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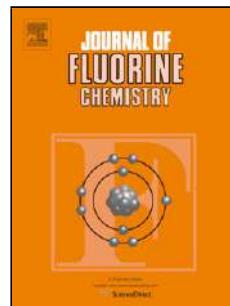
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Department of Chemistry

Journal Pre-proof

Synthesis, characterization, *in vitro* DNA photocleavage and cytotoxicity studies of
4-arylazo-1-phenyl-3-(2-thienyl)-5-hydroxy-5-trifluoromethylpyrazolines
and regioisomeric
4-arylazo-1-phenyl-5(3)-(2-thienyl)-3(5)-trifluoromethylpyrazoles



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Synthesis, Characterization, *in vitro* DNA Photocleavage and Cytotoxicity studies of 4-Arylazo-1-phenyl-3-(2-thienyl)-5-hydroxy-5-trifluoromethylpyrazolines and Regioisomeric 4-Arylazo-1-phenyl-5(3)-(2-thienyl)-3(5)-trifluoromethylpyrazoles

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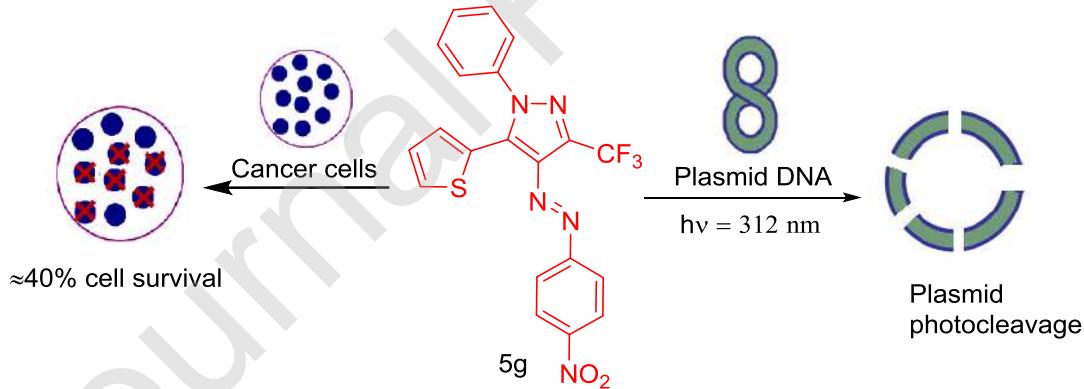
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Graphical Abstract:-



Highlights

- Synthesis of 4-arylazo-1-phenyl-3-(2-thienyl)-5-hydroxy-5-trifluoromethylpyrazolines and regioisomeric 4-arylazo-1-phenyl-5(3)-(2-thienyl)-3(5)-trifluoromethylpyrazoles was accomplished.
- All the synthesized compounds were screened for their DNA photocleavage on supercoiled pBr322 plasmoid.



Chitosan embedded with Ag/Au nanoparticles: investigation of their structural, optical and sensing properties

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Abstract

Quantitative detection of hydrogen peroxide (H_2O_2) is reported by utilizing an optical sensor based on the Surface Plasmon Resonances (SPR) of Ag and Au nanoparticles embedded in chitosan, a biopolymer. Ag and Au nanoparticles, fabricated by chemical reduction approach, were incorporated individually in chitosan matrix by solution casting method. Subsequently, their presence in the host matrix was confirmed using UV-visible spectroscopy, X-Ray diffractometer (XRD), High Resolution Transmission Electron Microscopy (HRTEM) and Field Emission Scanning Electron Microscopy (FESEM) along with Energy Dispersive Analysis of X-Ray (EDAX) spectroscopy. Structural changes induced in chitosan with addition of varying concentration of Ag or Au nanoparticles were studied using Fourier transform infrared (FTIR) spectroscopy. Optical energy gap of chitosan decreased from 3.82 ± 0.28 eV to 1.84 ± 0.19 eV for Ag-chitosan nanocomposite (Nc) film containing 0.50 wt% Ag nanoparticle while to a value of 2.14 ± 0.08 eV for Au-chitosan Nc film containing 0.5 wt% of Au nanoparticle. A significant difference in position and intensity of SPR absorption band was observed as a function of variable concentration of H_2O_2 . The detection limit of these optical sensors is upto 0.3 μM concentration of H_2O_2 .

Keywords Nanocomposite · Chitosan · Plasmon · Absorption · Sensor

Introduction

Chitosan is a linear cationic polysaccharide derived from the deacetylation of chitin, the second most abundant polysaccharide after cellulose [1]. Though like other biopolymers chitin is biodegradable, biocompatible and non-toxic but its applications and processing are limited due to its insolubility in

conventional solvents. Owing to this fact chitosan is derived by chemical/enzymatic deacetylation of chitin thus maintaining green properties of chitin [2].

Chitosan can be claimed as one of the best contenders for fabricating thin films owing to its excellent film-forming property. It contains free amino and hydroxyl groups which can be functionalized through binding with the cationic and anionic

Highlights

- Stable Ag-chitosan and Au-chitosan Nc films were fabricated.
- Optical energy gap reduces to 1.84 ± 0.19 eV and 2.14 ± 0.08 eV for Ag-chitosan Nc film and Au-chitosan Nc film respectively as compared to 3.82 ± 0.28 eV for chitosan.
- FTIR analysis confirms the strong interaction of Ag and Au nanoparticles with chitosan.
- The detection limit of these optical sensors is upto 0.3 μM concentration of H_2O_2 .

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forms of noble metals and can stabilize their nanoparticles [3]. Due to these unique physico-chemical properties chitosan is finding increasing applications in the fields of biotechnology, nanotechnology, food technology, agriculture, medicine, water purification and textiles [4].

Among different nanofillers metal nanoparticles are one of the most popular nanofillers as they exhibit size and shape dependent properties, resulting in their applications in optoelectronics, catalysis, coatings and sensors [5–7]. Nanoparticles of noble metals silver (Ag) and gold (Au) display peculiar characteristics like; surface plasmon resonance (SPR) in visible region, surface enhanced Raman scattering (SERS), outstanding plasmonic activity, catalytic activity etc. [8–10]. Such features of interest of Ag and Au nanoparticles make them potential candidates for diverse technological applications such as sensors [11], optics [12], filter technology [13], photovoltaic applications [14], etc. Therefore, incorporation of Ag or Au nanoparticles in chitosan matrix is a unique way to combine the properties of both noble metal nanoparticles as well as chitosan highlighting the need for comprehensive studies on synthesis and characterization of Ag-chitosan and Au-chitosan Ncs. There are several reports pertaining to antibacterial, antifungal as well as photoresponsive properties of Ag/Au-chitosan Ncs [15–18]. However, there is still a need for in-depth investigation of optical and sensing properties of Ag-chitosan and Au-chitosan Nc films in order to explore the full potential of applications of Ag-chitosan and Au-chitosan Nc films.

Sensors play an imperative role in analyzing the environment, providing information on industrial production processes, quality management of food products and numerous other applications. They provide valuable information by interacting with various chemical components. Hydrogen peroxide (H_2O_2) is one of the analytes that is routinely being used by the medical community. H_2O_2 is being used in several fields such as water treatment plants, fumigation as well as bleaching and cleaning microcircuits [19] however, it is hazardous to environment and living organisms. Even a little concentration of H_2O_2 can harm the cellular system. Hence, detection of H_2O_2 quantity in environment, food, pharmaceutical products and particularly in clinical laboratories is of utmost importance [20].

Thus, the development of new techniques for the quantitative detection of H_2O_2 is essential. Ning et al. [21] have utilized platinum, $ZnFe_2O_4$ functionalized reduced graphene oxide based electrode for determination of H_2O_2 . They observed high sensitivity and selectivity of electrode towards H_2O_2 . Zhang et al. [22] have employed Au/CeO₂-chitosan composite film as electrochemical biosensors for detection of H_2O_2 . However, these techniques suffer from several drawbacks such as use of highly sophisticated instruments and low selectivity/poor reproducibility. Therefore, other alternative methods are required for the determination of H_2O_2 . One of

the ideal candidates for the determination of H_2O_2 is SPR based optical sensor due to its high sensitivity and cost effective detection. SPR sensing is based on the principle of measurable shifts in the SPR wavelength due to its strong dependence on refractive index of surrounding medium. Noghabi et al. [23] investigated the colorimetric sensing of green synthesized Ag nanoparticles towards H_2O_2 . They observed a significant change in the intensity of SPR peak with increasing concentration of H_2O_2 . Mohan et al. [24] have also employed starch stabilized Ag nanoparticles for detection of H_2O_2 . However, fabrication of SPR based sensor for the detection of H_2O_2 by utilizing Ag-chitosan and Au-chitosan Nc films has seldom been reported.

The present research work reports the synthesis of Ag and Au nanoparticles via chemical reduction approach and fabrication of Ag-chitosan and Au-chitosan Nc films by solution casting technique. The structural and optical properties of synthesized Nc films have been investigated and their sensitivity towards H_2O_2 has been demonstrated.

Experimental section

Reagents and materials

Silver nitrate ($AgNO_3$) (Mol. wt. = 169.87 g mol⁻¹), glycerol (Mol. wt. = 92.10 g mol⁻¹) and tri-sodium citrate dihydrate (Mol. wt. = 294.10 g mol⁻¹) were procured from Rankem. Soluble starch (Mol. wt. = 342.30 g mol⁻¹), 36% H_2O_2 (Mol. wt. = 34.01 g mol⁻¹) and acetic acid (Mol. wt. = 60 g mol⁻¹) were bought from HIMEDIA. Chloauric acid ($HAuCl_4 \cdot xH_2O$) was purchased from Molychem. Chitosan (viscosity 47.78 cp) with degree of deacetylation 96.80% was procured from Central Institute of Fisheries (CIF) Kochi, India. Analytical grade chemicals were used as received and deionized water was used to prepare all solutions.

Methods

Preparation of Ag nanoparticles

For synthesizing Ag nanoparticles, 0.1 g of soluble starch was dissolved in 25 ml of distilled water at 85 °C. Subsequently, 10 ml of freshly prepared 0.04 M $AgNO_3$ was added to 25 ml of hot aqueous solution of soluble starch under vigorous stirring under dark. Then 15 ml of 0.12 M fructose solution was added to the reactive system, which was held at 85 °C under vigorous stirring for half an hour when colour of the mixture changed to light yellow indicating the formation of Ag nanoparticles. Colour of the mixture continued to darken and finally turned brown after 1 h [10, 25, 26].

25. Chevron P, Gouanvé F, Espuche E (2014) Green synthesis of colloid silver nanoparticles and resulting biodegradable starch/silver nanocomposites. *Carbohydr Polym* 108:291–298
26. Meena, Sharma A (2017). *Integr Ferroelectr* 184:158–165
27. Carlo GD, Curulli A, Toro RG, Bianchini C, Caro TD, Padeletti G, Zane D, Ingo GM (2012) Green Synthesis of Gold–Chitosan Nanocomposites for Caffeic Acid Sensing. *Langmuir* 28:5471–5479
28. Govindan S, Nivetha EAK, Saravanan R, Narayanan V, Stephen A (2012) Synthesis and characterization of chitosan–silver nanocomposite. *Appl Nanosci* 2:299–303
29. JCPDS-ICDD (Joint Committee on Powder Diffraction Standard–International Centre for Diffraction Data (2003). Silver file no. 67-0720
30. Kittel C (2005) Introduction to solid state physics. Wiley, USA
31. Cullity BD (1978) Elements of X-ray diffraction, Addison Wesley Pub.Co.
32. Youssef AM, Yousef HA, El-Sayed SM, Kamel S (2015) Mechanical and antibacterial properties of novel high performance chitosan/nanocomposite films. *Int J Biol Macromol* 76:25–32
33. Archana D, Singh BK, Dutta J, Dutta PK (2015) Chitosan-PVP-nano silver oxide wound dressing: In vitro and in vivo evaluation. *Int J Biol Macromol* 73:49–57
34. Metzler M, Chylińska M, Kaczmarek H (2015) Preparation and characteristics of nanosilver composite based on chitosan-graft-acrylic acid copolymer. *J Polym Res* 22:146
35. Caldera-Villalobos M, Serrano JG, Cid AAP, Herrera-Gonzalez AM (2017) Polyelectrolytes with sulfonate groups obtained by chemical modification of chitosan useful in green synthesis of Au and Ag nanoparticles. *J Appl Polym Sci* 134:45240
36. Venkateshan J, Lee JY, Kang DS, Anil S, Kim SK, Shim MS, Kim DG (2017) Antimicrobial and anticancer activities of porous chitosan-alginate biosynthesized silver nanoparticles. *Int J Biol Macromol* 98:515–525
37. Kizil R, Irudayaraj J, Seetharaman K (2002) Characterization of Irradiated Starches by Using FT-Raman and FTIR Spectroscopy. *J Agric Food Chem* 50:3912–3918
38. Futyra AR, Liskiewicz MK, Sebastian V, Irusta S, Arruebo M, Kyziola A, Stochel G (2017) Development of noncytotoxic silver–chitosan nanocomposites for efficient control of biofilm forming microbes. *RSC Adv* 7:52398–52413
39. Hussain ST, Iqbal M, Mazhar M (2009) Size control synthesis of starch capped-gold nanoparticles. *J Nanopart Res* 11:1383–1391
40. Badeggi UM, Ismail E, Adeloye AO, Botha S, Badmus JA, Marnewick JL, Cupido CN, Hussein AA (2020). *Biomolecules* 10:452
41. Seo DK, Homann R (1999) Direct and indirect band gap types in one-dimensional conjugated or stacked organic materials. *Theor Chem Accounts* 102:23–32
42. Hassanien AS, AKL AA (2016) Effect of Se addition on optical and electrical properties of chalcogenide CdSSe thin films. *Superlattice Microst* 89:153–169
43. Davis E, Mott NF (1970) Electronic processes in non-crystalline materials. Oxford University Press Inc., USA
44. Sonal, Sharma A, Aggarwal S (2018). *Opt Mater* 84:807

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Research paper

Tail approach synthesis of novel benzenesulfonamides incorporating 1,3,4-oxadiazole hybrids as potent inhibitor of carbonic anhydrase I, II, IX, and XII isoenzymes

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In silico enhancement of azo dye adsorption affinity for cellulose fibre through mechanistic interpretation under guidance of QSPR models using Monte Carlo method with index of ideality correlation

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ABSTRACT

Azo dyes are a group of chemical moieties joined by azo (-N=N-) group with potential usefulness in different industrial applications. But these dyes are not devoid of hazardous consequence because of poor affinity for the fibre and discharge into the water stream. The chemical aspects of 72 azo dyes towards cellulose fibre in terms of their affinity by QSPR have been explored in the present work. We have employed two approaches, namely balance of correlation without IIC (TF_1) and balance of correlation with IIC (TF_2), to generate 16 QSAR models from 8 splits. The determination coefficient of calibration and validation set was found higher when the QSPR models were developed using the index of ideality correlation (IIC) parameter (TF_2). The model developed with TF_2 for split 3 was considered as a prominent model because the determination coefficient of the validation set was maximum ($r^2 = 0.9468$). The applicability domain (AD) was also analysed based on 'statistical defect', d (A) for a SMILES attribute. The mechanistic interpretation was done by identifying the SMILES attributes responsible for the promoter of endpoint increase and promoter of endpoint decrease. These SMILES attributes were applied to design 15 new dyes with higher affinity for cellulose fibre.

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KEYWORDS

QSPR; azo dyes affinity; IIC; applicability domain (AD); OECD

Introduction

The use of colouring is a very sophisticated method used to change the colour features of various substrates, including cloth, paper and leather. Substances with dyeing abilities were derived from natural substances, mostly from animals or plants, before even the mid-nineteenth century [1]. But by the advent of the twentieth century, natural dyes were rendered almost obsolete by synthetic dyes. Today, nearly all commercially produced dyes and pigments are synthetic compounds, with a few inorganic pigments [2,3]. Dyes are used in almost all types of products in the market such as textiles, paper, food, packaging, plastics, lasers, biometrics, solar capture, diagnostic tools, cosmetics and household goods [4–6].

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The whole textile industry is highly dependent on the dyes, and approximately 700,000 tons of more than 100,000 dyes are produced annually for commercial purpose [7,8]. These display an enormous range of dazzling shades and good wet fastness. However, dye loss, which is mainly caused by relatively poor dye fibre fixation and the presence of unreactive hydrolysed dye, is a matter of concern as it leads to the release of dyes into the environment. Pollution of the environment resulting from the release of industrial effluents has attracted the attention of the scientific community worldwide [9]. Effluents from industries such as textiles, leather, paper and plastic contain different types of artificial dyestuffs. The groundwater quality is adversely affected by the haphazard discharge of these dyes into sewage, ponds and rivers. It has been proposed that nearly 10–15% of the total dye production is discharged into the environment through wastewater, which poses serious hazards to the flora and fauna [8]. Literature survey reveals that a few dyes can be degraded by microorganisms under anaerobic condition [10]. So, in most of cases, these dyes lead to the production of carcinogenic compounds [10,11].

Azo dyes are chemicals identified by the existence of one or more azo units ($-N=N-$), generally, in numbers one or four, connected to phenyl and naphthyl radicals, which are typically substituted by certain pairings of functional groups namely: amino ($-NH_2$), chlorine ($-Cl$), hydroxyl ($-OH$), methyl ($-CH_3$), nitro ($-NO_2$), sulphonic acid and sodium salts ($-SO_3Na$) [12–16]. Azo dyes constitute the biggest group of colours, as around 70% of all organic dyes products are azo dyes [17]. These are extensively consumed by the dyeing industry and are present in almost all types of dyes, such as direct, reactive, and disperse dyes [18–20]. Notwithstanding the challenge in the processing of the residues produced and the detrimental implications for their use, azo dyes, particularly sulphuric ones, are extensively employed for dyeing fabrics. This is partly because of low costs and good affinity features [21]. Levels of dye fixation can be measured in terms of the dye-fibre attractive force commonly known as ‘affinity’, and the design of high-affinity dyes is the need of the hour [22].

The fibre of cellulose is an incredible natural resource that has extensive uses in numerous industrial products, and particularly in textiles. In this, two glucose moieties are interconnected through a glycosidic linkage and polymer components form a hydrogen bond set-up with hydroxyl groups and ethereal oxygen. Cellulose cannot fixate ionic dyes due to a lack of ionic sites in its structure [23]. Therefore dye molecules with some special structural characteristics are required for direct dying of the cotton. Several factors such as the formation of hydrogen bonds, electrostatic fields, hydrophobicity, etc., influence the affinity between cotton and dye molecule, and therefore diverse experimental records exist in the literature [6,23–30]. For that reason, various computational approaches such as quantitative structure–property/activity relationships (QSPR/QSAR) have been extensively used to study this affinity [23,25–31].

QSAR modelling is a statistical approach correlating the structural information of chemicals with endpoints/response values (activity/property/toxicity) using chemometric techniques [32,33]. CORAL (<http://www.insilico.eu/coral/>) software is commonly used for the building of QSAR models in accordance with OECD principles [34,35]. It uses the SMILES notations of the molecules in the input file and extracts the best model using Monte Carlo optimization [36–38]. Many endpoints, including pharmacological and toxicological, have been modelled by this prestigious tool [39–45].



interpretation was done by identifying the SMILES attributes responsible for the promoter of endpoint increase and promoter of endpoint decrease. These SMILES attributes were applied to design 15 new dyes with higher affinity for cellulose fibre. Hence, it can be summarized that the present QSPR method enhances the azo dye adsorption affinity for cellulose fibre through mechanistic interpretation under the guidance of the QSPR model using the Monte Carlo method with index of ideality correlation.

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Author contributions

Authors have done equivalent contributions to this work.

Disclosure statement

No potential conflict of interest was reported by the authors.

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Data availability

The processed data required to reproduce these findings are available to download from supporting information.

References

- [1] L. Nambela, L.V. Haule, and Q. Mgani, *A review on source, chemistry, green synthesis and application of textile colorants*, J. Clean. Prod. 246 (2020), pp. 119036. <https://doi.org/10.1016/j.jclepro.2019.119036>.
- [2] N. Sekar and D.S. Deulgaonkar, *Azo styryl dyes: A brief review*, Colourage 51 (2004), pp. 71–79.
- [3] R. Mukherjee, R. Kumar, A. Sinha, Y. Lama, and A.K. Saha, *A review on synthesis, characterization, and applications of nano zero valent iron (nZVI) for environmental remediation*, Crit. Rev. Environ. Sci. Technol. 46 (2015), pp. 443–466. <https://doi.org/10.1080/10643389.2015.1103832>.
- [4] S. Venkata Mohan, N. Chandrasekhar Rao, K. Krishna Prasad, and J. Karthikeyan, *Treatment of simulated reactive Yellow 22 (azo) dye effluents using Spirogyra species*, Wast. Manag. 22 (2002), pp. 575–582. [https://doi.org/10.1016/s0956-053x\(02\)00030-2](https://doi.org/10.1016/s0956-053x(02)00030-2).
- [5] E. Forgacs, T. Cserhati, and G. Oros, *Removal of synthetic dyes from wastewaters: A review*, Environ. Int. 30 (2004), pp. 953–971. <https://doi.org/10.1016/j.envint.2004.02.001>.
- [6] A.B. Dos Santos, F.J. Cervantes, and J.B. van Lier, *Review paper on current technologies for decolourisation of textile wastewaters: Perspectives for anaerobic biotechnology*, Bioresour. Technol. 98 (2007), pp. 2369–2385. <https://doi.org/10.1016/j.biortech.2006.11.013>.

- descriptors, *J. Chem. Inf. Comput. Sci.* 43 (2003), pp. 1502–1512. <https://doi.org/10.1021/ci034064f>.
- [24] A. Welham, *The theory of dyeing (and the secret of life)*, *J. Soc. Dye. Colour.* 116 (2000), pp. 140–143.
- [25] S. Yu, Q. Zhou, X. Zhang, S. Jia, Y. Gan, Y. Zhang, J. Shi, and J. Yuan, *Hologram quantitative structure–activity relationship and topomer comparative molecular-field analysis to predict the affinities of azo dyes for cellulose fibers*, *Dyes Pigm.* 153 (2018), pp. 35–43. <https://doi.org/10.1016/j.dyepig.2018.01.053>.
- [26] S. Funar-Timofei, W.M.F. Fabian, L. Kurunczi, M. Goodarzi, S.T. Ali, and Y.V. Heyden, *Modelling heterocyclic azo dye affinities for cellulose fibres by computational approaches*, *Dyes Pigm.* 94 (2012), pp. 278–289. <https://doi.org/10.1016/j.dyepig.2012.01.015>.
- [27] S. Timofei, L. Kurunczi, and Z. Simon, *Structure-affinity relationships by the MTD method for binding to cellulose fibre of some heterocyclic monoazo dyes*, *Match* 44 (2001), pp. 349–360.
- [28] S. Timofei and W.M.F. Fabian, *Comparative molecular field analysis of heterocyclic monoazo dye–fiber affinities*, *J. Chem. Inf. Comput. Sci.* 38 (1998), pp. 1218–1222. <https://doi.org/10.1021/ci9704367>.
- [29] S. Timofei, L. Kurunczi, T. Suzuki, W.M.F. Fabian, and S. Mureşan, *Multiple linear regression (mlr) and neural network (nn) calculations of some disazo dye adsorption on cellulose*, *Dyes Pigm.* 34 (1997), pp. 181–193. [https://doi.org/10.1016/s0143-7208\(96\)00081-2](https://doi.org/10.1016/s0143-7208(96)00081-2).
- [30] G. Seu and L. Mura, *Adsorption of heterocyclic disperse azo dyes by cotton*, *Am. Dyest. Rep.* 73 (1984), pp. 43–44.
- [31] X. Wang, Y. Sun, L. Wu, S. Gu, R. Liu, L. Liu, X. Liu, and J. Xu, *Quantitative structure–affinity relationship study of azo dyes for cellulose fibers by multiple linear regression and artificial neural network*, *Chemom. Intel. Lab. Syst.* 134 (2014), pp. 1–9. <https://doi.org/10.1016/j.chemolab.2014.03.001>.
- [32] R.B. Aher, K. Khan, and K. Roy, *A brief introduction to quantitative structure–activity relationships as useful tools in predictive ecotoxicology*, in *Ecotoxicological QSARs*, K. Roy, ed., Springer US, New York, NY, 2020, pp. 27–53.
- [33] A.A. Toropov, A.P. Toropova, A. Roncaglioni, and E. Benfenati, *Prediction of biochemical endpoints by the coral software: Prejudices, paradoxes, and results*, in *Methods in Molecular Biology*, Humana Press Inc, New York, NY, 2018, pp. 573–583.
- [34] A.P. Toropova, A.A. Toropov, E. Benfenati, R. Rallo, D. Leszczynska, and J. Leszczynski, *Development of monte carlo approaches in support of environmental research*, in *Advances in QSAR Modeling: Applications in Pharmaceutical, Chemical, Food, Agricultural and Environmental Sciences*, K. Roy ed., Springer International Publishing, Cham, 2017, pp. 453–469.
- [35] P. Kumar and A. Kumar, *Monte Carlo method based QSAR studies of mer kinase inhibitors in compliance with OECD principles*, *Drug Res. (Stuttg.)* 68 (2018), pp. 189–195. <https://doi.org/10.1055/s-0043-119288>.
- [36] P. Kumar, A. Kumar, and J. Sindhu, *In silico design of diacylglycerol acyltransferase-1 (DGAT1) inhibitors based on SMILES descriptors using Monte-Carlo method*, *SAR QSAR Environ. Res.* 30 (2019), pp. 525–541. <https://doi.org/10.1080/1062936X.2019.1629998>.
- [37] A.P. Toropova, A.A. Toropov, E. Carnesecchi, E. Benfenati, and J.L. Dorne, *The index of ideality of correlation: Models for flammability of binary liquid mixtures*, *Chem. Pap.* 74 (2019), pp. 601–609. <https://doi.org/10.1007/s11696-019-00903-w>.
- [38] M. Marzo, G.J. Lavado, F. Como, A.P. Toropova, A.A. Toropov, D. Baderna, C. Cappelli, A. Lombardo, C. Toma, M. Blazquez, and E. Benfenati, *QSAR models for biocides: The example of the prediction of Daphnia magna acute toxicity*, *SAR QSAR Environ. Res.* 31 (2020), pp. 227–243. <https://doi.org/10.1080/1062936X.2019.1709221>.
- [39] E. Carnesecchi, A.A. Toropov, A.P. Toropova, N. Kramer, C. Svendsen, J.L. Dorne, and E. Benfenati, *Predicting acute contact toxicity of organic binary mixtures in honey bees (A. mellifera) through innovative QSAR models*, *Sci. Total Environ.* 704 (2020), pp. 135302. <https://doi.org/10.1016/j.scitotenv.2019.135302>.

- [40] A.A. Toropov, A.P. Toropova, G. Raitano, and E. Benfenati, *CORAL: Building up QSAR models for the chromosome aberration test*, Saudi J. Biol. Sci. 26 (2019), pp. 1101–1106. <https://doi.org/10.1016/j.sjbs.2018.05.013>.
- [41] P. Kumar, A. Kumar, and J. Sindhu, *Design and development of novel focal adhesion kinase (FAK) inhibitors using Monte Carlo method with index of ideality of correlation to validate QSAR*, SAR QSAR Environ. Res. 30 (2019), pp. 63–80. <https://doi.org/10.1080/1062936X.2018.1564067>.
- [42] M. Duhan, R. Singh, M. Devi, J. Sindhu, R. Bhatia, A. Kumar, and P. Kumar, *Synthesis, molecular docking and QSAR study of thiazole clubbed pyrazole hybrid as alpha-amylase inhibitor*, J. Biomol. Struct. Dyn. (2019), pp. 1–17. <https://doi.org/10.1080/07391102.2019.1704885>.
- [43] J.S. Choi, T.X. Trinh, T.H. Yoon, J. Kim, and H.G. Byun, *Quasi-QSAR for predicting the cell viability of human lung and skin cells exposed to different metal oxide nanomaterials*, Chemosphere 217 (2019), pp. 243–249. <https://doi.org/10.1016/j.chemosphere.2018.11.014>.
- [44] A.P. Toropova and A.A. Toropov, *CORAL: Monte Carlo method to predict endpoints for medical chemistry*, Mini-Rev. Med. Chem. 18 (2018), pp. 382–391. <https://doi.org/10.2174/1389557517666170927154931>.
- [45] A.P. Toropova and A.A. Toropov, *Use of the index of ideality of correlation to improve models of eco-toxicity*, Environ. Sci. Pollut. Res. Intenat. 25 (2018), pp. 31771–31775. <https://doi.org/10.1007/s11356-018-3291-5>.
- [46] S. Manisha, P.K. Chauhan, and A. Kumar, *Development of prediction model for fructose-1,6- bisphosphatase inhibitors using the Monte Carlo method*, SAR QSAR Environ. Res. 30 (2019), pp. 145–159. <https://doi.org/10.1080/1062936X.2019.1568299>.
- [47] P. Kumar and A. Kumar, *CORAL: QSAR models of CB1 cannabinoid receptor inhibitors based on local and global SMILES attributes with the index of ideality of correlation and the correlation contradiction index*, Chemom. Intel. Lab. Syst. 200 (2020), pp. 103982. <https://doi.org/10.1016/j.chemolab.2020.103982>.
- [48] A. Kumar, K.B. Manisha, and P. Kumar, *Use of graph based descriptors for determination of structural features causing modulation of fructose-1,6-bisphosphatase*, Drug Res. (Stuttg.) 70 (2020), pp. 226–232. <https://doi.org/10.1055/a-1138-8725>.
- [49] Marvin-Sketch-v.14.11.17.0, ChemAxon, XchemAxon KFT, Budapest, Hungary.
- [50] J.B. Veselinovic, G.M. Nikolic, N.V. Trutic, J.V. Zivkovic, and A.M. Veselinovic, *Monte Carlo QSAR models for predicting organophosphate inhibition of acetylcholinesterase*, SAR QSAR Environ. Res. 26 (2015), pp. 449–460. <https://doi.org/10.1080/1062936X.2015.1049665>.
- [51] A. Gissi, A.A. Toropov, A.P. Toropova, O. Nicolotti, A. Carotti, and E. Benfenati, *Building up QSAR model for toxicity of psychotropic drugs by the Monte Carlo method*, Struct. Chem. 25 (2013), pp. 1067–1073. <https://doi.org/10.1007/s11224-013-0380-4>.
- [52] A.A. Toropov, I. Raska Jr., A.P. Toropova, M. Raskova, A.M. Veselinovic, and J.B. Veselinovic, *The study of the index of ideality of correlation as a new criterion of predictive potential of QSPR/QSAR-models*, Sci. Total Environ. 659 (2019), pp. 1387–1394. <https://doi.org/10.1016/j.scitotenv.2018.12.439>.
- [53] S. Ahmadi, F. Mardinia, N. Azimi, M. Qomi, and E. Balali, *Prediction of chalcone derivative cytotoxicity activity against MCF-7 human breast cancer cell by Monte Carlo method*, J. Mol. Struct. 1181 (2019), pp. 305–311. <https://doi.org/10.1016/j.molstruc.2018.12.089>.
- [54] P.G. Achary, *Simplified molecular input line entry system-based optimal descriptors: QSAR modelling for voltage-gated potassium channel subunit Kv7.2*, SAR QSAR Environ. Res. 25 (2014), pp. 73–90. <https://doi.org/10.1080/1062936X.2013.842930>.
- [55] S. Bhargava, N. Adhikari, S.A. Amin, K. Das, S. Gayen, and T. Jha, *Hydroxyethylamine derivatives as HIV-1 protease inhibitors: A predictive QSAR modelling study based on Monte Carlo optimization*, SAR QSAR Environ. Res. 28 (2017), pp. 973–990. <https://doi.org/10.1080/1062936X.2017.1388281>.
- [56] A. Kumar, J. Sindhu, and P. Kumar, *In-silico identification of fingerprint of pyrazolyl sulfonamide responsible for inhibition of N-myristoyltransferase using Monte Carlo method with index of ideality of correlation*, J. Biomol. Struct. Dyn. (2020), pp. 1–12. <https://doi.org/10.1080/07391102.2020.1784286>.

- [57] A.A. Toropov and A.P. Toropova, *Optimal descriptor as a translator of eclectic data into end-point prediction: Mutagenicity of fullerene as a mathematical function of conditions*, Chemosphere 104 (2014), pp. 262–264. <https://doi.org/10.1016/j.chemosphere.2013.10.079>.
- [58] A.P. Toropova, A.A. Toropov, E. Carnesecchi, E. Benfenati, and J.L. Dorne, *The using of the Index of Ideality of Correlation (IIC) to improve predictive potential of models of water solubility for pesticides*, Environ. Sci. Pollut. Res. Internat. 27 (2020), pp. 13339–13347. <https://doi.org/10.1007/s11356-020-07820-6>.
- [59] M. Nimbhal, K. Bagri, P. Kumar, and A. Kumar, *The index of ideality of correlation: A statistical yardstick for better QSAR modeling of glucokinase activators*, Struct. Chem. 31 (2019), pp. 831–839. <https://doi.org/10.1007/s11224-019-01468-w>.
- [60] A. Kumar and P. Kumar, *Construction of pioneering quantitative structure activity relationship screening models for abuse potential of designer drugs using index of ideality of correlation in Monte Carlo optimization*, Arch. Toxicol. (2020). <https://doi.org/10.1007/s00204-020-02828-w>.
- [61] P. Kumar and A. Kumar, *Nucleobase sequence based building up of reliable QSAR models with the index of ideality correlation using Monte Carlo method*, J. Biomol. Struct. Dyn. 38 (2020), pp. 3296–3306. <https://doi.org/10.1080/07391102.2019.1656109>.
- [62] A. Golbraikh and A. Tropsha, *Beware of $q^2!$* , J. Mol. Graph Mod. 20 (2002), pp. 269–276. [https://doi.org/10.1016/s1093-3263\(01\)00123-1](https://doi.org/10.1016/s1093-3263(01)00123-1).
- [63] K. Roy, R.N. Das, P. Ambure, and R.B. Aher, *Be aware of error measures. Further studies on validation of predictive QSAR models*, Chemom. Intel. Lab. Syst. 152 (2016), pp. 18–33. <https://doi.org/10.1016/j.chemolab.2016.01.008>.
- [64] N. Chirico and P. Gramatica, *Real external predictivity of QSAR models: How to evaluate it? Comparison of different validation criteria and proposal of using the concordance correlation coefficient*, J. Chem. Inf. Model. 51 (2011), pp. 2320–2335. <https://doi.org/10.1021/ci200211n>.
- [65] A.P. Toropova and A.A. Toropov, *Does the index of ideality of correlation detect the better model correctly?* Mol. Inform. 38 (2019), pp. e1800157. <https://doi.org/10.1002/minf.201800157>.
- [66] C. Rucker, G. Rucker, and M. Meringer, *γ -Randomization and its variants in QSPR/QSAR*, J. Chem. Inf. Model. 47 (2007), pp. 2345–2357. <https://doi.org/10.1021/ci700157b>.
- [67] I. Mitra, A. Saha, and K. Roy, *Exploring quantitative structure–activity relationship studies of antioxidant phenolic compounds obtained from traditional Chinese medicinal plants*, Molecul. Simulat. 36 (2010), pp. 1067–1079. <https://doi.org/10.1080/08927022.2010.503326>.
- [68] N.I. Zhokhova, I.I. Baskin, V.A. Palyulin, A.N. Zefirov, and N.S. Zefirov, *A study of the affinity of dyes for cellulose fiber within the framework of a fragment approach in QSPR*, Russ. J. Appl. Chem. 78 (2005), pp. 1013–1017. <https://doi.org/10.1007/s11167-005-0439-0>.
- [69] J. Polanski, R. Gieleciak, and M. Wyszomirski, *Comparative molecular surface analysis (CoMSA) for modeling dye-fiber affinities of the azo and anthraquinone dyes*, J. Chem. Inf. Comput. Sci. 43 (2003), pp. 1754–1762. <https://doi.org/10.1021/ci0340761>.



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In-silico identification of fingerprint of pyrazolyl sulfonamide responsible for inhibition of N-myristoyltransferase using Monte Carlo method with index of ideality of correlation

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ABSTRACT

Human African trypanosomiasis (HAT) or sleeping sickness like infections remain a serious health concern around the globe due to unavailability of safe and potential drugs for their treatment. Moreover, developing safe, potential and highly specific target based treatments is still a challenge for present drug discovery programs. A series of pyrazole based sulfonamides are identified as an inhibitor of *Trypanosoma brucei* N-myristoyltransferase (TbNMT). In the present manuscript, we have developed robust and reliable QSAR models by using the balance of correlation method in CORAL software. The chemical structures are represented by simplified molecular input line entry system (SMILES). The significance of the index of ideality correlation (IIC) with applicability domain (AD) is also studied at depth. The models developed by considering the index of ideality of correlation (IIC) were found to statistically more significant and robust. One QSAR model with best $R^2_{calibration} = 0.8638$ for split 2 was considered as the leading model. A greater value of cR^2 i.e. 0.5 for all models in Y-randomization test showed the robustness of developed models. The outliers and promoters of increase and decrease of endpoint were also extracted independently from the leading models. The mechanistic interpretation of developed models explains the role of different structural attributes in predicting the pIC_{50} of pyrazole sulfonamides extracted from the crystal structure of Leishmania major N-myristoyltransferase (NMT) along with co-crystallized myristoyl-CoA and ligands NMT106, NMT157, NMT187 and NMT236 (PDB ID: 4A2Z, 4A30, 4A32, 2WSA).

Abbreviations: HAT: Human African trypanosomiasis; IIC: Index of Ideality of Correlation; TbNMT: *Trypanosoma brucei* N-myristoyltransferase; CW: Correlation weight; OECD: Organization of Economic Co-operation and Development; QSAR: Quantitative Structure Activity Relationship; CORAL: CORrelation And Logic; AD: Applicability Domain

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Introduction

Human African trypanosomiasis (HAT) or sleeping sickness is caused by the kinetoplastid parasite *Trypanosoma brucei* and it is transmitted by the bite of an infected tsetse fly (*Glossina* genus) (Nagle et al., 2014; Njoroge et al., 2014). Mainly two species i.e. *Trypanosoma brucei gambiense* and *Trypanosoma brucei rhodesiense* of the above protozoan parasite is responsible for the HAT disease which leads to death if not treated properly or treatment is delayed (Klug et al., 2020; Lepovitz et al., 2020; Scarim et al., 2020). The clinical features are characterized by lymphadenopathy, fever and excessive sleepiness due to encephalopathy or encephalitis (Macleod et al., 2020; Singh et al., 2019; Watson et al., 2019). This disease is progressed by two distinct stages: the hemolymphatic stage and central nervous system (CNS) involvement or meningoencephalitic stage. The first stage is initially an acute

stage, starts with the proliferation of parasite in the hemolymphatic system and give rise to non-specific symptoms (Bayliss et al., 2017; Harrison et al., 2018; Klug et al., 2020; Montalvo-Quiros et al., 2015; Patrick et al., 2017). However, the second stage onsets with the migration of parasite from the hemolymphatic system to CNS and cause classical symptoms of sleeping sickness, ultimately leading to coma and death. In the present scenario, the drugs available for HAT treatment are unsafe and frequently allied with severe or life-threatening side effects such as fatal encephalopathy, agranulocytosis, drug-resistance, and myocardial damage. Mainly five drugs (suramin, pentamidine, melarsoprol, eflornithine and nifurtimox) are used to treat various stages of HAT disease (Hagen et al., 2020). For 1st stage treatment, suramin and pentamidine are used, whereas melarsoprol and eflornithine are recommended for stage 2 infection. Recently, the nifurtimox-eflornithine combination therapy (NECT) has

- Kumar, A., & Chauhan, S. (2017). Monte Carlo method based QSAR modelling of natural lipase inhibitors using hybrid optimal descriptors. *SAR and QSAR in Environmental Research*, 28(3), 179–197. <https://doi.org/10.1080/1062936X.2017.1293729>
- Kumar, P., Duhan, M., Sindhu, J., Kadyan, K., Saini, S., & Panigar, N. (2020). Thiazolidine-4-one clubbed pyrazoles hybrids: Potent α -amylase and α -glucosidase inhibitors with NLO properties. *Journal of Heterocyclic Chemistry*, 57(4), 1573–1587. <https://doi.org/10.1002/jhet.3882>
- Kumar, P., & Kumar, A. (2018). Monte Carlo method based QSAR studies of Mer Kinase inhibitors in compliance with OECD principles. *Drug Research*, 68(4), 189–195. <https://doi.org/10.1055/s-0043-119288>
- Kumar, P., & Kumar, A. (2019). Nucleobase sequence based building up of reliable QSAR models with the index of ideality correlation using Monte Carlo method. *Journal of Biomolecular Structure and Dynamics*, online published. <https://doi.org/10.1080/07391102.2019.1656109>
- Kumar, P., & Kumar, A. (2020). CORAL: QSAR models of CB1 cannabinoid receptor inhibitors based on local and global SMILES attributes with the index of ideality of correlation and the correlation contradiction index. *Chemometrics and Intelligent Laboratory Systems*, 200 103982. <https://doi.org/10.1016/j.chemolab.2020...>
- Kumar, P., Kumar, A., & Sindhu, J. (2019a). Design and development of novel focal adhesion kinase (FAK) inhibitors using Monte Carlo method with index of ideality of correlation to validate QSAR. *SAR and QSAR in Environmental Research*, 30(2), 63–80. <https://doi.org/10.1080/1062936X.2018.1564067>
- Kumar, P., Kumar, A., & Sindhu, J. (2019b). In silico design of diacylglycerol acyltransferase-1 (DGAT1) inhibitors based on SMILES descriptors using Monte-Carlo method. *SAR and QSAR in Environmental Research*, 30(8), 525–541. <https://doi.org/10.1080/1062936X.2019.1629998>
- Kumar, P., Kumar, A., Sindhu, J., & Lal, S. (2019). QSAR Models for nitrogen containing monophosphonate and bisphosphonate derivatives as human farnesyl pyrophosphate synthase inhibitors based on Monte Carlo method. *Drug Research*, 69(3), 159–167. <https://doi.org/10.1055/a-0652-5290>
- Lepovitz, L. T., Meis, A. R., Thomas, S. M., Wiedeman, J., Pham, A., Mensa-Wilmot, K., & Martin, S. F. (2020). Design, synthesis, and evaluation of novel anti-trypanosomal compounds. *Tetrahedron*, 76(16), 131086. <https://doi.org/10.1016/j.tet.2020.131086>
- Macleod, O. J. S., Bart, J.-M., MacGregor, P., Peacock, L., Savill, N. J., Hester, S., Ravel, S., Sunter, J. D., Trevor, C., Rust, S., Vaughan, T. J., Minter, R., Mohammed, S., Gibson, W., Taylor, M. C., Higgins, M. K., & Carrington, M. (2020). A receptor for the complement regulator factor H increases transmission of trypanosomes to tsetse flies. *Nature Communications*, 11(1), 1326. <https://doi.org/10.1038/s41467-020-15125-y>
- Manisha, Chauhan, S., Kumar, P., & Kumar, A. (2019). Development of prediction model for fructose-1,6- bisphosphatase inhibitors using the Monte Carlo method. *SAR and QSAR in Environmental Research*, 30(3), 145–159. <https://doi.org/10.1080/1062936X.2019.1568299>
- Masand, V. H., El-Sayed, N. N. E., Bambole, M. U., Patil, V. R., & Thakur, S. D. (2019). Multiple quantitative structure-activity relationships (QSARs) analysis for orally active trypanocidal N-myristoyltransferase inhibitors. *Journal of Molecular Structure*, 1175, 481–487. <https://doi.org/10.1016/j.molstruc.2018.07.080>
- Montalvo-Quiros, S., Taladriz-Sender, A., Kaiser, M., & Dardonville, C. (2015). Antiprotozoal activity and DNA binding of dicationic acridones. *Journal of Medicinal Chemistry*, 58(4), 1940–1949. <https://doi.org/10.1021/jm5018303>
- Nagle, A. S., Khare, S., Kumar, A. B., Supek, F., Buchynskyy, A., Mathison, C. J. N., Chennamaneni, N. K., Pendem, N., Buckner, F. S., Gelb, M. H., & Molteni, V. (2014). Recent developments in drug discovery for leishmaniasis and human African trypanosomiasis. *Chemical Reviews*, 114(22), 11305–11347. <https://doi.org/10.1021/cr500365f>
- Nimbal, M., Bagri, K., Kumar, P., & Kumar, A. (2020). The index of ideality of correlation: A statistical yardstick for better QSAR modeling of glucokinase activators. *Structural Chemistry*, 31(2), 831–839. <https://doi.org/10.1007/s11224-019-01468-w>
- Njoroge, M., Njuguna, N. M., Mutai, P., Ongarora, D. S., Smith, P. W., & Chibale, K. (2014). Recent approaches to chemical discovery and development against malaria and the neglected tropical diseases human African trypanosomiasis and schistosomiasis. *Chemical Reviews*, 114(22), 11138–11163. <https://doi.org/10.1021/cr500098f>
- Ojha, P. K., Mitra, I., Das, R. N., & Roy, K. (2011). Further exploring rm2 metrics for validation of QSPR models. *Chemometrics and Intelligent Laboratory Systems*, 107(1), 194–205. <https://doi.org/10.1016/j.chemolab.2011.03.011>
- Patrick, D. A., Gillespie, J. R., McQueen, J., Hulverson, M. A., Ranade, R. M., Creason, S. A., Herbst, Z. M., Gelb, M. H., Buckner, F. S., & Tidwell, R. R. (2017). Urea Derivatives of 2-Aryl-benzothiazol-5-amines: A new class of potential drugs for human African trypanosomiasis. *Journal of Medicinal Chemistry*, 60(3), 957–971. <https://doi.org/10.1021/acs.jmedchem.6b01163>
- Pratim Roy, P., Paul, S., Mitra, I., & Roy, K. (2009). On two novel parameters for validation of predictive QSAR models. *Molecules (Basel, Switzerland)*, 14(5), 1660–1701. <https://doi.org/10.3390/molecules14051660>
- Roy, K., Chakraborty, P., Mitra, I., Ojha, P. K., Kar, S., & Das, R. N. (2013). Some case studies on application of "r(m)²" metrics for judging quality of quantitative structure-activity relationship predictions: Emphasis on scaling of response data. *Journal of Computational Chemistry*, 34(12), 1071–1082. <https://doi.org/10.1002/jcc.23231>
- Rücker, C., Rücker, G., & Meringer, M. (2007). γ -Randomization and its variants in QSPR/QSAR. *Journal of Chemical Information and Modeling*, 47(6), 2345–2357. <https://doi.org/10.1021/ci700157b>
- Santos, S. M., Nascimento, D. C., Costa, M. C., Neto, A. M. B., & Fregolente, L. V. (2020). Flash point prediction: Reviewing empirical models for hydrocarbons, petroleum fraction, biodiesel, and blends. *Fuel*, 263, 116375. <https://doi.org/10.1016/j.fuel.2019.116375>
- Scarim, C. B., Chelucci, R. C., Dos Santos, J. L., & Chin, C. M. (2020). The use of sulfonamide derivatives in the treatment of trypanosomatid parasites including Trypanosoma cruzi, Trypanosoma brucei, and Leishmania spp. *Medicinal Chemistry (Shariqah (United Arab Emirates))*, 16(1), 24–38. <https://doi.org/10.2174/1573406415666190620141109>
- Schuurmann, G., Ebert, R. U., Chen, J., Wang, B., & Kuhne, R. (2008). External validation and prediction employing the predictive squared correlation coefficient test set activity mean vs training set activity mean. *Journal of Chemical Information and Modeling*, 48(11), 2140–2145. <https://doi.org/10.1021/ci800253u>
- Shi, L. M., Fang, H., Tong, W., Wu, J., Perkins, R., Blair, R. M., Branham, W. S., Dial, S. L., Moland, C. L., & Sheehan, D. M. (2001). QSAR models using a large diverse set of estrogens. *Journal of Chemical Information and Computer Sciences*, 41(1), 186–195. <https://doi.org/10.1021/ci000066d>
- Singh, B., Varikuti, S., Halsey, G., Volpedo, G., Hamza, O. M., & Satoskar, A. R. (2019). Host-directed therapies for parasitic diseases. *Future Medicinal Chemistry*, 11(15), 1999–2018. <https://doi.org/10.4155/fmc-2018-0439>
- Tear, W. F., Bag, S., Diaz-Gonzalez, R., Ceballos-Pérez, G., Rojas-Barros, D. I., Cordon-Obras, C., Pérez-Moreno, G., García-Hernández, R., Martínez-Martínez, M. S., Ruiz-Perez, L. M., Gamarro, F., González Pacanowska, D., Caffrey, C. R., Ferrins, L., Manzano, P., Navarro, M., & Pollastri, M. P. (2020). Selectivity and Physicochemical Optimization of Repurposed Pyrazolo[1,5-b]pyridazines for the Treatment of Human African Trypanosomiasis. *Journal of Medicinal Chemistry*, 63(2), 756–783. <https://doi.org/10.1021/acs.jmedchem.9b01741>
- Toropova, A. P., & Toropov, A. A. (2017). The index of ideality of correlation: A criterion of predictability of QSAR models for skin permeability? *The Science of the Total Environment*, 586, 466–472. <https://doi.org/10.1016/j.scitotenv.2017.01.198>
- Toropova, A. P., & Toropov, A. A. (2019a). Does the index of ideality of correlation detect the better model correctly? *Molecular Informatics*, 38(8-9), 1800157. <https://doi.org/10.1002/minf.201800157>
- Toropova, A. P., & Toropov, A. A. (2019b). The index of ideality of correlation: Improvement of models for toxicity to algae. *Natural Product Research*, 33(15), 2200–2207. <https://doi.org/10.1080/14786419.2018.1493591>

- Toropova, A. P., & Toropov, A. A. (2019c). QSPR and nano-QSPR: What is the difference? *Journal of Molecular Structure*, 1182, 141–149. <https://doi.org/10.1016/j.molstruc.2019.01.040>
- Toropova, A. P., & Toropov, A. A. (2019d). Whether the Validation of the Predictive Potential of Toxicity Models is a Solved Task? *Current Topics in Medicinal Chemistry*, 19(29), 2643–2657. <https://doi.org/10.2174/1568026619666191105111817>
- Toropova, A. P., Toropov, A. A., Carnesecchi, E., Benfenati, E., & Dorne, J. L. (2020). The using of the Index of Ideality of Correlation (IIC) to improve predictive potential of models of water solubility for pesticides. *Environmental Science and Pollution Research International*, 27(12), 13339–13347. <https://doi.org/10.1007/s11356-020-07820-6>
- Toropova, A. P., Toropov, A. A., Veselinovic, A. M., Veselinovic, J. B., Benfenati, E., Leszczynska, D., & Leszczynski, J. (2016). Nano-QSAR: Model of mutagenicity of fullerene as a mathematical function of different conditions. *Ecotoxicology and Environmental Safety*, 124, 32–36. <https://doi.org/10.1016/j.ecoenv.2015.09.038>
- Toropov, A. A., Carbo-Dorca, R., & Toropova, A. P. (2018). Index of Ideality of Correlation: New possibilities to validate QSAR: A case study. *Structural Chemistry*, 29(1), 33–38. <https://doi.org/10.1007/s11224-017-0997-9>
- Toropov, A. A., Raska, I., Jr., Toropova, A. P., Raskova, M., Veselinovic, A. M., & Veselinovic, J. B. (2019). The study of the index of ideality of correlation as a new criterion of predictive potential of QSPR/QSAR-models. *The Science of the Total Environment*, 659, 1387–1394. <https://doi.org/10.1016/j.scitotenv.2018.12.439>
- Toropov, A. A., & Toropova, A. P. (2017). The index of ideality of correlation: A criterion of predictive potential of QSPR/QSAR models? *Mutation Research*, 819, 31–37. <https://doi.org/10.1016/j.mrgentox.2017.05.008>
- Toropov, A. A., & Toropova, A. P. (2018). Predicting cytotoxicity of 2-phenoxyindole derivatives against breast cancer cells using index of ideality of correlation. *Anticancer Research*, 38(11), 6189–6194. <https://doi.org/10.21873/anticanres.12972>
- Toropov, A. A., & Toropova, A. P. (2019). Use of the index of ideality of correlation to improve predictive potential for biochemical endpoints. *Toxicology Mechanisms and Methods*, 29(1), 43–52. <https://doi.org/10.1080/15376516.2018.1506851>
- Toropov, A. A., & Toropova, A. P. (2020). QSPR/QSAR: State-of-Art, Weirdness, the Future. *Molecules*, 25(6), 1292. <https://doi.org/10.3390/molecules25061292>
- Toropov, A. A., Toropova, A. P., Raitano, G., & Benfenati, E. (2019). CORAL: Building up QSAR models for the chromosome aberration test. *Saudi Journal of Biological Sciences*, 26(6), 1101–1106. <https://doi.org/10.1016/j.sjbs.2018.05.013>
- Toropov, A. A., Toropova, A. P., Selvestrel, G., & Benfenati, E. (2019). Idealization of correlations between optimal simplified molecular input-line entry system-based descriptors and skin sensitization. *SAR and QSAR in Environmental Research*, 30(6), 447–455. <https://doi.org/10.1080/1062936X.2019.1615547>
- Varghese, S., Rahmani, R., Russell, S., Deora, G. S., Ferrins, L., Toynton, A., Jones, A., Sykes, M., Kessler, A., Eufrasio, A., Cordeiro, A. T., Sherman, J., Rodriguez, A., Avery, V. M., Piggott, M. J., & Baell, J. B. (2020). Discovery of Potent N-Ethylurea Pyrazole Derivatives as Dual Inhibitors of Trypanosoma brucei and Trypanosoma cruzi. *ACS Medicinal Chemistry Letters*, 11(3), 278–285. <https://doi.org/10.1021/acsmmedchemlett.9b00218>
- Watson, J. A., Strub-Wourgraft, N., Tarral, A., Ribeiro, I., Tarning, J., & White, N. J. (2019). Pharmacokinetic-Pharmacodynamic Assessment of the Hepatic and Bone Marrow Toxicities of the New Trypanoside Fexinidazole. *Antimicrobial Agents and Chemotherapy*, 63(4): e02515–18. <https://doi.org/10.1128/AAC.02515-18>
- Xin, W., Li, Z., Wang, Q., Du, J., Zhu, M., & Zhou, H. (2020). Design and synthesis of α -phenoxy-N-sulfonylphenyl acetamides as Trypanosoma brucei Leucyl-tRNA synthetase inhibitors. *European Journal of Medicinal Chemistry*, 185, 111827. <https://doi.org/10.1016/j.ejmech.2019.111827>

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A Monte Carlo method based QSPR model for prediction of reaction rate constants of hydrated electrons with organic contaminants

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ABSTRACT

The Monte Carlo algorithm was applied to formulate a robust quantitative structure–property relationship (QSPR) model to compute the reaction rate constants of hydrated electron values for a data set of 309 water contaminants containing 125 aliphatic and 184 phenyl-based chemicals. The QSPR models were computed with the hybrid optimal descriptors which were procured by combining the SMILES and hydrogen-suppressed molecular graph for both classes of compounds. Approximately 75% of the total experimental data set was randomly divided into training and invisible training sets, while approximately 25% was divided into calibration and validation sets. The authenticity and robustness of the developed QSPR models were also judged by the Index of Ideality of Correlation. In QSPR modelling of aliphatic compounds, the numerical values of r^2_{Training} , $r^2_{\text{Validation}}$, Q^2_{Training} and $Q^2_{\text{Validation}}$ were in the range of 0.852–0.905, 0.815–0.894, 0.839–0.897 and 0.737–0.867, respectively. Whereas, in the QSPR modelling of phenyl-based compounds, the numerical values of r^2_{Training} , $r^2_{\text{Validation}}$, Q^2_{Training} and $Q^2_{\text{Validation}}$ were in the range of 0.867–0.896, 0.852–0.865, 0.816–0.850 and 0.760–0.762, respectively. The structural attributes, which are promoters of $\log K_{e_{aq}}$ increase/decrease are also extracted from the SMILES notation for mechanistic interpretation. These QSPR models can also be applied to compute the reaction rate constants of organic contaminants.

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Introduction

Currently, one notable problem is water pollution associated with contaminants released by industrial effluent wastes, leakage from water tanks, marine dumping, radioactive waste and atmospheric deposition. The quality of water is important for health in both developing and developed countries. Water pollution has a direct effect on human health, crops and the industrial sector. A literature survey shows that various methods such as chemical oxidation technologies, nanofiltration, reverse osmosis and advanced oxidation process have been used to eliminate contaminants, especially for organic compounds [1]. Biological or chemical

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results of the models were obtained using the hybrid descriptor, that is both molecular graph (HSG) and SMILES descriptor. Split 2 gave the leading model for the aliphatic data set, and in the case of phenyl-based compounds the leading model was given by split 3. The described approach provides the possibility of predictions of rate constants of hydrated electrons with new organic contaminants. The authenticity and robustness of the developed models were predicted by various statistical parameters such as r^2 , CCC, IIC, Q_{F1}^2 , Q_{F2}^2 , Q_{F3}^2 , r_m^2 , CR_p^2 etc. The structural attributes, which are promoters of $\log K_{e_{aq}^-}$ increase or decrease, are recognized from the leading models. The present hybrid QSPR models are more robust and predictive than models reported in the literature. In the present paper, the developed QSPR models are based on molecular structures and these do not involve the use of 3D molecular descriptors, physicochemical descriptors, and/or descriptors of quantum mechanics. Hence, it can be concluded that the present QSPR method can be used to predict the $\log K_{e_{aq}^-}$ of 309 water contaminants containing 125 aliphatic and 184 phenyl-based compounds using the Monte Carlo method with index of ideality correlation.

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Disclosure statement

No potential conflict of interest was reported by the authors.

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References

- [1] X. Li, Y.-M. Sun, Z.-Y. Zhang, N.-X. Feng, H. Song, Y.-L. Liu, L. Hai, J.-M. Cao, and G.P. Wang, *Visible light-driven multi-motion modes CNC/TiO₂ nanomotors for highly efficient degradation of emerging contaminants*, Carbon 155 (2019), pp. 195–203. doi:[10.1016/j.carbon.2019.08.039](https://doi.org/10.1016/j.carbon.2019.08.039).
- [2] H. Kušić, B. Rasulev, D. Leszczynska, J. Leszczynski, and N. Koprivanac, *Prediction of rate constants for radical degradation of aromatic pollutants in water matrix: A QSAR study*, Chemosphere 75 (2009), pp. 1128–1134. doi:[10.1016/j.chemosphere.2009.01.019](https://doi.org/10.1016/j.chemosphere.2009.01.019).
- [3] I. Ivančev-Tumbas, *The fate and importance of organics in drinking water treatment: A review*, Environ. Sci. Pollut. R. 21 (2014), pp. 11794–11810.
- [4] K. Yu, X. Li, L. Chen, J. Fang, H. Chen, Q. Li, N. Chi, and J. Ma, *Mechanism and efficiency of contaminant reduction by hydrated electron in the sulfite/iodide/UV process*, Water Res. 129 (2018), pp. 357–364. doi:[10.1016/j.watres.2017.11.030](https://doi.org/10.1016/j.watres.2017.11.030).
- [5] G.V. Buxton, C.L. Greenstock, W.P. Helman, and A.B. Ross, *Critical review of rate constants for reactions of hydrated electrons, hydrogen atoms and hydroxyl radicals (OH/O⁻ in aqueous solution*, J. Phys. Chem. Ref. Data 17 (1988), pp. 513–886. doi:[10.1063/1.555805](https://doi.org/10.1063/1.555805).
- [6] Y. Gu, T. Liu, H. Wang, H. Han, and W. Dong, *Hydrated electron based decomposition of perfluorooctane sulfonate (PFOS) in the VUV/sulfite system*, Sci. Total Environ. 607 (2017), pp. 541–548. doi:[10.1016/j.scitotenv.2017.06.197](https://doi.org/10.1016/j.scitotenv.2017.06.197).

- [24] Manisha, S. Chauhan, P. Kumar, and A. Kumar, *Development of prediction model for fructose-1, 6-bisphosphatase inhibitors using the Monte Carlo method*, SAR QSAR Environ. Res. 30 (2019), pp. 145–159. doi:[10.1080/1062936X.2019.1568299](https://doi.org/10.1080/1062936X.2019.1568299).
- [25] P. Kumar, A. Kumar, and J. Sindhu, *Design and development of novel focal adhesion kinase (FAK) inhibitors using Monte Carlo method with index of ideality of correlation to validate QSAR*, SAR QSAR Environ. Res. 30 (2019), pp. 63–80. doi:[10.1080/1062936X.2018.1564067](https://doi.org/10.1080/1062936X.2018.1564067).
- [26] S. Gupta, N. Basant, D. Mohan, and K.P. Singh, *Modeling the reactivities of hydroxyl radical and ozone towards atmospheric organic chemicals using quantitative structure-reactivity relationship approaches*, Environ. Sci. Pollut. Res. 23 (2016), pp. 14034–14046. doi:[10.1007/s11356-016-6527-2](https://doi.org/10.1007/s11356-016-6527-2).
- [27] H. Su, C. Yu, Y. Zhou, L. Gong, Q. Li, P.J. Alvarez, and M. Long, *Quantitative structure–activity relationship for the oxidation of aromatic organic contaminants in water by TAML/H₂O₂*, Water Res. 140 (2018), pp. 354–363. doi:[10.1016/j.watres.2018.04.062](https://doi.org/10.1016/j.watres.2018.04.062).
- [28] T. Ye, Z. Wei, R. Spinney, D.D. Dionysiou, S. Luo, L. Chai, Z. Yang, and R. Xiao, *Quantitative structure–activity relationship for the apparent rate constants of aromatic contaminants oxidized by ferrate (VI)*, Chem. Eng. J. 317 (2017), pp. 258–266. doi:[10.1016/j.cej.2017.02.061](https://doi.org/10.1016/j.cej.2017.02.061).
- [29] X. Jin, S. Peldszus, and D.I. Sparkes, *Modeling ozone reaction rate constants of micropollutants using quantitative structure–property relationships*, Ozone Sci. Eng. 36 (2014), pp. 289–302. doi:[10.1080/01919512.2014.910444](https://doi.org/10.1080/01919512.2014.910444).
- [30] X. Long and J. Niu, *Estimation of gas-phase reaction rate constants of alkylnaphthalenes with chlorine, hydroxyl and nitrate radicals*, Chemosphere 67 (2007), pp. 2028–2034. doi:[10.1016/j.chemosphere.2006.11.021](https://doi.org/10.1016/j.chemosphere.2006.11.021).
- [31] R. Xiao, T. Ye, Z. Wei, S. Luo, Z. Yang, and R. Spinney, *Quantitative structure–activity relationship (QSAR) for the oxidation of trace organic contaminants by sulfate radical*, Environ. Sci. Technol. 49 (2015), pp. 13394–13402. doi:[10.1021/acs.est.5b03078](https://doi.org/10.1021/acs.est.5b03078).
- [32] C. Li, S. Zheng, T. Li, J. Chen, J. Zhou, L. Su, Y.-N. Zhang, J.C. Crittenden, S. Zhu, and Y. Zhao, *Quantitative structure–activity relationship models for predicting reaction rate constants of organic contaminants with hydrated electrons and their mechanistic pathways*, Water Res. 151 (2019), pp. 468–477. doi:[10.1016/j.watres.2018.12.010](https://doi.org/10.1016/j.watres.2018.12.010).
- [33] P. Achary, A. Toropova, and A. Toropov, *Combinations of graph invariants and attributes of simplified molecular input-line entry system (SMILES) to build up models for sweetness*, Food Res. Int. 122 (2019), pp. 40–46. doi:[10.1016/j.foodres.2019.03.067](https://doi.org/10.1016/j.foodres.2019.03.067).
- [34] A. Kumar, J. Sindhu, and P. Kumar, *In-silico identification of fingerprint of pyrazolyl sulfonamide responsible for inhibition of N-myristoyltransferase using Monte Carlo method with index of ideality of correlation*, J. Biomol. Struct. Dyn. (2020), pp. 1–12. doi:[10.1080/07391102.2020.1784286](https://doi.org/10.1080/07391102.2020.1784286).
- [35] K. Bagri, A. Kumar, M. Nimbhal, and P. Kumar, *Index of ideality of correlation and correlation contradiction index: A confluent perusal on acetylcholinesterase inhibitors*, Mol. Simulat. (2020), pp. 1–10. doi:[10.1080/08927022.2020.1770753](https://doi.org/10.1080/08927022.2020.1770753).
- [36] A.P. Toropova and A.A. Toropov, *CORAL software: Prediction of carcinogenicity of drugs by means of the Monte Carlo method*, Eur. J. Pharm. Sci. 52 (2014), pp. 21–25. doi:[10.1016/j.ejps.2013.10.005](https://doi.org/10.1016/j.ejps.2013.10.005).
- [37] S. Lotfi, S. Ahmadi, and P. Zohrabi, *QSAR modeling of toxicities of ionic liquids toward *Staphylococcus aureus* using SMILES and graph invariants*, Struct. Chem. 31 (2020), pp. 717–739.
- [38] P. Kumar, A. Kumar, and J. Sindhu, *In silico design of diacylglycerol acyltransferase-1 (DGAT1) inhibitors based on SMILES descriptors using Monte-Carlo method*, SAR QSAR Environ. Res. 30 (2019), pp. 525–541. doi:[10.1080/1062936X.2019.1629998](https://doi.org/10.1080/1062936X.2019.1629998).
- [39] A.A. Toropov, R. Carbó-Dorca, and A.P. Toropova, *Index of ideality of correlation: New possibilities to validate QSAR: A case study*, Struct. Chem. 29 (2018), pp. 33–38. doi:[10.1007/s11224-017-0997-9](https://doi.org/10.1007/s11224-017-0997-9).
- [40] A.A. Toropov and A.P. Toropova, *The index of ideality of correlation: A criterion of predictive potential of QSPR/QSAR models?*, Mutat. Res.-Gen. Tox. En. 819 (2017), pp. 31–37. doi:[10.1016/j.mrgentox.2017.05.008](https://doi.org/10.1016/j.mrgentox.2017.05.008).

- [41] S. Ahmadi, *Mathematical modeling of cytotoxicity of metal oxide nanoparticles using the index of ideality correlation criteria*, Chemosphere 242 (2020), pp. 125192. doi:[10.1016/j.chemosphere.2019.125192](https://doi.org/10.1016/j.chemosphere.2019.125192).
- [42] P. Kumar and A. Kumar, *CORAL: QSAR models of CB1 cannabinoid receptor inhibitors based on local and global SMILES attributes with the index of ideality of correlation and the correlation contradiction index*, Chemom. Intell. Lab. 200 (2020), pp. 103982. doi:[10.1016/j.chemolab.2020.103982](https://doi.org/10.1016/j.chemolab.2020.103982).
- [43] C. Rücker, G. Rücker, and M. Meringer, *y-Randomization and its variants in QSPR/QSAR*, J. Chem. Inf. Model. 47 (2007), pp. 2345–2357. doi:[10.1021/ci700157b](https://doi.org/10.1021/ci700157b).
- [44] N. Chirico and P. Gramatica, *Real external predictivity of QSAR models: How to evaluate it? Comparison of different validation criteria and proposal of using the concordance correlation coefficient*, J. Chem. Inf. Model. 51 (2011), pp. 2320–2335. doi:[10.1021/ci200211n](https://doi.org/10.1021/ci200211n).
- [45] G. Schüürmann, R.-U. Ebert, J. Chen, B. Wang, and R. Kühne, *External validation and prediction employing the predictive squared correlation coefficient test set activity mean vs training set activity mean*, J. Chem. Inf. Model. 48 (2008), pp. 2140–2145. doi:[10.1021/ci800253u](https://doi.org/10.1021/ci800253u).
- [46] V. Consonni, R. Todeschini, and M. Pavan, *Structure/response correlations and similarity/diversity analysis by GETAWAY descriptors. 1. Theory of the novel 3D molecular descriptors*, J. Chem. Inform. Comput. Sci. 42 (2002), pp. 682–692. doi:[10.1021/ci015504a](https://doi.org/10.1021/ci015504a).
- [47] V. Consonni, D. Ballabio, and R. Todeschini, *Comments on the definition of the Q₂ parameter for QSAR validation*, J. Chem. Inf. Model. 49 (2009), pp. 1669–1678. doi:[10.1021/ci900115y](https://doi.org/10.1021/ci900115y).
- [48] I. Lawrence and K. Lin, *Assay validation using the concordance correlation coefficient*, Biometrics 45 (1992), pp. 599–604.
- [49] I. Kuzmanovski, A. Wagner, and M. Novič, *Development of models for prediction of the antioxidant activity of derivatives of natural compounds*, Anal. Chim. Acta 868 (2015), pp. 23–35. doi:[10.1016/j.aca.2015.01.050](https://doi.org/10.1016/j.aca.2015.01.050).
- [50] S. Ahmadi, A.P. Toropova, and A.A. Toropov, *Correlation intensity index: Mathematical modeling of cytotoxicity of metal oxide nanoparticles*, Nanotoxicology 14 (2020), pp. 1118–1126.
- [51] M. Javidfar and S. Ahmadi, *QSAR modelling of larvicidal phytocompounds against Aedes aegypti using index of ideality of correlation*, SAR QSAR Environ. Res. 31 (2020), pp. 717–739. doi:[10.1080/1062936X.2020.1806922](https://doi.org/10.1080/1062936X.2020.1806922).
- [52] P. Kumar and A. Kumar, *In silico enhancement of azo dye adsorption affinity for cellulose fibre through mechanistic interpretation under guidance of QSPR models using Monte Carlo method with index of ideality correlation*, SAR QSAR Environ. Res. 31 (2020), pp. 697–715. doi:[10.1080/1062936X.2020.1806105](https://doi.org/10.1080/1062936X.2020.1806105).



In-silico identification of fingerprint of pyrazolyl sulfonamide responsible for inhibition of N-myristoyltransferase using Monte Carlo method with index of ideality of correlation

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In-silico identification of fingerprint of pyrazolyl sulfonamide responsible for inhibition of N-myristoyltransferase using Monte Carlo method with index of ideality of correlation

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ABSTRACT

Human African trypanosomiasis (HAT) or sleeping sickness like infections remain a serious health concern around the globe due to unavailability of safe and potential drugs for their treatment. Moreover, developing safe, potential and highly specific target based treatments is still a challenge for present drug discovery programs. A series of pyrazole based sulfonamides are identified as an inhibitor of *Trypanosoma brucei* N-myristoyltransferase (TbNMT). In the present manuscript, we have developed robust and reliable QSAR models by using the balance of correlation method in CORAL software. The chemical structures are represented by simplified molecular input line entry system (SMILES). The significance of the index of ideality correlation (IIC) with applicability domain (AD) is also studied at depth. The models developed by considering the index of ideality of correlation (IIC) were found to statistically more significant and robust. One QSAR model with best $R^2_{calibration} = 0.8638$ for split 2 was considered as the leading model. A greater value of cR^2 i.e. 0.5 for all models in Y-randomization test showed the robustness of developed models. The outliers and promoters of increase and decrease of endpoint were also extracted independently from the leading models. The mechanistic interpretation of developed models explains the role of different structural attributes in predicting the pIC_{50} of pyrazole sulfonamides extracted from the crystal structure of Leishmania major N-myristoyltransferase (NMT) along with co-crystallized myristoyl-CoA and ligands NMT106, NMT157, NMT187 and NMT236 (PDB ID: 4A2Z, 4A30, 4A32, 2WSA).

Abbreviations: HAT: Human African trypanosomiasis; IIC: Index of Ideality of Correlation; TbNMT: *Trypanosoma brucei* N-myristoyltransferase; CW: Correlation weight; OECD: Organization of Economic Co-operation and Development; QSAR: Quantitative Structure Activity Relationship; CORAL: CORrelation And Logic; AD: Applicability Domain

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Introduction

Human African trypanosomiasis (HAT) or sleeping sickness is caused by the kinetoplastid parasite *Trypanosoma brucei* and it is transmitted by the bite of an infected tsetse fly (*Glossina* genus) (Nagle et al., 2014; Njoroge et al., 2014). Mainly two species i.e. *Trypanosoma brucei gambiense* and *Trypanosoma brucei rhodesiense* of the above protozoan parasite is responsible for the HAT disease which leads to death if not treated properly or treatment is delayed (Klug et al., 2020; Lepovitz et al., 2020; Scarim et al., 2020). The clinical features are characterized by lymphadenopathy, fever and excessive sleepiness due to encephalopathy or encephalitis (Macleod et al., 2020; Singh et al., 2019; Watson et al., 2019). This disease is progressed by two distinct stages: the hemolymphatic stage and central nervous system (CNS) involvement or meningoencephalitic stage. The first stage is initially an acute

stage, starts with the proliferation of parasite in the hemolymphatic system and give rise to non-specific symptoms (Bayliss et al., 2017; Harrison et al., 2018; Klug et al., 2020; Montalvo-Quiros et al., 2015; Patrick et al., 2017). However, the second stage onsets with the migration of parasite from the hemolymphatic system to CNS and cause classical symptoms of sleeping sickness, ultimately leading to coma and death. In the present scenario, the drugs available for HAT treatment are unsafe and frequently allied with severe or life-threatening side effects such as fatal encephalopathy, agranulocytosis, drug-resistance, and myocardial damage. Mainly five drugs (suramin, pentamidine, melarsoprol, eflornithine and nifurtimox) are used to treat various stages of HAT disease (Hagen et al., 2020). For 1st stage treatment, suramin and pentamidine are used, whereas melarsoprol and eflornithine are recommended for stage 2 infection. Recently, the nifurtimox-eflornithine combination therapy (NECT) has

- Kumar, A., & Chauhan, S. (2017). Monte Carlo method based QSAR modelling of natural lipase inhibitors using hybrid optimal descriptors. *SAR and QSAR in Environmental Research*, 28(3), 179–197. <https://doi.org/10.1080/1062936X.2017.1293729>
- Kumar, P., Duhan, M., Sindhu, J., Kadyan, K., Saini, S., & Panigar, N. (2020). Thiazolidine-4-one clubbed pyrazoles hybrids: Potent α -amylase and α -glucosidase inhibitors with NLO properties. *Journal of Heterocyclic Chemistry*, 57(4), 1573–1587. <https://doi.org/10.1002/jhet.3882>
- Kumar, P., & Kumar, A. (2018). Monte Carlo method based QSAR studies of Mer Kinase inhibitors in compliance with OECD principles. *Drug Research*, 68(4), 189–195. <https://doi.org/10.1055/s-0043-119288>
- Kumar, P., & Kumar, A. (2019). Nucleobase sequence based building up of reliable QSAR models with the index of ideality correlation using Monte Carlo method. *Journal of Biomolecular Structure and Dynamics*, online published. <https://doi.org/10.1080/07391102.2019.1656109>
- Kumar, P., & Kumar, A. (2020). CORAL: QSAR models of CB1 cannabinoid receptor inhibitors based on local and global SMILES attributes with the index of ideality of correlation and the correlation contradiction index. *Chemometrics and Intelligent Laboratory Systems*, 200 103982. <https://doi.org/10.1016/j.chemolab.2020...>
- Kumar, P., Kumar, A., & Sindhu, J. (2019a). Design and development of novel focal adhesion kinase (FAK) inhibitors using Monte Carlo method with index of ideality of correlation to validate QSAR. *SAR and QSAR in Environmental Research*, 30(2), 63–80. <https://doi.org/10.1080/1062936X.2018.1564067>
- Kumar, P., Kumar, A., & Sindhu, J. (2019b). In silico design of diacylglycerol acyltransferase-1 (DGAT1) inhibitors based on SMILES descriptors using Monte-Carlo method. *SAR and QSAR in Environmental Research*, 30(8), 525–541. <https://doi.org/10.1080/1062936X.2019.1629998>
- Kumar, P., Kumar, A., Sindhu, J., & Lal, S. (2019). QSAR Models for nitrogen containing monophosphonate and bisphosphonate derivatives as human farnesyl pyrophosphate synthase inhibitors based on Monte Carlo method. *Drug Research*, 69(3), 159–167. <https://doi.org/10.1055/a-0652-5290>
- Lepovitz, L. T., Meis, A. R., Thomas, S. M., Wiedeman, J., Pham, A., Mensa-Wilmot, K., & Martin, S. F. (2020). Design, synthesis, and evaluation of novel anti-trypanosomal compounds. *Tetrahedron*, 76(16), 131086. <https://doi.org/10.1016/j.tet.2020.131086>
- Macleod, O. J. S., Bart, J.-M., MacGregor, P., Peacock, L., Savill, N. J., Hester, S., Ravel, S., Sunter, J. D., Trevor, C., Rust, S., Vaughan, T. J., Minter, R., Mohammed, S., Gibson, W., Taylor, M. C., Higgins, M. K., & Carrington, M. (2020). A receptor for the complement regulator factor H increases transmission of trypanosomes to tsetse flies. *Nature Communications*, 11(1), 1326. <https://doi.org/10.1038/s41467-020-15125-y>
- Manisha, Chauhan, S., Kumar, P., & Kumar, A. (2019). Development of prediction model for fructose-1,6- bisphosphatase inhibitors using the Monte Carlo method. *SAR and QSAR in Environmental Research*, 30(3), 145–159. <https://doi.org/10.1080/1062936X.2019.1568299>
- Masand, V. H., El-Sayed, N. N. E., Bambole, M. U., Patil, V. R., & Thakur, S. D. (2019). Multiple quantitative structure-activity relationships (QSARs) analysis for orally active trypanocidal N-myristoyltransferase inhibitors. *Journal of Molecular Structure*, 1175, 481–487. <https://doi.org/10.1016/j.molstruc.2018.07.080>
- Montalvo-Quiros, S., Taladriz-Sender, A., Kaiser, M., & Dardonville, C. (2015). Antiprotozoal activity and DNA binding of dicationic acridones. *Journal of Medicinal Chemistry*, 58(4), 1940–1949. <https://doi.org/10.1021/jm5018303>
- Nagle, A. S., Khare, S., Kumar, A. B., Supek, F., Buchynskyy, A., Mathison, C. J. N., Chennamaneni, N. K., Pendem, N., Buckner, F. S., Gelb, M. H., & Molteni, V. (2014). Recent developments in drug discovery for leishmaniasis and human African trypanosomiasis. *Chemical Reviews*, 114(22), 11305–11347. <https://doi.org/10.1021/cr500365f>
- Nimbal, M., Bagri, K., Kumar, P., & Kumar, A. (2020). The index of ideality of correlation: A statistical yardstick for better QSAR modeling of glucokinase activators. *Structural Chemistry*, 31(2), 831–839. <https://doi.org/10.1007/s11224-019-01468-w>
- Njoroge, M., Njuguna, N. M., Mutai, P., Ongarora, D. S., Smith, P. W., & Chibale, K. (2014). Recent approaches to chemical discovery and development against malaria and the neglected tropical diseases human African trypanosomiasis and schistosomiasis. *Chemical Reviews*, 114(22), 11138–11163. <https://doi.org/10.1021/cr500098f>
- Ojha, P. K., Mitra, I., Das, R. N., & Roy, K. (2011). Further exploring rm2 metrics for validation of QSPR models. *Chemometrics and Intelligent Laboratory Systems*, 107(1), 194–205. <https://doi.org/10.1016/j.chemolab.2011.03.011>
- Patrick, D. A., Gillespie, J. R., McQueen, J., Hulverson, M. A., Ranade, R. M., Creason, S. A., Herbst, Z. M., Gelb, M. H., Buckner, F. S., & Tidwell, R. R. (2017). Urea Derivatives of 2-Aryl-benzothiazol-5-amines: A new class of potential drugs for human African trypanosomiasis. *Journal of Medicinal Chemistry*, 60(3), 957–971. <https://doi.org/10.1021/acs.jmedchem.6b01163>
- Pratim Roy, P., Paul, S., Mitra, I., & Roy, K. (2009). On two novel parameters for validation of predictive QSAR models. *Molecules (Basel, Switzerland)*, 14(5), 1660–1701. <https://doi.org/10.3390/molecules14051660>
- Roy, K., Chakraborty, P., Mitra, I., Ojha, P. K., Kar, S., & Das, R. N. (2013). Some case studies on application of "r(m)²" metrics for judging quality of quantitative structure-activity relationship predictions: Emphasis on scaling of response data. *Journal of Computational Chemistry*, 34(12), 1071–1082. <https://doi.org/10.1002/jcc.23231>
- Rücker, C., Rücker, G., & Meringer, M. (2007). γ -Randomization and its variants in QSPR/QSAR. *Journal of Chemical Information and Modeling*, 47(6), 2345–2357. <https://doi.org/10.1021/ci700157b>
- Santos, S. M., Nascimento, D. C., Costa, M. C., Neto, A. M. B., & Fregolente, L. V. (2020). Flash point prediction: Reviewing empirical models for hydrocarbons, petroleum fraction, biodiesel, and blends. *Fuel*, 263, 116375. <https://doi.org/10.1016/j.fuel.2019.116375>
- Scarim, C. B., Chelucci, R. C., Dos Santos, J. L., & Chin, C. M. (2020). The use of sulfonamide derivatives in the treatment of trypanosomatid parasites including Trypanosoma cruzi, Trypanosoma brucei, and Leishmania spp. *Medicinal Chemistry (Shariqah (United Arab Emirates))*, 16(1), 24–38. <https://doi.org/10.2174/1573406415666190620141109>
- Schuurmann, G., Ebert, R. U., Chen, J., Wang, B., & Kuhne, R. (2008). External validation and prediction employing the predictive squared correlation coefficient test set activity mean vs training set activity mean. *Journal of Chemical Information and Modeling*, 48(11), 2140–2145. <https://doi.org/10.1021/ci800253u>
- Shi, L. M., Fang, H., Tong, W., Wu, J., Perkins, R., Blair, R. M., Branham, W. S., Dial, S. L., Moland, C. L., & Sheehan, D. M. (2001). QSAR models using a large diverse set of estrogens. *Journal of Chemical Information and Computer Sciences*, 41(1), 186–195. <https://doi.org/10.1021/ci000066d>
- Singh, B., Varikuti, S., Halsey, G., Volpedo, G., Hamza, O. M., & Satoskar, A. R. (2019). Host-directed therapies for parasitic diseases. *Future Medicinal Chemistry*, 11(15), 1999–2018. <https://doi.org/10.4155/fmc-2018-0439>
- Tear, W. F., Bag, S., Diaz-Gonzalez, R., Ceballos-Pérez, G., Rojas-Barros, D. I., Cordon-Obras, C., Pérez-Moreno, G., García-Hernández, R., Martínez-Martínez, M. S., Ruiz-Perez, L. M., Gamarro, F., González Pacanowska, D., Caffrey, C. R., Ferrins, L., Manzano, P., Navarro, M., & Pollastri, M. P. (2020). Selectivity and Physicochemical Optimization of Repurposed Pyrazolo[1,5-b]pyridazines for the Treatment of Human African Trypanosomiasis. *Journal of Medicinal Chemistry*, 63(2), 756–783. <https://doi.org/10.1021/acs.jmedchem.9b01741>
- Toropova, A. P., & Toropov, A. A. (2017). The index of ideality of correlation: A criterion of predictability of QSAR models for skin permeability? *The Science of the Total Environment*, 586, 466–472. <https://doi.org/10.1016/j.scitotenv.2017.01.198>
- Toropova, A. P., & Toropov, A. A. (2019a). Does the index of ideality of correlation detect the better model correctly? *Molecular Informatics*, 38(8-9), 1800157. <https://doi.org/10.1002/minf.201800157>
- Toropova, A. P., & Toropov, A. A. (2019b). The index of ideality of correlation: Improvement of models for toxicity to algae. *Natural Product Research*, 33(15), 2200–2207. <https://doi.org/10.1080/14786419.2018.1493591>

- Toropova, A. P., & Toropov, A. A. (2019c). QSPR and nano-QSPR: What is the difference? *Journal of Molecular Structure*, 1182, 141–149. <https://doi.org/10.1016/j.molstruc.2019.01.040>
- Toropova, A. P., & Toropov, A. A. (2019d). Whether the Validation of the Predictive Potential of Toxicity Models is a Solved Task? *Current Topics in Medicinal Chemistry*, 19(29), 2643–2657. <https://doi.org/10.2174/1568026619666191105111817>
- Toropova, A. P., Toropov, A. A., Carnesecchi, E., Benfenati, E., & Dorne, J. L. (2020). The using of the Index of Ideality of Correlation (IIC) to improve predictive potential of models of water solubility for pesticides. *Environmental Science and Pollution Research International*, 27(12), 13339–13347. <https://doi.org/10.1007/s11356-020-07820-6>
- Toropova, A. P., Toropov, A. A., Veselinovic, A. M., Veselinovic, J. B., Benfenati, E., Leszczynska, D., & Leszczynski, J. (2016). Nano-QSAR: Model of mutagenicity of fullerene as a mathematical function of different conditions. *Ecotoxicology and Environmental Safety*, 124, 32–36. <https://doi.org/10.1016/j.ecoenv.2015.09.038>
- Toropov, A. A., Carbo-Dorca, R., & Toropova, A. P. (2018). Index of Ideality of Correlation: New possibilities to validate QSAR: A case study. *Structural Chemistry*, 29(1), 33–38. <https://doi.org/10.1007/s11224-017-0997-9>
- Toropov, A. A., Raska, I., Jr., Toropova, A. P., Raskova, M., Veselinovic, A. M., & Veselinovic, J. B. (2019). The study of the index of ideality of correlation as a new criterion of predictive potential of QSPR/QSAR-models. *The Science of the Total Environment*, 659, 1387–1394. <https://doi.org/10.1016/j.scitotenv.2018.12.439>
- Toropov, A. A., & Toropova, A. P. (2017). The index of ideality of correlation: A criterion of predictive potential of QSPR/QSAR models? *Mutation Research*, 819, 31–37. <https://doi.org/10.1016/j.mrgentox.2017.05.008>
- Toropov, A. A., & Toropova, A. P. (2018). Predicting cytotoxicity of 2-phenoxyindole derivatives against breast cancer cells using index of ideality of correlation. *Anticancer Research*, 38(11), 6189–6194. <https://doi.org/10.21873/anticanres.12972>
- Toropov, A. A., & Toropova, A. P. (2019). Use of the index of ideality of correlation to improve predictive potential for biochemical endpoints. *Toxicology Mechanisms and Methods*, 29(1), 43–52. <https://doi.org/10.1080/15376516.2018.1506851>
- Toropov, A. A., & Toropova, A. P. (2020). QSPR/QSAR: State-of-Art, Weirdness, the Future. *Molecules*, 25(6), 1292. <https://doi.org/10.3390/molecules25061292>
- Toropov, A. A., Toropova, A. P., Raitano, G., & Benfenati, E. (2019). CORAL: Building up QSAR models for the chromosome aberration test. *Saudi Journal of Biological Sciences*, 26(6), 1101–1106. <https://doi.org/10.1016/j.sjbs.2018.05.013>
- Toropov, A. A., Toropova, A. P., Selvestrel, G., & Benfenati, E. (2019). Idealization of correlations between optimal simplified molecular input-line entry system-based descriptors and skin sensitization. *SAR and QSAR in Environmental Research*, 30(6), 447–455. <https://doi.org/10.1080/1062936X.2019.1615547>
- Varghese, S., Rahmani, R., Russell, S., Deora, G. S., Ferrins, L., Toynton, A., Jones, A., Sykes, M., Kessler, A., Eufrasio, A., Cordeiro, A. T., Sherman, J., Rodriguez, A., Avery, V. M., Piggott, M. J., & Baell, J. B. (2020). Discovery of Potent N-Ethylurea Pyrazole Derivatives as Dual Inhibitors of Trypanosoma brucei and Trypanosoma cruzi. *ACS Medicinal Chemistry Letters*, 11(3), 278–285. <https://doi.org/10.1021/acsmmedchemlett.9b00218>
- Watson, J. A., Strub-Wourgraft, N., Tarral, A., Ribeiro, I., Tarning, J., & White, N. J. (2019). Pharmacokinetic-Pharmacodynamic Assessment of the Hepatic and Bone Marrow Toxicities of the New Trypanoside Fexinidazole. *Antimicrobial Agents and Chemotherapy*, 63(4): e02515–18. <https://doi.org/10.1128/AAC.02515-18>
- Xin, W., Li, Z., Wang, Q., Du, J., Zhu, M., & Zhou, H. (2020). Design and synthesis of α -phenoxy-N-sulfonylphenyl acetamides as Trypanosoma brucei Leucyl-tRNA synthetase inhibitors. *European Journal of Medicinal Chemistry*, 185, 111827. <https://doi.org/10.1016/j.ejmech.2019.111827>



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In silico exploration of the fingerprints triggering modulation of glutaminyl cyclase inhibition for the treatment of Alzheimer's disease using SMILES based attributes in Monte Carlo optimization

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ABSTRACT

Alzheimer's disease is the most common neurodegenerative disorder and being a social burden Alzheimer's has become an economic liability on developing countries. With limited understanding regarding the cause of disease, it is commonly identified by extracellular deposit of amyloid β ($A\beta$) peptides as senile plaques. Pyroglutamated $A\beta$ is identified from the brain of AD patients and constituted the majority of total $A\beta$ present. The formation of Pyroglutamated $A\beta$ could be hindered by the use of Glutaminyl cyclase inhibitors and could efficiently improve the symptoms of Alzheimer's. The literature revealed the competence of quantitative structure activity/property relationship studies in drug discovery. The present work explores the efficiency of Monte Carlo based QSAR modelling studies on a dataset of 125 Glutaminyl cyclase inhibitors with pKi taken as the endpoint for QSAR analysis. The dataset is divided into training, subtraining, calibration and validation sets resulting in the generation of five random splits. The validation is performed in accordance with the Organization of Economic Corporation and Development principles. The values of R^2 , Q^2 , index of ideality of correlation, concordance correlation coefficient, av. r_m^2 and delta r_m^2 of calibration set of the best split are found to be 0.9012, 0.8775, 0.9479, 0.9435, 0.8347 and 0.0847, respectively. The structural features responsible for increasing the inhibitory activity are identified. These structural features are added to a base compound from the dataset to design six novel molecules. These new molecules possess improved inhibitory activity as compare to the base compound. The results are further supported by docking studies.

Abbreviations: AD: Alzheimer's disease; APP: amyloid precursor protein; ASP: aspartic acid; $A\beta$: amyloid beta; BACE: beta site amyloid precursor protein cleaving enzyme; CCC: concordance correlation coefficient; DCW: descriptors of correlation weight; GLN: glutamine; GLU: glutamic acid; HIS: histidine; IIC: index of ideality of correlation; ILE: isoleucine; LEU: leucine; MAE: mean absolute error; OECD: organisation for economic cooperation and development; PDB: protein data bank; PHE: phenylalanine; QSAR: quantitative structure activity relationship; RMSE: root mean square error; SMILES: simplified molecular input line entry system; TRP: tryptophan; ZBG: zinc binding group

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Introduction

Alzheimer's disease (AD) is a prevalent irreversible neurological disease and according to the Brightfocus Foundation, worldwide 50 million people are suffering from Alzheimer's or neurological dementia (Alzheimer's-Disease-International, 2019). The disease is responsible for the social and economic burden. In 2018, the estimated cost of Alzheimer's and dementia care in the US was \$277 billion (Alzheimer's-Association, 2018). The most common identifiable symptom of Alzheimer's is memory loss. As the disease advances from preclinical to late-stage, the symptoms encompass behavioural, cognitive, functional and complete mental decline of the patient (Kumar et al., 2015). Now a days, neuropsychiatric

symptoms like depression, apathy, aggression, and psychosis are identified as main features of AD, and symptoms complexity characterize the disease progression (Lancôt et al., 2017; Li et al., 2014). In addition to genetics, head injuries, depression, diabetes and obesity, age is the most prominent factor leading to AD (Lao et al., 2019).

Clinical diagnosis of AD includes patient history, collateral history from relatives, and clinical remarks, based on the existence of neurological and neuropsychological features. With limited understanding regarding the cause of disease, it is commonly identified by extracellular deposits of amyloid β ($A\beta$) peptides as senile plaques, transmission loss at synapsis, tangles formed by deposition of tau protein and degeneration of cholinergic neurons (Du et al., 2018; Verma et al.,

hydrogen bonding with ILE 303. In GC06, the benzene group was substituted with amino group. In GC06, the coordination bond with Zinc was absent but subsequently, the benzene ring of the molecule showed π - π stacking interactions with HIS 330, TRP 329 and imidazole with PHE 325. The 5-methyl substituent of imidazole ring was seen involved in π -alkyl interactions with ILE 303. The substituted amino group showed hydrogen bonding interactions with ASP 159 and GLU 201.

Conclusion

Five different SMILES based QSAR models were developed on 125 glutaminyl cyclase inhibitors adopting logical and scientific process. The developed models were thoroughly validated, and various validation parameters were found to be within described limits. The studies pointed towards the importance of five-membered heterocyclic ring with three methylene unit spacer attached to two heavy atoms with a terminal phenyl ring. The results also demonstrated the importance of 5-methyl substituted imidazole ring and alkyl-substituted benzene ring in activity enhancement. Structural features extracted from the best QSAR model helped in designing of novel 06 glutaminyl cyclase inhibitor compounds. The newly designed compounds were found to have improved inhibitory potential. The pKi of compound GC05 was found to be 4.05. Docking studies further verified the results as the designed compound showed the crucial binding interactions with the residues of the active site and the binding affinity was in well correlation with the predicted inhibition potential. The results of the present study are very encouraging and would be very useful for researchers working in the field of AD treatment.

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Disclosure statement

There is no conflict of interest.

ORCID

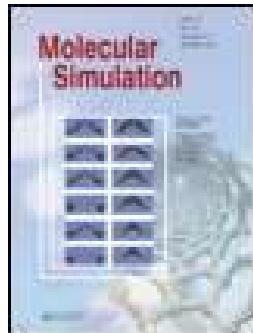
Parvin Kumar  <http://orcid.org/0000-0002-2635-6465>

References

- Ahmadi, S. (2020). Mathematical modeling of cytotoxicity of metal oxide nanoparticles using the index of ideality correlation criteria. *Chemosphere*, 242, 125192. <https://doi.org/10.1016/j.chemosphere.2019.125192>
- Ahmadi, S., Ghanbari, H., Lotfi, S., & Azimi, N. (2020). Predictive QSAR modeling for the antioxidant activity of natural compounds derivatives based on Monte Carlo method. *Molecular Diversity*. <https://doi.org/10.1007/s11030-019-10026-9>
- Ahmadi, S., Mardinia, F., Azimi, N., Qomi, M., & Balali, E. (2019). Prediction of chalcone derivative cytotoxicity activity against MCF-7 human breast cancer cell by Monte Carlo method. *Journal of Molecular Structure*, 1181, 305–311. <https://doi.org/10.1016/j.molstruc.2018.12.089>
- Alzheimer's-Association. (2017). *Beta-amyloid and the amyloid hypothesis*. https://www.alz.org/national/documents/topicssheet_betaamyloid.pdf
- Alzheimer's-Association. (2018). 2018 Alzheimer's disease facts and figures. *Alzheimer's & Dementia*, 14(3), 367–429. <https://doi.org/https://doi.org/10.1016/j.jalz.2018.02.001>
- Alzheimer's-Disease-International. (2019). *World Alzheimer Report 2019: Attitudes to dementia*. Alzheimer's Disease International. <https://www.alz.co.uk/research/WorldAlzheimerReport2019.pdf>
- Amin, S. A., Bhargava, S., Adhikari, N., Gayen, S., & Jha, T. (2018). Exploring pyrazolo[3,4-d]pyrimidine phosphodiesterase 1 (PDE1) inhibitors: A predictive approach combining comparative validated multiple molecular modelling techniques. *Journal of Biomolecular Structure & Dynamics*, 36(3), 590–608. <https://doi.org/10.1080/07391102.2017.1288659>
- Benfenati, E., Toropov, A. A., Toropova, A. P., Manganaro, A., & Gonella Diaza, R. (2011). CORAL Software: QSAR for anticancer agents. *Chemical Biology & Drug Design*, 77(6), 471–476. <https://doi.org/10.1111/j.1747-0285.2011.01117.x>
- Bloom, G. S. (2014). Amyloid- β and Tau: The trigger and bullet in Alzheimer disease pathogenesis. *JAMA Neurology*, 71(4), 505–508. <https://doi.org/10.1001/jamaneurol.2013.5847>
- Bridel, C., Hoffmann, T., Meyer, A., Durieux, S., Koel-Simmelink, M. A., Orth, M., Scheltens, P., Lues, I., & Teunissen, C. E. (2017). Glutaminyl cyclase activity correlates with levels of $\text{A}\beta$ peptides and mediators of angiogenesis in cerebrospinal fluid of Alzheimer's disease patients. *Alzheimer's Research & Therapy*, 9(1), 38. <https://doi.org/10.1186/s13195-017-0266-6>
- Buchholz, M., Hamann, A., Aust, S., Brandt, W., Böhme, L., Hoffmann, T., Schilling, S., Demuth, H.-U., & Heiser, U. (2009). Inhibitors for human glutaminyl cyclase by structure based design and bioisosteric replacement. *Journal of Medicinal Chemistry*, 52(22), 7069–7080. <https://doi.org/10.1021/jm900969p>
- Buchholz, M., Heiser, U., Schilling, S., Niestroj, A. J., Zunkel, K., & Demuth, H.-U. (2006). The first potent inhibitors for human glutaminyl cyclase: Synthesis and structure-activity relationship. *Journal of Medicinal Chemistry*, 49(2), 664–677. <https://doi.org/10.1021/jm050756e>
- Chen, G.-F., Xu, T.-H., Yan, Y., Zhou, Y.-R., Jiang, Y., Melcher, K., & Xu, H. E. (2017). Amyloid beta: Structure, biology and structure-based therapeutic development. *Acta Pharmacologica Sinica*, 38(9), 1205–1235. <https://doi.org/10.1038/aps.2017.28>
- Chirico, N., & Gramatica, P. (2011). Real external predictivity of QSAR models: How to evaluate it? Comparison of different validation criteria and proposal of using the concordance correlation coefficient. *Journal of Chemical Information and Modeling*, 51(9), 2320–2335. <https://doi.org/10.1021/ci200211n>
- Coimbra, J. R. M., Sobral, P. J. M., Santos, A. E., Moreira, P. I., & Salvador, J. A. R. (2019). An overview of glutaminyl cyclase inhibitors for Alzheimer's disease. *Future Medicinal Chemistry*, 11(24), 3179–3194. <https://doi.org/10.4155/fmc-2019-0163>
- Dassault-Systèmes-BIOVIA. (2019). *Discovery studio*. Dassault Systèmes.
- Du, X., Wang, X., & Geng, M. (2018). Alzheimer's disease hypothesis and related therapies. *Translational Neurodegeneration*, 7(1), 2. <https://doi.org/10.1186/s40035-018-0107-y>
- Duhan, M., Singh, R., Devi, M., Sindhu, J., Bhatia, R., Kumar, A., & Kumar, P. (2019). Synthesis, molecular docking and QSAR study of thiazole clubbed pyrazole hybrid as alpha-amylase inhibitor. *Journal of Biomolecular Structure and Dynamics*, 1–17. <https://doi.org/10.1080/07391102.2019.1704885>
- ENV/JM/MONO, O. D. J. (2007). *Guidance Document on the Validation of (Quantitative) Structure-Activity Relationships [(Q) SARs] Models*, 2. Paris: OECD Publishing.
- Golbraikh, A., & Tropsha, A. (2002). Beware of q2!. *Journal of Molecular Graphics and Modelling*, 20(4), 269–276. [https://doi.org/10.1016/S1093-3263\(01\)00123-1](https://doi.org/10.1016/S1093-3263(01)00123-1)
- Golbraikh, A., Wang, X. S., Zhu, H., & Tropsha, A. (2012). Predictive QSAR modeling: Methods and applications in drug discovery and chemical risk assessment. In J. Leszczynski (Ed.), *Handbook of computational chemistry* (pp. 1309–1342). Springer Netherlands.

- Gramatica, P. (2007). Principles of QSAR models validation: Internal and external. *QSAR & Combinatorial Science*, 26(5), 694–701. <https://doi.org/10.1002/qsar.200610151>
- Gramatica, P. (2013). On the development and validation of QSAR models. In B. Reisfeld & A. N. Mayeno (Eds.), *Computational toxicology: Volume II* (pp. 499–526). Humana Press.
- Harigaya, Y., Saido, T. C., Eckman, C. B., Prada, C.-M., Shoji, M., & Younkin, S. G. (2000). Amyloid beta protein starting pyroglutamate at position 3 is a major component of the amyloid deposits in the Alzheimer's disease brain. *Biochemical and Biophysical Research Communications*, 276(2), 422–427. <https://doi.org/10.1006/bbrc.2000.3490>
- Hennekens, C. H., A., Bensadon, B., Zivin, R., & Gaziano, J. M. (2015). Hypothesis: Glutaminyl cyclase inhibitors decrease risks of Alzheimer's disease and related dementias. *Expert Review of Neurotherapeutics*, 15(11), 1245–1248. <https://doi.org/10.1586/14737175.2015.1088784>
- Hillen, H. (2019). The Beta Amyloid Dysfunction (BAD) hypothesis for Alzheimer's disease. *Frontiers in Neuroscience*, 13(1154), 1154. <https://doi.org/10.3389/fnins.2019.01154>
- Hoffmann, T., Meyer, A., Heiser, U., Kurat, S., Böhme, L., Kleinschmidt, M., Bühring, K.-U., Hutter-Paier, B., Farcher, M., Demuth, H.-U., Lues, I., & Schilling, S. (2017). Glutaminyl cyclase inhibitor PQ912 improves cognition in mouse models of Alzheimer's disease—studies on relation to effective target occupancy. *Journal of Pharmacology and Experimental Therapeutics*, 362(1), 119–130. <https://doi.org/10.1124/jpet.117.240614>
- Huang, K.-F., Liaw, S.-S., Huang, W.-L., Chia, C.-Y., Lo, Y.-C., Chen, Y.-L., & Wang, A. H.-J. (2011). Structures of human golgi-resident glutaminyl cyclase and its complexes with inhibitors reveal a large loop movement upon inhibitor binding. *The Journal of Biological Chemistry*, 286(14), 12439–12449. <https://doi.org/10.1074/jbc.M110.208595>
- Islam, M. L., & Gupta, G. K. (2018). Application of Monte Carlo algorithm to explore simplified molecular-input line-entry system based molecular descriptors of BACE1 inhibitors for therapeutic application in Alzheimer's disease. *International Journal of Computer Applications*, 182(11), 40–47. <https://doi.org/10.5120/ijca2018917745>
- Jain, S., Amin, S. A., Adhikari, N., Jha, T., & Gayen, S. (2020). Good and bad molecular fingerprints for human rhinovirus 3C protease inhibition: Identification, validation, and application in designing of new inhibitors through Monte Carlo-based QSAR study. *Journal of Biomolecular Structure & Dynamics*, 38(1), 66–77. <https://doi.org/10.1080/07391102.2019.1566093>
- Jain, S., Bhardwaj, B., Amin, S. A., Adhikari, N., Jha, T., & Gayen, S. (2020). Exploration of good and bad structural fingerprints for inhibition of indoleamine-2,3-dioxygenase enzyme in cancer immunotherapy using Monte Carlo optimization and Bayesian classification QSAR modeling. *Journal of Biomolecular Structure & Dynamics*, 38(6), 1683–1696. <https://doi.org/10.1080/07391102.2019.1615000>
- Kumar, A., & Chauhan, S. (2017). QSAR differential model for prediction of SIRT1 modulation using Monte Carlo method. *Drug Research*, 67(3), 156–162. <https://doi.org/10.1055/s-0042-119725>
- Kumar, A., Sindhu, J., & Kumar, P. (2020). In-silico identification of finger-print of pyrazolyl sulfonamide responsible for inhibition of N-myristoyltransferase using Monte Carlo method with index of ideality of correlation. *Journal of Biomolecular Structure and Dynamics*, 1–12. <https://doi.org/10.1080/07391102.2020.1784286>
- Kumar, A., Singh, A., & Ekaivali, (2015). A review on Alzheimer's disease pathophysiology and its management: An update. *Pharmacological Reports*, 67(2), 195–203. <https://doi.org/10.1016/j.pharep.2014.09.004>
- Kumar, P., & Kumar, A. (2018). Monte Carlo method based QSAR studies of mer kinase inhibitors in compliance with OECD principles. *Drug Research*, 68(4), 189–195. <https://doi.org/10.1055/s-0043-119288>
- Kumar, P., & Kumar, A. (2019). Nucleobase sequence based building up of reliable QSAR models with the index of ideality correlation using Monte Carlo method. *Journal of Biomolecular Structure and Dynamics*, 38(11), 3296–3306. <https://doi.org/10.1080/07391102.2019.1656109>
- Kumar, P., & Kumar, A. (2020). CORAL: QSAR models of CB1 cannabinoid receptor inhibitors based on local and global SMILES attributes with the index of ideality of correlation and the correlation contradiction index. *Chemometrics and Intelligent Laboratory Systems*, 200, 103982. <https://doi.org/10.1016/j.chemolab.2020.103982>
- Kuo, Y.-M., Emmerling, M. R., Woods, A. S., Cotter, R. J., & Roher, A. E. (1997). Isolation, chemical characterization, and quantitation of A beta 3-pyroglutamyl peptide from neuritic plaques and vascular amyloid deposits . *Biochemical and Biophysical Research Communications*, 237(1), 188–191. <https://doi.org/10.1006/bbrc.1997.7083>
- Lanctôt, K. L., Amatniek, J., Ancoli-Israel, S., Arnold, S. E., Ballard, C., Cohen-Mansfield, J., Ismail, Z., Lyketsos, C., Miller, D. S., Musiek, E., Osorio, R. S., Rosenberg, P. B., Satlin, A., Steffens, D., Tariot, P., Bain, L. J., Carrillo, M. C., Hendrix, J. A., Jurgens, H., & Boot, B. (2017). Neuropsychiatric signs and symptoms of Alzheimer's disease: New treatment paradigms. *Alzheimer's & Dementia (New York, N.Y.)*, 3(3), 440–449. <https://doi.org/10.1016/j.trci.2017.07.001>
- Lao, K., Ji, N., Zhang, X., Qiao, W., Tang, Z., & Gou, X. (2019). Drug development for Alzheimer's disease: Review. *Journal of Drug Targeting*, 27(2), 164–173. <https://doi.org/10.1080/1061186X.2018.1474361>
- Li, X.-L., Hu, N., Tan, M.-S., Yu, J.-T., & Tan, L. (2014). Behavioral and psychological symptoms in Alzheimer's disease. *BioMed Research International*, 2014, 927804. <https://doi.org/10.1155/2014/927804>
- Manisha, Chauhan, S., Kumar, P., & Kumar, A. (2019). Development of prediction model for fructose- 1,6- bisphosphatase inhibitors using the Monte Carlo method. *SAR and QSAR in Environmental Research*, 30(3), 145–159. <https://doi.org/10.1080/1062936X.2019.1568299>
- Marvin-Sketch-v.14.11.17.0. (2014). ChemAxon, XhemAxon KFT. Hungary.
- Mitra, A., & Dey, B. (2013). Therapeutic interventions in Alzheimer disease. In U. Kishore (Ed.), *Neurodegenerative diseases*. IntechOpen. <https://www.intechopen.com/books/neurodegenerative-diseases/therapeutic-interventions-in-alzheimer-disease>.
- Nimbal, M., Bagri, K., Kumar, P., & Kumar, A. (2020). The index of ideality of correlation: A statistical yardstick for better QSAR modeling of glucokinase activators. *Structural Chemistry*, 31(2), 831–839. <https://doi.org/10.1007/s11224-019-01468-w>
- O'Boyle, N. M., Banck, M., James, C. A., Morley, C., Vandermeersch, T., & Hutchison, G. R. (2011). Open Babel: An open chemical toolbox. *Journal of Cheminformatics*, 3(1), 33. <https://doi.org/10.1186/1758-2946-3-33>
- Ojha, P. K., Mitra, I., Das, R. N., & Roy, K. (2011). Further exploring rm2 metrics for validation of QSPR models. *Chemometrics and Intelligent Laboratory Systems*, 107(1), 194–205. <https://doi.org/10.1016/j.chemolab.2011.03.011>
- Pedretti, A., Villa, L., & Vistoli, G. (2002). VEGA: A versatile program to convert, handle and visualize molecular structure on Windows-based PCs. *J. Mol. Graph. Model.*, 21(1), 47–49. [https://doi.org/10.1016/s1093-3263\(02\)00123-7](https://doi.org/10.1016/s1093-3263(02)00123-7)
- Ramsbeck, D., Buchholz, M., Koch, B., Böhme, L., Hoffmann, T., Demuth, H.-U., & Heiser, U. (2013). Structure-activity relationships of benzimidazole-based glutaminyl cyclase inhibitors featuring a heteroaryl scaffold. *Journal of Medicinal Chemistry*, 56(17), 6613–6625. <https://doi.org/10.1021/jm4001709>
- Ricciarelli, R., & Fedele, E. (2017). The amyloid cascade hypothesis in Alzheimer's disease: It's time to change our mind. *Current Neuropharmacology*, 15(6), 926–935. <https://doi.org/10.2174/1570159X1566170116143743>
- Roy, K. (2017). *MLR Plus Validation*. Kolkata, India: Jadhavpur University. Retrieved from <http://dtclab.webs.com/software-tools> and http://teqip.jdvu.ac.in/QSAR_Tools/
- Roy, K., & Kar, S. (2014). The rm2 metrics and regression through origin approach: Reliable and useful validation tools for predictive QSAR models (Commentary on 'Is regression through origin useful in external validation of QSAR models?'). *European Journal of Pharmaceutical Sciences: Official Journal of the European Federation for Pharmaceutical Sciences*, 62, 111–114. <https://doi.org/10.1016/j.ejps.2014.05.019>
- Roy, K., Das, R. N., Ambure, P., & Aher, R. B. (2016). Be aware of error measures. Further studies on validation of predictive QSAR models. *Chemometrics and Intelligent Laboratory Systems*, 152, 18–33. <https://doi.org/10.1016/j.chemolab.2016.01.008>
- Roy, K., Kar, S., & Ambure, P. (2015). On a simple approach for determining applicability domain of QSAR models. *Chemometrics and Intelligent Laboratory Systems*, 145, 22–29. <https://doi.org/10.1016/j.chemolab.2015.04.013>
- Roy, K., Kar, S., & Das, R. N. (2015). Chapter 7 - Validation of QSAR models. In K. Roy, S. Kar, & R. N. Das (Eds.), *Understanding the basics of*

- QSAR for applications in pharmaceutical sciences and risk assessment* (pp. 231–289). Academic Press.
- Roy, P. P., Leonard, J. T., & Roy, K. (2008). Exploring the impact of size of training sets for the development of predictive QSAR models. *Chemometrics and Intelligent Laboratory Systems*, 90(1), 31–42. <https://doi.org/10.1016/j.chemolab.2007.07.004>
- Scheltens, P., Hallikainen, M., Grimmer, T., Duning, T., Gouw, A. A., Teunissen, C. E., Wink, A. M., Maruff, P., Harrison, J., van Baal, C. M., Bruins, S., Lues, I., & Prins, N. D. (2018). Safety, tolerability and efficacy of the glutaminyl cyclase inhibitor PQ912 in Alzheimer's disease: Results of a randomized, double-blind, placebo-controlled phase 2a study. *Alzheimer's Research & Therapy*, 10(1), 107. <https://doi.org/10.1186/s13195-018-0431-6>
- Schilling, S., Zeitschel, U., Hoffmann, T., Heiser, U., Francke, M., Kehlen, A., Holzer, M., Hutter-Paier, B., Prokesch, M., Windisch, M., Jagla, W., Schlenzig, D., Lindner, C., Rudolph, T., Reuter, G., Cynis, H., Montag, D., Demuth, H.-U., & Rossner, S. (2008). Glutaminyl cyclase inhibition attenuates pyroglutamate A β and Alzheimer's disease-like pathology. *Nature Medicine*, 14(10), 1106–1111. <https://doi.org/10.1038/nm.1872>
- Selkoe, D. J., & Hardy, J. (2016). The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Molecular Medicine*, 8(6), 595–608. <https://doi.org/10.15252/emmm.201606210>
- Toropov, A. A., & Toropova, A. P. (2020). The Monte Carlo method as a tool to build up predictive QSPR/QSAR. *Current Computer-Aided Drug Design*, 16(3), 197–206. <https://doi.org/10.2174/1573409915666190328123112>
- Toropov, A. A., Toropova, A. P., Raitano, G., & Benfenati, E. (2019). CORAL: Building up QSAR models for the chromosome aberration test. *Saudi Journal of Biological Sciences*, 26(6), 1101–1106. <https://doi.org/10.1016/j.sjbs.2018.05.013>
- Toropov, A. A., Toropova, A. P., Selvestrel, G., & Benfenati, E. (2019). Idealization of correlations between optimal simplified molecular input-line entry system-based descriptors and skin sensitization. *SAR and QSAR in Environmental Research*, 30(6), 447–455. <https://doi.org/10.1080/1062936X.2019.1615547>
- Toropova, A. P., Toropov, A. A., Beeg, M., Gobbi, M., & Salmona, M. (2017). Utilization of the Monte Carlo method to build up QSAR models for hemolysis and cytotoxicity of antimicrobial peptides. *Current Drug Discovery Technologies*, 14(4), 229–243. <https://doi.org/10.2174/157016381466170525114128>
- Toropova, A. P., Toropov, A. A., Veselinović, A. M., Veselinović, J. B., Leszczynska, D., & Leszczynski, J. (2016). Monte Carlo-based quantitative structure-activity relationship models for toxicity of organic chemicals to *Daphnia magna*. *Environmental Toxicology and Chemistry*, 35(11), 2691–2697. <https://doi.org/10.1002/etc.3466>
- Toropova, M. A., Toropov, A. A., Raška, I., & Rašková, M. (2015). Searching therapeutic agents for treatment of Alzheimer disease using the Monte Carlo method. *Computers in Biology and Medicine*, 64, 148–154. <https://doi.org/10.1016/j.combiomed.2015.06.019>
- Trott, O., & Olson, A. J. (2010). AutoDock Vina: Improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *Journal of Computational Chemistry*, 31(2), 455–461. <https://doi.org/10.1002/jcc.21334>
- Velázquez-Libera, J. L., Caballero, J., Toropova, A. P., & Toropov, A. A. (2019). Estimation of 2D autocorrelation descriptors and 2D Monte Carlo descriptors as a tool to build up predictive models for acetylcholinesterase (AChE) inhibitory activity. *Chemometrics and Intelligent Laboratory Systems*, 184, 14–21. <https://doi.org/10.1016/j.chemolab.2018.11.008>
- Verma, S., Kumar, A., Tripathi, T., & Kumar, A. (2018). Muscarinic and nicotinic acetylcholine receptor agonists: Current scenario in Alzheimer's disease therapy. *The Journal of Pharmacy and Pharmacology*, 70(8), 985–993. <https://doi.org/10.1111/jphp.12919>
- Veselinović, J. B., Nikolić, G. M., Trutić, N. V., Živković, J. V., & Veselinović, A. M. (2015). Monte Carlo QSAR models for predicting organophosphate inhibition of acetylcholinesterase. *SAR and QSAR in Environmental Research*, 26(6), 449–460. <https://doi.org/10.1080/1062936X.2015.1049665>
- Vijayan, D. K., & Zhang, K. Y. J. (2019). Human glutaminyl cyclase: Structure, function, inhibitors and involvement in Alzheimer's disease. *Pharmacological Research*, 147, 104342. <https://doi.org/10.1016/j.phrs.2019.104342>



Index of ideality of correlation and correlation contradiction index: a confluent perusal on acetylcholinesterase inhibitors

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Index of ideality of correlation and correlation contradiction index: a confluent perusal on acetylcholinesterase inhibitors

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ABSTRACT

Alzheimer's disease is one of the leading causes of disability and death in the global scenario. Acetylcholinesterase inhibitors are symptomatically involved in the therapeutic management of Alzheimer's disease. Due to the prophetic capability of SMILES-based QSAR studies, the current method explored its efficiency for designing novel inhibitors of acetylcholinesterase enzyme. Two newly introduced validation parameters (the ideality of correlation (IIC) and the correlation contradiction index (CCI)) were studied for further validating the predictive capability of developed models. The index of ideality of correlation was found to have a positive effect on models developed in comparison to models developed without IIC. The structural features accountable for intensifying the inhibitory activity were identified by performing QSAR modelling studies on 60 acetylcholinesterase inhibitors from the literature. Based on the molecular features identified designing of new molecules was accomplished and was found to have satisfactory inhibitory potential. Docking interactions of designed molecules pointed the importance of position of nitro group, aromatic ring and alkyl substitution in influencing the inhibitory activity and binding interactions. The designed compound DD2 was found to have highest inhibitory potential (4.33) and binding affinity ($-11.2 \text{ kcal mol}^{-1}$).

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1. Introduction

With over 40 million people suffering and the sixth leading cause of death, Alzheimer's disease has become the most widespread neurodegenerative disorder. It is not only a social burden but also an economic burden for developing countries. The present lifestyle has also contributed towards the growth of the disease. Regardless of extensive research over the last century the knowledge regarding the exact cause and the ways for its treatment are still incomplete [1–3]. In light of the above facts there are only four FDA-approved drugs e.g. donepezil, rivastigmine, galantamine and memantine that can only slow down the advancement of the disease. The first three drugs are Acetylcholinesterase inhibitors (AChEIs) and are based on Cholinergic hypothesis which states that the reduction in the level of acetylcholine and loss of cholinergic neurons are responsible for learning difficulties and cognitive impairment [4,5]. AChEIs causes inhibition of cholinesterase enzyme responsible for the breakdown of acetylcholine. This improves acetylcholine levels in brains and slows the memory loss.

The last few decades have witnessed a considerable increase in the use of computational methods in drug discovery and development collectively called as Rational Drug Design [6]. Reduction in laboratory and animal testing experiments are the reasons responsible for the diversion of traditional drug design to rational drug design. One of the

cost-effective and predictive techniques of Rational Drug Design is Quantitative Structure Activity Relationship (QSAR). QSAR studies include the mathematical correlation of descriptors of a bioactive molecule with its activity. In QSAR-based drug designing the pharmacokinetic/toxicological parameters are appropriately correlated with potency/selectivity [7–11]. The literature revealed that SMILES-based QSAR studies have remarkable prophetic capability [12–18]. The purpose of the current work is the explanation of CORAL software as a means for QSAR modelling [19], which uses simplified molecular input line entry system (SMILES)-based optimal descriptor [20–22]. Prediction capability is the most important criterion for QSAR model development process. To determine this criterion many statistical methods have been reported in the literature. However, none of them is capable of estimating the prediction ability of QSAR model individually and all are associated with one or more drawbacks. Recently a new criterion of predictability i.e. Index of ideality of correlation (IIC) has been suggested by Toropov [23–25] which is based on correlation coefficient and mean absolute error. In 2019, same authors have introduced a new criterion for prediction i.e. correlation contradiction index (CCI) [26]. It has been shown that there is good correlation between correlation coefficient of validation and CCI. In this work, comparison of IIC and CCI has been performed using a new dataset of 60 AChEIs. The further effect of IIC on CCI has also been investigated. In addition, the

- [22] Toropov AA, Toropova AP, Benfenati E. SMILES-based optimal descriptors: QSAR modeling of carcinogenicity by balance of correlations with ideal slopes. *Eur J Med Chem*. 2010;45(9):3581–3587.
- [23] Toropov AA, Toropova AP. The index of ideality of correlation: a criterion of predictive potential of QSPR/QSAR models. *Mutat Res*. 2017;819:31–37.
- [24] Toropov AA, Raska Jr I, Toropova AP, et al. The study of the index of ideality of correlation as new criterion of predictive potential of QSPR/QSAR-models. *Sci Total Environ*. 2019;659:1387–1394.
- [25] Toropov AA, Toropova AP. Predicting cytotoxicity of 2-phenylindole derivatives against breast cancer cells using index of ideality of correlation. *Anticancer Res*. 2018;38(11):6189–6194.
- [26] Toropov AA, Toropova AP. QSAR as a random event: criteria of predictive potential for a chance model. *Struc Chem*. 2019;30(5):1677–1683.
- [27] Piplani P, Danta C. Design and synthesis of newer potential 4-(N-acetylaminophenol derived piperazine derivatives as potential cognition enhancers. *Bioorg Chem*. 2015;60:64–73.
- [28] Kulshreshtha A, Piplani P. Ameliorative effects of amide derivatives of 1,3,4-thiadiazoles on scopolamine induced cognitive dysfunction. *Eur J Med Chem*. 2016;122:557–573.
- [29] Sharma A, Piplani P. Design and synthesis of some acridine-piperazine hybrids for the improvement of cognitive dysfunction. *Chem Biol Drug Des*. 2017;90(5):926–935.
- [30] Piplani P, Sharma M, Mehta P, et al. N-(4-Hydroxyphenyl)-3,4,5-trimethoxybenzamide derivatives as potential memory enhancers: synthesis, biological evaluation and molecular simulation studies. *J Biomol Struct Dyn*. 2018;36(7):1867–1877.
- [31] Piplani P, Jain A, Devi D, et al. Design, synthesis and pharmacological evaluation of some novel indanone derivatives as acetylcholinesterase inhibitors for the management of cognitive dysfunction. *Bioorg Med Chem*. 2018;26(1):215–224.
- [32] MarvinSketch v.14.11.17.0. ChemAxon, XchemAxon Kft. Budapest, Hungary; 2014.
- [33] O’Boyle NM, Banck M, James CA, et al. Open Babel: an open chemical toolbox. *J Cheminform*. 2011;3:33.
- [34] Kumar A, Chauhan S. Monte Carlo method based QSAR modelling of natural lipase inhibitors using hybrid optimal descriptors. *SAR QSAR Environ Res*. 2017;28(3):179–197.
- [35] Nimbhal M, Chauhan S, Kumar P, et al. Development of prediction model for fructose-1,6-bisphosphatase inhibitors using the Monte Carlo method. *SAR QSAR Environ Res*. 2019;30(3):145–159.
- [36] Worachartcheewan A, Mandi P, Prachayasittikul V, et al. Large-scale QSAR study of aromatase inhibitors using SMILES-based descriptors. *Chemometr Intell Lab*. 2014;138:120–126.
- [37] Toropova AP, Toropov AA, Veselinovic AM, et al. Monte Carlo based quantitative structure-activity relationship models for toxicity of organic chemicals to *Daphnia magna*. *Environ Toxicol Chem*. 2016;35(11):2691–2697.
- [38] Toropov AA, Toropova AP, Como F, et al. Quantitative structure-activity relationship models for bee toxicity. *Toxicol Environ Chem*. 2017;99(7–8):1117–1128.
- [39] Toropova AP, Toropov AA, Rasulev BF, et al. QSAR models for ACE-inhibitor activity of tri-peptides based on representation of the molecular structure by graph of atomic orbitals and SMILES. *Struct Chem*. 2012;23(6):1873–1878.
- [40] Kumar A, Chauhan S. Use of simplified molecular input line entry system and molecular graph based descriptors in prediction and design of pancreatic lipase inhibitors. *Future Med Chem*. 2018;10(13):1603–1622.
- [41] OECD Document. Guidance document on the validation of (quantitative) 1226 structure activity relationships (Q)SARs models. ENV/JM/MONO. 2007;2(2007):1–154.
- [42] Roy K, Kar S, Das RN. Understanding the basics of QSAR for applications in pharmaceutical sciences and risk assessment. San Diego: Academic Press, Elsevier; 2015.
- [43] Roy PP, Leonard JT, Roy K. Exploring the impact of size of training sets for the development of predictive QSAR models. *Chemometr Intell Lab*. 2008;90(1):31–42.
- [44] Roy K. On some aspects of validation of predictive quantitative structure-activity relationship models. *Expert Opin Drug Discov*. 2007;2(12):1567–1577.
- [45] Chirico N, Gramatica P. Real external predictivity of QSAR models: how to evaluate it? Comparison of different validation criteria and proposal of using the concordance correlation coefficient. *J Chem Inf Model*. 2011;51(9):2320–2335.
- [46] Gramatica P. On the development and validation of QSAR models. *Methods Mol Biol*. 2013;930:499–526.
- [47] Gramatica P. Principles of QSAR models validation: internal and external. *QSAR Comb Sci*. 2007;26:694–701.
- [48] Golbraikh A, Tropsha A. Beware of q^2 ! *J Mol Graph Model*. 2002;20(4):269–276.
- [49] Consonni V, Ballabio D, Todeschini R. Evaluation of model predictive ability by external validation techniques. *J Chemometrics*. 2010;24(3–4):194–201.
- [50] Ojha PK, Mitra I, Das RN, et al. Further exploring rm^2 metrics for validation of QSPR models. *Chemometr Intell Lab*. 2011;107(1):194–205.
- [51] Roy K, Kar S. The rm^2 metrics and regression through origin approach: reliable and useful validation tools for predictive QSAR models (commentary on ‘Is regression through origin useful in external validation of QSAR models?’). *Eur J Pharm Sci*. 2014;62:111–114.
- [52] Shayanfar A, Shayanfar S. Is regression through origin useful in external validation of QSAR models? *Eur J Pharm Sci*. 2014;59(1):31–35.
- [53] Aptula N, Jeliazkova G, Schultz TW, et al. The better predictive model: high q^2 for the training set or low root mean square error of prediction for the test set? *QSAR Comb Sci*. 2005;24(3):385–396.
- [54] Roy K, Das RN, Ambure P, et al. Be aware of error measures. Further studies on validation of predictive QSAR models. *Chemometr Intell Lab*. 2016;152:18–33.
- [55] Chirico N, Gramatica P. Real external predictivity of QSAR models. Part 2. New intercomparable thresholds for different validation criteria and the need for scatter plot inspection. *J Chem Inf Model*. 2012;52(8):2044–2058.
- [56] Gramatica P, Sangion A. A historical excursus on the statistical validation parameters for QSAR models: a clarification concerning metrics and terminology. *J Chem Inf Model*. 2016;56(6):1127–1131.
- [57] Toropov AA, Toropova AP. Use of the index of ideality of correlation to improve predictive potential for biochemical endpoints. *Toxicol Mech Methods*. 2019;29(1):43–52.
- [58] Toropova AP, Toropov AA. The index of ideality of correlation: improvement of models for toxicity to algae. *Nat Prod Res*. 2019;33(15):2200–2207.
- [59] Toropova AP, Toropov AA. Does the index of ideality of correlation detect the better model correctly? *Mol. Inform*. 2019;38:1–9.
- [60] Toropov AA, Toropova AP. The correlation contradictions index (CCI): building up reliable models of mutagenic potential of silver nanoparticles under different conditions using quasi-SMILES. *Sci Total Environ*. 2019;681:102–109.
- [61] Mitra I, Saha A, Roy K. Exploring quantitative structure-activity relationship studies of antioxidant phenolic compounds obtained from traditional Chinese medicinal plants. *Mol Simulat*. 2010;36(13):1067–1079.
- [62] Roy K, Kar S, Ambure P. On a simple approach for determining applicability domain of QSAR models. *Chemometr Intell Lab*. 2015;145:22–29.
- [63] Veselinović JB, Veselinovic AM, Toropova AP, et al. The Monte Carlo technique as a tool to predict LOAEL. *Eur J Med Chem*. 2016;116:71–75.
- [64] Trott O, Olson AJ. Autodock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *J Comput Chem*. 2010;31(2):455–461.
- [65] Cheung J, Gary EN, Shiomi K, et al. Structures of human acetylcholinesterase bound to dihydrotanshinone I and territrem B show peripheral site flexibility. *ACS Med Chem Lett*. 2013;4(11):1091–1096.
- [66] Dassault Systèmes BIOVIA. Discovery Studio, 2019. San Diego: Dassault Systèmes; 2019.



Synthesis and characterization of water-soluble chitosan derivatives: spectral, thermal and biological studies

Sohan Lal, Sanjiv Arora, Shikha Rani, Parvin Kumar, Pooja Dabas & Jaideep Malik

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Synthesis and characterization of water-soluble chitosan derivatives: spectral, thermal and biological studies

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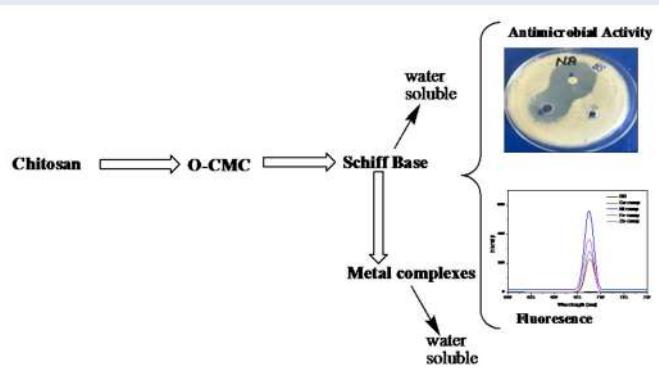
ABSTRACT

In this study, different water-soluble derivatives of chitosan were synthesized by treating chitosan with monochloroacetic acid to yield O-carboxymethyl chitosan (O-CMC) and this O-CMC treated with vanillin to form Schiff base. Metal complexes of this Schiff base were prepared with different salt of metal ions like Cu(II), Ni(II), Fe(II) and Zn(II). O-CMC and its Schiff base were characterized by FTIR, ¹H-NMR and all other derivatives were characterized by using FTIR, UV-visible and fluorescence spectroscopic techniques. In FTIR study, the main characteristic peaks were observed at 1744 cm⁻¹ (C=O str. in O-CMC), 1666 cm⁻¹ (C=N str. peak in Schiff base) and 600–640 cm⁻¹ (metal-ligand str.). In ¹H-NMR different signals were obtained at δ 9.58 ppm due to the proton of imine group (–CH=N– in Schiff base) and signals in between δ 6.5 and 7.5 ppm due to aromatic protons. Antimicrobial activity of all these derivatives was also investigated against *Bacillus subtilis* (Gram-positive bacteria), *Escherichia coli* (Gram-negative bacteria) and *Aspergillus niger* (fungi). The result shows that Ni- and Zn-complexes of O-CMC Schiff base has almost similar activity as that of standard drug ampicillin. Thermal behavior of all these derivatives was also examined by using TG/DTG techniques.

RESEARCH HIGHLIGHTS

- Water-soluble derivatives of chitosan were synthesized (O-CMC, Schiff base and metal complexes).
- All derivatives were characterized by using different techniques (FTIR, ¹H-NMR).
- Thermal, spectral and antimicrobial studies were also carried out.
- All derivatives are water-soluble and thermally stable and exhibited enhanced antimicrobial activity.

GRAPHICAL ABSTRACT



1. Introduction

Natural biopolymers become very popular among the peoples because they have some unique properties of biocompatibility, biodegradability, they can be extracted from natural resources like agriculture and marine food resources and with the additional benefit of their use is that they are

safe for our environment. Chitin is also a natural biopolymer which has all these properties and can be obtained as waste material from the exoskeleton of crustaceans (shrimp, lobster, crab, crawfish, etc.) after food processing.^[1] But chitin is not soluble as such in most of the solvents that reduce its applications so; it is converted into chitosan, which is

which showed that Zn-metal ion binds very efficiently with Schiff base.

4.5. Antimicrobial results

The antimicrobial activity of chitosan and its different water-soluble derivatives were investigated against *B. subtilis* (Gram-positive strain), *E. coli* (Gram-negative strain) and their antifungal activity against *A. niger*. The antibacterial activities of all these compounds were calculated by measuring the zone of inhibition method against the test organisms/specimen. The tested biopolymers possessed variable antibacterial activity against *B. subtilis* with a diameter of zone of inhibitions ranging in between 10 and 32 mm (Figure 5) and against *E. coli* the diameter of zones in between 10 and 26 mm (Figure 5). Among all the tested samples, Zn-complex and Ni-complex were found to be most effective against Gram-positive and Gram-negative bacteria because their zones of inhibition had the largest diameter. The overall results of antibacterial activity showed that all the newly synthesized derivatives are more effective than chitosan against both the strains (*B. subtilis* and *E. coli*) but Zn- and Ni-complexes of Schiff base showed almost equal activity as that of standard drug ampicillin.

On the basis of very good antibacterial activity of Ni-complex and Zn-complex, further investigation was carried out to find their least but sufficient amount to show their antibacterial action in terms of MIC. For this purpose, the optical density (OD) of solution of Ni-complex and Zn-complex of different concentrations containing inocula of *B. subtilis* and *E. coli* at 600 nm was recorded by using spectrophotometer.^[38] The initial concentration of metal complexes of Schiff base was prepared 2000 ppm against *B. subtilis* but the concentration of metal complex against *E. coli* was 3000 ppm and then they were decreased to get optimum result. As the concentration of metal complex solution decreased turbidity of solution increases which showed more growth of bacteria in the solution. The results showed that MIC of Ni-complex and Zn-complex against *B. subtilis* was nearby 1 mg per ml (1000 ppm) and against *E. coli* was between 1.75 mg and 2 mg per ml (1750–2000 ppm).

The antifungal activity of chitosan, O-CMC and its derivatives was tested against *A. niger* by using poisoned food method (Figure 6). In the antifungal activity, among all the tested compounds Zn and Ni metal complexes of Schiff base showed good activity with more than 0.55 inhibition against *A. niger*, whereas other tested compounds showed moderate activity. Thus, the results of antifungal activity showed that all the synthesized derivatives have moderate activity, but Schiff base and its metal complexes were found to be most active as compared to chitosan.

The mechanism of biological activity of chitosan and its derivatives can be explained as negatively charged bacterial membrane interact with the positively charged chitosan molecules and eventually the normal cell got destroyed.^[39] Greater positive charge on -N in case of Schiff base of (-RC=N-) as compared to amine group of polymeric chain of chitosan (-NH₂ group) may be the reason for higher

antimicrobial activity of Schiff base of chitosan. The metal complexes were more biologically active than Schiff base because the density of positive charge on -N increased after chelation of Schiff base with metal ions.^[24]

5. Conclusion

The present work describes the synthesis and characterization of water-soluble derivatives of chitosan i.e., O-CMC, Schiff base of O-CMC and metal complexes of Schiff base with Cu(II), Ni(II), Fe(II) and Zn(II) ions. O-CMC was prepared by reacting chitosan with monochloroacetic acid in strong basic condition and this O-CMC was treated with vanillin to obtain Schiff base and due to the chelating property of Schiff base different metal complexes were prepared. FTIR, ¹H-NMR, UV-visible and fluorescence are some spectroscopic techniques which were used to confirm the structure of all these derivatives. Antimicrobial property of chitosan and its derivatives was investigated against bacteria (*B. subtilis*, *E. coli*) and fungi (*A. niger*). Analysis of results showed that Schiff base and its metal complexes have enhanced antimicrobial activities as compared to chitosan. Among these two derivatives, Zn and Ni-complexes of Schiff base have comparable antimicrobial activity to the standard drug ampicillin. Investigations were also carried out to find out MIC of these compounds by using a broth dilution method. The thermal inquiry of chitosan and its synthesized derivatives showed that they were quite thermally stable and stability of metal complexes was slightly more than Schiff base with also increased in their char yield due to the formation of their metal oxides.

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References

- [1] Prashanth, K. H.; Tharanathan, R. N. Chitin/Chitosan: Modifications and Their Unlimited Application Potential—An Overview. *Trends Food Sci. Technol.* **2007**, *18*, 117–131. DOI: [10.1016/j.tifs.2006.10.022](https://doi.org/10.1016/j.tifs.2006.10.022).
- [2] Saud, R.; Pokhrel, S. P.; Yadav, N. Synthesis, Characterization and Antimicrobial Activity of Maltol Functionalized Chitosan Derivatives. *J. Macromol. Sci. A* **2019**, *56*, 375–383. DOI: [10.1080/10601325.2019.1578616](https://doi.org/10.1080/10601325.2019.1578616).
- [3] Baskar, D.; Kumar, T. S. Effect of Deacetylation Time on the Preparation, Properties and Swelling Behavior of Chitosan Films. *Carbohydr. Polym.* **2009**, *78*, 767–772. DOI: [10.1016/j.carbpol.2009.06.013](https://doi.org/10.1016/j.carbpol.2009.06.013).
- [4] Sabaa, M. W.; Mohamed, N. A.; Mohamed, R. R.; Khalil, N. M.; El Latif, S. M. Synthesis, Characterization and Antimicrobial

- Activity of Poly(N-Vinyl Imidazole) Grafted Carboxymethyl Chitosan. *Carbohydr. Polym.* **2010**, *79*, 998–1005. DOI: [10.1016/j.carbpol.2009.10.024](https://doi.org/10.1016/j.carbpol.2009.10.024).
- [5] Muzzarelli, R. A.; Ilari, P. Chitosans Carrying the Methoxyphenyl Functions Typical of Lignin. *Carbohydr. Polym.* **1994**, *23*, 155–160. DOI: [10.1016/0144-8617\(94\)90097-3](https://doi.org/10.1016/0144-8617(94)90097-3).
- [6] Cocarta, A. I.; Gutanu, V.; Dragan, E. S. Structural, Morphological and Magnetic Characterization of Metal-Chitosan/Poly (Vinyl Amine) Complexes. *J. Polym. Res.* **2017**, *24*, 1182–1193. <https://doi.org/10.1007/s10965-016-1182-3>.
- [7] Pokhrel, S.; Yadav, P. N. Functionalization of Chitosan Polymer and Their Applications. *J. Macromol. Sci. A* **2019**, *56*, 450–475. DOI: [10.1080/10601325.2019.1581576](https://doi.org/10.1080/10601325.2019.1581576).
- [8] Fabris, R.; Chow, C. W.; Drikas, M. Evaluation of Chitosan as a Natural Coagulant for Drinking Water Treatment. *Water Sci. Technol.* **2010**, *61*, 2119–2128. DOI: [10.2166/wst.2010.833](https://doi.org/10.2166/wst.2010.833).
- [9] Guo, Z.; Chen, R.; Xing, R.; Liu, S.; Yu, H.; Wang, P.; Li, C.; Li, P. Novel Derivatives of Chitosan and Their Antifungal Activities *In Vitro*. *Carbohydr. Res.* **2006**, *341*, 351–354. DOI: [10.1016/j.carres.2005.11.002](https://doi.org/10.1016/j.carres.2005.11.002).
- [10] Kong, X. Simultaneous Determination of Degree of Deacetylation, Degree of Substitution and Distribution Fraction of -COONa in Carboxymethyl Chitosan by Potentiometric Titration. *Carbohydr. Polym.* **2012**, *88*, 336–341. DOI: [10.1016/j.carbpol.2011.12.019](https://doi.org/10.1016/j.carbpol.2011.12.019).
- [11] Liu, X.; Song, L.; Li, L.; Li, S.; Yao, K. Antibacterial Effects of Chitosan and Its Water-Soluble Derivatives on *E. coli*, Plasmids DNA, and mRNA. *J. Appl. Polym. Sci.* **2007**, *103*, 3521–3528. DOI: [10.1002/app.25421](https://doi.org/10.1002/app.25421).
- [12] Muzzarelli, R. A. Carboxymethylated Chitins and Chitosans. *Carbohydr. Polym.* **1988**, *8*, 1–21. DOI: [10.1016/0144-8617\(88\)90032-X](https://doi.org/10.1016/0144-8617(88)90032-X).
- [13] Zheng, M.; Han, B.; Yang, Y.; Liu, W. Synthesis, Characterization and Biological Safety of O-Carboxymethyl Chitosan Used to Treat Sarcoma 180 Tumor. *Carbohydr. Polym.* **2011**, *86*, 231–238. DOI: [10.1016/j.carbpol.2011.04.038](https://doi.org/10.1016/j.carbpol.2011.04.038).
- [14] Sun, S.; Wang, A. Adsorption Kinetics of Cu(II) ions Using N,O-Carboxymethyl-Chitosan. *J. Hazard. Mater.* **2006**, *131*, 103–111. DOI: [10.1016/j.jhazmat.2005.09.012](https://doi.org/10.1016/j.jhazmat.2005.09.012).
- [15] Jin, X.; Wang, J.; Bai, J. Synthesis and Antimicrobial Activity of the Schiff Base from Chitosan and Citral. *Carbohydr. Res.* **2009**, *344*, 825–829. DOI: [10.1016/j.carres.2009.01.022](https://doi.org/10.1016/j.carres.2009.01.022).
- [16] Tirkistani, F. A. Thermal Analysis of Some Chitosan Schiff Bases. *Polym. Degrad. Stabil.* **1998**, *60*, 67–70. DOI: [10.1016/S0141-3910\(97\)00020-7](https://doi.org/10.1016/S0141-3910(97)00020-7).
- [17] Jiao, F. T.; Zhou, J.; Zhou, J. X.; Gao, L.; Xing, Y. Y.; Li, X. H. Synthesis and Characterization of Chitosan-Based Schiff Base Compounds with Aromatic Substituent Groups. *Iran. Polym. J.* **2011**, *20*, 123–136.
- [18] dos Santos, J. E.; Dockal, E. R.; Cavalheiro, E. T. Synthesis and Characterization of Schiff Bases from Chitosan and Salicylaldehyde Derivatives. *Carbohydr. Polym.* **2005**, *60*, 277–282. DOI: [10.1016/j.carbpol.2004.12.008](https://doi.org/10.1016/j.carbpol.2004.12.008).
- [19] Becker, T.; Schlaak, M.; Strasdeit, H. Adsorption of Nickel (II), Zinc (II) and Cadmium (II) by New Chitosan Derivatives. *React. Funct. Polym.* **2000**, *44*, 289–298. DOI: [10.1016/S1381-5148\(99\)00104-2](https://doi.org/10.1016/S1381-5148(99)00104-2).
- [20] Monier, M. Adsorption of Hg^{2+} , Cu^{2+} and Zn^{2+} Ions from Aqueous Solution Using Formaldehyde Cross-Linked Modified Chitosan-Thioglyceraldehyde Schiff's Base. *Int. J. Biol. Macromol.* **2012**, *50*, 773–781. DOI: [10.1016/j.ijbiomac.2011.11.026](https://doi.org/10.1016/j.ijbiomac.2011.11.026).
- [21] Wang, R. M.; He, N. P.; Song, P. F.; He, Y. F.; Ding, L.; Lei, Z. Q. Preparation of Nano-Chitosan Schiff-Base Copper Complexes and Their Anticancer Activity. *Polym. Adv. Technol.* **2009**, *20*, 959–964. DOI: [10.1002/pat.1348](https://doi.org/10.1002/pat.1348).
- [22] Adhikari, H. S.; Yadav, P. N. Anticancer Activity of Chitosan, Chitosan Derivatives, and Their Mechanism of Action. *Int. J. Biomater.* **2018**, *2018*, 2952085. DOI: [10.1155/2018/2952085](https://doi.org/10.1155/2018/2952085).
- [23] Wang, X.; Du, Y.; Fan, L.; Liu, H.; Hu, Y. Chitosan-Metal Complexes as Antimicrobial Agent: Synthesis, Characterization and Structure-Activity Study. *Polym. Bull.* **2005**, *55*, 105–113. DOI: [10.1007/s00289-005-0414-1](https://doi.org/10.1007/s00289-005-0414-1).
- [24] Wang, X.; Du, Y.; Liu, H. Preparation, Characterization and Antimicrobial Activity of Chitosan-Zn Complex. *Carbohydr. Polym.* **2004**, *56*, 21–26. DOI: [10.1016/j.carbpol.2003.11.007](https://doi.org/10.1016/j.carbpol.2003.11.007).
- [25] Hardy, J. J.; Hubert, S.; Macquarrie, D. J.; Wilson, A. J. Chitosan-Based Heterogeneous Catalysts for Suzuki and Heck Reactions. *Green Chem.* **2004**, *6*, 53–56. DOI: [10.1039/b312145n](https://doi.org/10.1039/b312145n).
- [26] Lal, S.; Arora, S.; Kumar, V.; Rani, S.; Sharma, C.; Kumar, P. Thermal and Biological Studies of Schiff Bases of Chitosan Derived from Heteroaryl Aldehydes. *J. Therm. Anal. Calorim.* **2018**, *132*, 1707–1716. DOI: [10.1007/s10973-018-7147-5](https://doi.org/10.1007/s10973-018-7147-5).
- [27] Arora, S.; Lal, S.; Kumar, S.; Kumar, M.; Kumar, M. Comparative Degradation Kinetic Studies of Three Biopolymers: Chitin, Chitosan and Cellulose. *Arch. Appl. Sci. Res.* **2011**, *3*, 188–201.
- [28] Chen, X. G.; Park, H. J. Chemical Characteristics of O-Carboxymethylchitosans Related to the Preparation Conditions. *Carbohydr. Polym.* **2003**, *53*, 355–359. DOI: [10.1016/S0144-8617\(03\)00051-1](https://doi.org/10.1016/S0144-8617(03)00051-1).
- [29] Baran, T.; Menteş, A.; Arslan, H. Synthesis and Characterization of Water Soluble O-Carboxymethyl Chitosan Schiff Bases and Cu(II) Complexes. *Int. J. Biol. Macromol.* **2015**, *72*, 94–103. DOI: [10.1016/j.ijbiomac.2014.07.029](https://doi.org/10.1016/j.ijbiomac.2014.07.029).
- [30] Ahmad, I.; Beg, A. Z. Antimicrobial and Phytochemical Studies on 45 Indian Medicinal Plants against Multi-Drug Resistant Human Pathogens. *J. Ethnopharmacol.* **2001**, *74*, 113–123. DOI: [10.1016/S0378-8741\(00\)00335-4](https://doi.org/10.1016/S0378-8741(00)00335-4).
- [31] Andrews, J. M. Determination of Minimum Inhibitory Concentrations. *J. Antimicrob. Chemother.* **2001**, *48*, 5–16. DOI: [10.1093/jac/48.suppl_1](https://doi.org/10.1093/jac/48.suppl_1).
- [32] Abdel-Monem, R. A.; Khalil, A. M.; Darwesh, O. M.; Hashim, A. I.; Rabie, S. T. Antibacterial Properties of Carboxymethyl Chitosan Schiff-Base Nanocomposites Loaded with Silver Nanoparticles. *J. Macromol. Sci. A* **2020**, *57*, 145–155. DOI: [10.1080/10601325.2019.1674666](https://doi.org/10.1080/10601325.2019.1674666).
- [33] Cocarta, A. I.; Gutanu, V.; Dragan, E. S. Structural, Morphological and Magnetic Characterization of Metal-Chitosan/Poly (Vinyl Amine) Complexes. *J. Polym. Res.* **2017**, *24*, 20.
- [34] Boghaei, D. M.; Asl, F. B. Synthesis, Characterization and Fluorescence Spectra of Mixed Ligand Zn (II), Cd (II) and Hg (II) Complexes with 1,10-Phenanthroline-5,6-Dione Ligand. *J. Coord. Chem.* **2007**, *60*, 1629–1635. DOI: [10.1080/00958970601099183](https://doi.org/10.1080/00958970601099183).
- [35] Pawlak, A.; Mucha, M. Thermogravimetric and FTIR Studies of Chitosan Blends. *Thermochim. Acta* **2003**, *396*, 153–166. DOI: [10.1016/S0040-6031\(02\)00523-3](https://doi.org/10.1016/S0040-6031(02)00523-3).
- [36] Antony, R.; David, S. T.; Saravanan, K.; Karuppasamy, K.; Balakumar, S. Synthesis, Spectrochemicalcharacterisation and Catalytic Activity of Transition Metal Complexes Derived from Schiff Base Modified Chitosan. *Spectrochim. Acta Part A* **2013**, *103*, 423–430. DOI: [10.1016/j.saa.2012.09.101](https://doi.org/10.1016/j.saa.2012.09.101).
- [37] Singh, K.; Kumar, Y.; Puri, P.; Kumar, M.; Sharma, C. Cobalt, Nickel, Copper and Zinc Complexes with 1,3-Diphenyl-1H-Pyrazole-4-Carboxaldehyde Schiff Bases: Antimicrobial, Spectroscopic, Thermal and Fluorescence Studies. *Eur. J. Med. Chem.* **2012**, *52*, 313–321. DOI: [10.1016/j.ejmech.2012.02.053](https://doi.org/10.1016/j.ejmech.2012.02.053).
- [38] Devienne, K. F.; Raddi, M. S. Screening for Antimicrobial Activity of Natural Products Using a Microplate Photometer. *Braz. J. Microbiol.* **2002**, *33*, 166–168. DOI: [10.1590/S1517-83822002000200014](https://doi.org/10.1590/S1517-83822002000200014).
- [39] Gupta, D.; Haile, A. Multifunctional Properties of Cotton Fabric Treated with Chitosan and Carboxymethyl Chitosan. *Carbohydr. Polym.* **2007**, *69*, 164–171. DOI: [10.1016/j.carbpol.2006.09.023](https://doi.org/10.1016/j.carbpol.2006.09.023).

Use of Graph Based Descriptors for Determination of Structural Features Causing Modulation of Fructose-1,6-Bisphosphatase

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ABSTRACT

Fructose-1,6-bisphosphatase performs a significant function in regulating the blood glucose level in type 2 diabetes by controlling the process gluconeogenesis. In this research work optimal descriptor (graph) based quantitative structural activity relationship studies of a set of 203 fructose-1,6-bisphosphatase has been performed with the help of Monte Carlo optimization. Distribution of compounds into different sets such as training set, invisible training set, calibration set and validation sets resulted in formation of splits. Statistical parameters obtained from quantitative structural activity relationship modeling were good for various designed splits. The statistical parameters such as R^2 and Q^2 for calibration and validation sets of best split developed were found to be 0.8338, 0.7908 & 0.7920 and 0.7036 respectively. Based on the results obtained for correlation weights, different structural attributes were described as promoters and demoters of the endpoint. Further these structural attributes were used in designing of new fructose-1,6-bisphosphatase inhibitors and molecular docking study was accomplished for the determination of interactions of designed molecules with the enzyme.

Introduction

Diabetes mellitus (DM) is a disorder related to imbalance in carbohydrate, protein and fat metabolism and characterized by hyperglycemia [1]. Hyperglycemia results into microvascular and macrovascular problems such as failure of visualization, heart disease etc [2]. World Health Organization (WHO) has included diabetes in chronic diseases [3]. About 80 % of the total deaths occur due to diabetes every year [4] and the rate at which it is spreading, diabetic patients will enhance up to 300 million in 2025 [5]. Diabetes Mellitus or non-insulin-dependent diabetes mellitus (NIDDM) is regarded as the mostly widespread type of diabetes in all known forms [6]. Although various therapeutics agents are available as anti-diabetic drugs, but still many of them have a number of serious side effects. In medicinal chemistry, development of anti-diabetic drugs with less side effects and relatively low price is still a challenge [7]. From different radioisotopic analysis & some experimentation by

means of ^{13}C NMR, it has been reported that the glucose (in non-absorptive state) is produced in the liver by a process known as the GNG (gluconeogenesis). In Type-2 diabetic (T2D) patients GNG flux is excessive. Hence, to maintain the level of blood glucose in T2D patients, GNG as a pharmacological target represents an attractive approach [8]. Gluconeogenesis involves a principle regulatory enzyme fructose-1,6-bisphosphatase which converts the fructose-1,6-bisphosphate into fructose-6-phosphate and an inorganic phosphate is also released with it [9].

In the modern medicine development practices, to predict the pharmacological action of novel molecules by Quantitative structure activity relationship (QSAR) is assumed as the best option [10]. The major motive of all QSAR modeling process is the development of a model which can correlate pharmacological activity of a molecule with its properties by a simple mathematical equation [11]. For QSPR/QSAR study, the CORALSEA software is one of the best

compounds are given in **Table S7** along with their structures, which are comparable with experimental data given by Dang et al. [16].

Conclusion

In current research work, three QSAR models of a dataset of 203 Fructose-1,6-bisphosphatase inhibitors using graph based optimal descriptors have been developed by CORAL. During QSAR modeling process OECD guidelines were strictly taken into consideration and developed models have good predictions regarding all the statistical parameters. Extracted structural features emphasized the importance of Morgan's extended connectivity, path lengths, vertex numbers and nearest neighboring code in modulation of FBPase inhibitory activity. Incorporation of these graph attributes in the structure of compound D155 resulted in design of novel compounds with increased activity. Further the prediction ability of best QSAR model was confirmed by higher docking score of designed molecules. The suggested novel inhibitors could be tested for their *in vitro* activity in the laboratory and this study will assist the researchers in further design and development of novel FBPase inhibitors.

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Conflict of Interest

There is no conflict of interest.

References

- [1] Blaslov K. Curcumin-a polyphenol with molecular targets for diabetes control. *Endocr Oncol Metab* 2017; 3: 43–48
- [2] Poelje PDV, Dang Q, Erion MD. Fructose-1,6-bisphosphatase as a therapeutic target for type 2 diabetes. *Drug Discov. Today: Therapeutic Strategies* 2007; 4: 103–109
- [3] Tiwari N, Thakur AK, Kumar V et al. Therapeutic targets for diabetes mellitus: An update. *Clin Pharmacol Biopharm* 2014; 3: 1–10
- [4] Kaushik P, Khokra SL, Rana AC et al. Pharmacophore Modeling and Molecular Docking Studies on *Pinus roxburghii* as a Target for Diabetes Mellitus. *Adv Bioinf* 2014; <http://dx.doi.org/10.1155/2014/903246>
- [5] Kaur R, Dahiya L, Kumar M. Fructose-1,6-bisphosphatase inhibitors: A new valid approach for management of type 2 diabetes mellitus. *Eur J Med Chem* 2017; 141: 473–505
- [6] Havale SH, Pal M. Medicinal chemistry approaches to the inhibition of dipeptidyl peptidase-4 for the treatment of type 2 diabetes. *Bioorg Med Chem* 2009; 17: 1783–1802
- [7] Naim MJ, Alam O, Nawaz F. Recent target based discovery of anti-diabetic agents. *IJPSPR* 2015; 6: 4544–4554
- [8] Kitas E, Mohr P, Kuhn B et al. Sulfonylureido thiazoles as fructose-1,6-bisphosphatase inhibitors for the treatment of Type-2 diabetes. *Bioorg Med Chem Lett* 2010; 20: 594–599
- [9] Hines JK, Fromm HJ, Honzatko RB. Novel allosteric activation site in escherichiafructose-1,6-bisphosphatase. *ASBMB* 2006; <http://www.jbc.org/cgi/doi/10.1074/jbc>
- [10] Golubović M, Lazarević M, Zlatanović D et al. The anesthetic action of some polyhalogenated ethers – Monte Carlo method based QSAR study. *Comput Biol Med* 2018; 75: 32–38
- [11] Dearden JC. The history and development of Quantitative Structure-Activity Relationships (QSARs). *IJQSPR*. 2016; 1: 1–44
- [12] Toropov AA, Toropova AP. CORAL. software available at: <http://www.insilico.eu/coral>
- [13] Prachayasittikul V, Worachartcheewan A, Toropova AP et al. Large-scale classification of P-glycoprotein inhibitors using SMILES-based descriptors. *SAR QSAR Environ Res* 2017; 28: 1–16
- [14] Toropova AP, Toropov AA, Martyanov SE et al. CORAL: Monte Carlo Method as a tool for the prediction of the bioconcentration factor of industrial pollutants. *Mol Inf* 2013; 32: 145–154
- [15] Toropov AA, Toropova AP, Benfenati E et al. CORAL: QSPR model of water solubility based on local and global SMILES attributes. *Chemosphere* 2013; 90: 877–880
- [16] Dang Q, Brown BS, Liu Y et al. Fructose-1,6-bisphosphatase Inhibitors. 1. purine phosphonic acids as novel AMP mimics. *J Med Chem* 2009; 52: 2880–2898
- [17] Dang Q, Reddy KR, Kasibhatla SR et al. Discovery of Phosphonic acid-containing desaminobenzimidazoles as Fructose 1,6-Bisphosphatase Inhibitors that are Suitable for oral delivery via Prodrugs. *J Diabetes Metab* 2010; doi:10.4172/2155-6156.1000105
- [18] Dang Q, Kasibhatla SR, Xiao W et al. Fructose-1,6-bisphosphatase Inhibitors. 2. design, synthesis, and structure-activity relationship of a series of phosphonic acid containing benzimidazoles that function as 5'-Adenosinemonophosphate (AMP) Mimics. *J Med Chem* 2010; 53: 441–451
- [19] Dang Q, Kasibhatla SR, Jiang T et al. Oxazole phosphonic acids as fructose 1,6-bisphosphatase inhibitors with potent glucose-lowering activity. *Med Chem Commun* 2011; 2: 287–290
- [20] Dang Q, Liu Y, Cashion DK et al. Discovery of a series of phosphonic acid-containing thiazoles and orally bioavailable diamide prodrugs that lower glucose in diabetic animals through inhibition of fructose-1,6-bisphosphatase. *J Med Chem* 2011; 54: 153–165
- [21] Toropova AP, Toropov AA, Veselinovic JB et al. QSAR models for HEPT derivates as NNRTI inhibitors based on Monte Carlo method. *Eur J Med Chem* 2014; 77: 298–305
- [22] Chadha N, Jasuja H, Kaur M et al. Imidazo[1,2-a]pyrazine inhibitors of phosphoinositide 3-kinase alpha (PI3K α): 3D-QSAR analysis utilizing the Hybrid Monte Carlo algorithm to refine receptor-ligand complexes for molecular alignment. *SAR QSAR Environ Res* 2014; 25: 221–247
- [23] Marvin Sketch v.14.11.17.0 chemAxon, xhemAxon kft. Budapest, Hungary 2014, <https://chemaxon.com/products/marvin>
- [24] O'Boyle N, Banck M, James CA et al. Open Babel: an open chemical toolbox. *J Cheminform* 2011; 3: 33
- [25] Toropov AA, Toropova AP, Como F et al. Quantitative structure-activity relationship models for bee toxicity. *Toxicol Environ Chem* 2016; DOI: 10.1080/02772248.2016.1242006

- [26] Toropov AA, Toropova AP, Martyanov SE et al. CORAL: Predictions of rate constants of hydroxyl radical reaction using representation of the molecular structure obtained by combination of SMILES and Graph approaches. *Chemom Intell Lab Syst* 2012; 112: 65–70
- [27] Kumar A, Chauhan S. QSAR Differential Model for Prediction of SIRT1 Modulation using Monte Carlo Method. *Drug Res* 2016; 67: 156–162
- [28] Toropov AA, Benfenati E. Correlation weighting of valence shells in QSAR analysis of toxicity. *Bioorg Med Chem* 2006; 14: 3923–3928
- [29] Stoičkov V, Šarić S, Golubović M et al. Development of non-peptide ACE inhibitors as novel and potent cardiovascular therapeutics: An in silico modelling approach. *SAR QSAR Environ Res* 2018; 29: 503–515
- [30] Acharya PGR. Simplified molecular input line entry system-based optimal descrip-tors: QSAR modelling for voltage-gated potassium channel subunit Kv7.2. *SAR QSAR Environ Res* 2014; 25: 73–90
- [31] Kumar A, Chauhan S. Use of Simplified Molecular Input Line Entry System and molecular graph based descriptors in prediction and design of pancreatic lipase inhibitors. *Future Med Chem* 2018; 10: 1603–1622
- [32] Roy K. MLRPlusValidation. software available at: <http://dtclab.webs.com/software-tools> and http://teqip.jdvu.ac.in/QSAR_Tools/
- [33] Roy K, Kar S, Ambure P. On a simple approach for determining applicability domain of QSAR models. *Chemom Intell Lab Syst* 2015; 145: 22–29
- [34] Kumar A, Chauhan S. Monte Carlo method based QSAR modeling of natural lipase inhibitors using hybrid optimal descriptors. *SAR QSAR Environ Res* 2017; 28: 179–197
- [35] Kumar A, Chauhan S. Use of the Monte Carlo Method for OECD Principles-Guided QSAR Modeling of SIRT1 Inhibitors. *Arch Pharm Chem Life Sci* 2017; 349: 1–9
- [36] Duchowicz PR, Comelli NC, Ortiz EV et al. QSAR Study for Carcinogenicity in a Large Set of Organic Compounds. *Curr Drug Saf* 2012; 7: 282–288
- [37] Sokolović D, Aleksić D, Milenković V et al. QSAR modeling of bis-quinolinium and bis-isoquinolinium compounds as acetylcholine esterase inhibitors based on the Monte Carlo method—the implication for Myasthenia gravis treatment. *Med Chem Res* 2016; 25: 2989–2998
- [38] Kumar P, Kumar A, Sindhu J et al. QSAR Models for Nitrogen Containing Monophosphonate and Bisphosphonate Derivatives as Human Farnesyl Pyrophosphate Synthase Inhibitors Based on Monte Carlo Method. *Drug Res* 2018; DOI <https://doi.org/10.1055/a-0652-5290>
- [39] Manisha Chauhan S, Kumar A et al. Development of prediction model for fructose-1,6-bisphosphatase inhibitors using the Monte Carlo method. *SAR and QSAR in Environ Res* 2019; 30: 145–159
- [40] Nimbhal M, Bagri K, Kumar A et al. The index of ideality of correlation: A statistical yardstick for better QSAR modeling of glucokinase activators. *Struct Chem* 2019; <https://doi.org/10.1007/s11224-019-01468-w>
- [41] Trott O, Olson AJ. AutoDockVina: Improving the speed and accuracy of docking with a newscore function, efficient optimization, and multithreading. *J Comput Chem* 2010; 31: 455–461
- [42] Pedretti A, Villa L, Vistoli G. “VEGA: a versatile program to convert, handle and visualize molecular structure on windows-based pcs”. *J Mol Graph* 2002; 21: 47–49
- [43] Tsukada T, Takahashi M, Takemoto T et al. Structure-based drug design of tricyclic 8H-indeno[1,2-d][1,3]thiazoles as potent FBPase inhibitors. *Bioorg Med Chem Lett* 2010; 20: 1004–1007
- [44] Lan P, Wu Z, Chen W et al. Molecular modeling studies on phosphonic acid-containing thiazole derivatives: Design for fructose-1, 6-bisphosphatase inhibitors. *J Mol Model* 2011; 18: 973–990
- [45] Discovery Studio Visualizer v19.1.0.18287, Dassault Systemes Biovia Corp, 2018

Bicyclic 5-5 Systems With One Bridgehead (Ring Junction) Nitrogen Atom: Three Extra Heteroatoms 3:0[☆]

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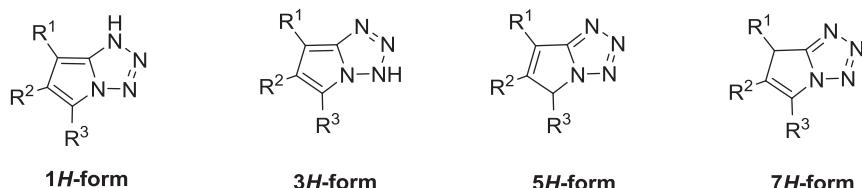
Introduction

The present chapter is an extension of review presented in CHEC-II(1996) (Chapter 11.06) which covers five-membered fused bicyclic systems with three heteroatoms in the same ring and one bridgehead nitrogen atom.¹ Some additional entries based on such fused systems appeared in the literature after the publication of CHEC-II (2015). In this chapter, some of the fused bicyclic compounds are shown which are reported in the literature in terms of scientific findings. The ring systems covered by this chapter are fused tetrazoles.

Theoretical methods

Structure and tautomerism

Various theoretical aspects related to the structure and tautomerism of pyrrolotetrazoles have been discussed in this chapter. The studies on structure and tautomerism were done using quantum-chemical methods (HF/6-31G** and DFT B3LYP/ANO-S) in the gas phase. In the case of pyrrolotetrazoles based heterocyclic systems, theoretically four neutral prototropic forms (1*H*-, 3*H*, 5*H*- and 7*H*-tautomers) are possible (Scheme 1).



Scheme 1

The four theoretically possible tautomeric forms are further categorized into two class based on the aromaticity of the pyrrole ring. In the first case, 1*H*- and 3*H*-tautomers are placed where aromaticity of half pyrrole ring remains intact. On the contrary, in the second case maintenance of aromaticity for pyrrole ring is impossible for 5*H*- and 7*H*-tautomers. The aromaticity of the pyrrole ring in both the cases is controlled solely by the proton transfer from the pyrrole ring to the tetrazole ring or vice versa (Scheme 2).



Scheme 2

[☆]Change History: April 2020. P Kumar and J Sindhu prepared the update. In the section "Energetics of Intramolecular cyclisation", the citation of Table 2 has been changed.

The annular tautomerism in pyrrolo[1,2-*d*]tetrazoles was explored for the first time by Zubarev et al. using quantum-chemical methods (HF/6-31G** and DFT B3LYP/ANO-S) (DFT calculations) in the gas phase.² The total energy was calculated for the four possible tautomers (*i.e.*, 1*H*-, 3*H*-, 5*H*, and 7*H*-forms) to find the stable one among them. The calculation was done using unsubstituted as well as mono-, di- and tri-substituted derivatives to explore the effect of the substituent on the stability of tautomeric forms. Both electron-donating and electron-withdrawing substituents (Me, CN and Cl) were used in various positions of the pyrrole ring help in the evaluation of the influence of electronic nature and their positions on the prototropic tautomerism of these annulated systems. The structures of all possible forms were optimized using DFT method, no negative frequencies were found and the total energies were calculated taking ZPE-corrections into account. It was observed that 5*H*-tautomer is the most stable form for unsubstituted pyrrolotetrazoles and its derivatives containing methyl substituents at the carbon atom(s) in which aromaticity of the tetrazole fragment is maintained. On the contrary, 1*H*-tautomer is the most stable form for pyrrolotetrazoles with electron-withdrawing substituents (CN or Cl) in which the pyrrole fragment is aromatic. A difference in the relative electron-accepting ability of heterocyclic half-rings may affect the stability of such tautomeric forms.

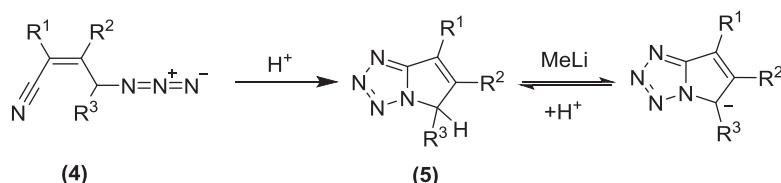
The aromaticity for various tautomeric forms (1*H*-, 3*H*-, 5*H*-, and 7*H*-forms) was calculated using various geometrical parameters (bond lengths) obtained from the optimized structure, where optimization was done using HF/6-31G** method. Pozharskii criterion, based on deviations of interatomic bond orders from the average value in each annulated heterocyclic system, was used for the determination of aromaticity of each of the above-mentioned prototropic forms (Table 1).

Table 1 Aromaticity (in percentage) of various prototropic forms of pyrrolotetrazoles calculated according to the Pozharskii criterion optimized by the HF/6-31G** method.

Form	Aromaticity (A, %)		Integral aromaticity of annulated system
	Pyrrole ring	Tetrazole ring	
1 <i>H</i>	35	28	32
3 <i>H</i>	38	32	35
5 <i>H</i>	10	50	30
7 <i>H</i>	12	50	31

Energetics of intramolecular cycloaddition

An attempt was made by Santelli and co-workers to study the theoretical aspect of intramolecular cycloaddition of azidoenynes and azidobutenenitriles to give 5*H*-pyrrolo[1,2-*d*]tetrazoles.³ The theoretical calculations were done by DFT using B3LYP/6-311+ +G(3df,3pd) level of theory in ideal gas and H₂O as a solvent. The same level of theory was used for the transition state during the cyclisation of the parent compound. The synthesis and alkylation/acylation of 5*H*-pyrrolo[1,2-*d*]tetrazoles was reported by the cyclization of 4-azidobut-2-enenitriles in the acidic medium.^{4,5} In the present chapter, the calculated thermodynamic and kinetic data concerning various steps reported by these workers have been discussed. The thermal cyclization of 4-azidobut-2-enenitriles (4a-4f) to 5a-5f was weakly exergonic and explained the lack of experimental observation in neutral medium (Scheme 3, Table 2). Nevertheless, the use of H₂O as solvent facilitated this thermal cyclization.



Scheme 3

Table 2 Energetics of the thermal cycloaddition of 4-azidobut-2-enenitriles (4a-4f) to 5*H*-pyrrolo[1,2-*d*]tetrazoles (5a-5f).

R	$\Delta E = E_5 - E_4$	$\Delta G = G_5 - G_4$	$\Delta E = E_5 - E_4$	$\Delta G = G_5 - G_4$
	[Kcal mol ⁻¹] ^a	[Kcal mol ⁻¹] ^a	[Kcal mol ⁻¹] ^b	[Kcal mol ⁻¹] ^b
4a → 5a	R ¹ =R ² =R ³ =H	-9.66	-4.37	-13.02
4b → 5b	R ¹ =Me, R ² =R ³ =H	-11.40	-6.28	-14.55
4c → 5c	R ¹ =R ³ =H, R ² =Me	-8.87	-3.58	-12.60
4d → 5d	R ¹ =R ² =Me, R ³ =H	-12.69	-7.17	-16.22
4e → 5e	R ¹ =H, R ² =R ³ =Me	-11.07	-5.86	-14.61
4f → 5f	R ¹ =R ² =R ³ =Me	-14.48	-9.41	-17.81

^aCalculated in ideal gas.

^bCalculated using H₂O as solvent.

Table 6 (Continued)

S. No	Compound	Thallium Flux IC ₅₀
31j		0.082
31k		0.087
31l		0.027
31m		0.036
31n		0.029

References

- Hajós, G.; Riedl, Z. In *Comprehensive Heterocyclic Chemistry III*; Katritzky, A. R., et al. Eds.; Elsevier: Oxford, 2008; pp 645–670.
- Zubarev, V. Y.; et al. *Chem. Heterocycl. Compd.* **2015**, *51* (3), 246–249.
- Audran, G.; et al. *Helv. Chim. Acta* **2015**, *98* (7), 1018–1027.
- Dulcere, J. P.; Tawil, M.; Santelli, M. *J. Org. Chem.* **1990**, *55* (2), 571–575.
- Moderhack, D.; Decker, D. *J. Org. Chem.* **1996**, *61* (16), 5646–5647.
- Sarvary, A.; Khosravi, F.; Ghanbari, M. *Monatshefte für Chem.-Chem. Monthly* **2018**, *149* (1), 39–45.
- Mitsuoka, Y.; et al. *J. Org. Chem.* **2016**, *82* (1), 12–24.
- BIOVIA, D. S. *BIOVIA Discovery Studio Visualizer, v16. 1.0. 15350; Dassault SystemesSan Diego2015*. [cited: Mar 20, 2017].
- Hanessian, S.; et al. *Org. Lett.* **2008**, *10* (7), 1381–1384.
- Carpenter, W. R. *J. Org. Chem.* **1962**, *27* (6), 2085–2088.
- Davis, B. G.; et al. *Tetrahedron* **1999**, *55* (14), 4489–4500.
- Brandstetter, T. W.; et al. *Tetrahedron Lett.* **1995**, *36* (41), 7511–7514.
- Paz, N. R.; et al. *Org. Lett.* **2012**, *14* (13), 3388–3391.
- Miao, H. M.; et al. *Helv. Chim. Acta* **2011**, *94* (11), 1981–1993.
- Kalir, A.; Balderman, D. *Org. Syn.* **1981**, *60*, 104–108.
- Pasternak, A.; et al. *Inhibitors of the Renal Outer Medullary Potassium Channel*; . Google Patents.



Quantitative structure activity relationship studies of novel hydrazone derivatives as α -amylase inhibitors with index of ideality of correlation

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Quantitative structure activity relationship studies of novel hydrazone derivatives as α -amylase inhibitors with index of ideality of correlation

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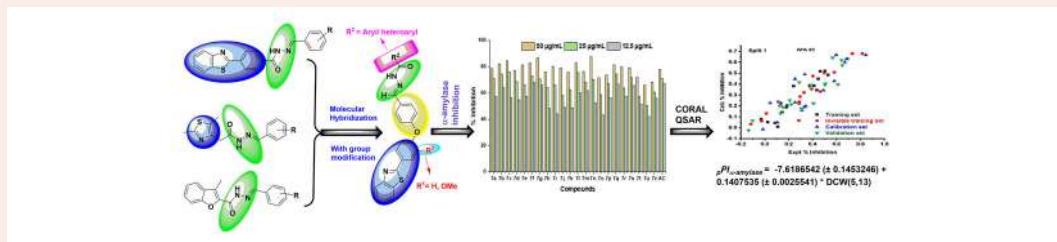
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ABSTRACT

The present manuscript describes the synthesis, α -amylase inhibition, *in silico* studies and in-depth quantitative structure–activity relationship (QSAR) of a library of aroyl hydrazones based on benzothiazole skeleton. All the compounds of the developed library are characterized by various spectral techniques. α -Amylase inhibitory potential of all compounds has been explored, where compound **7n** exhibits remarkable α -amylase inhibition of 87.5% at 50 μ g/mL. Robust QSAR models are made by using the balance of correlation method in CORAL software. The chemical structures at different concentration with optimal descriptors are represented by SMILES. A data set of 66 SMILES of 22 hydrazones at three distinct concentrations are prepared. The significance of the index of ideality of correlation (IIC) with applicability domain (AD) is also studied at depth. A QSAR model with best $R^2_{validation} = 0.8587$ for split 1 is considered as a leading model. The outliers and promoters of increase and decrease of endpoint are also extracted. The binding modes of the most active compound, that is, **7n** in the active site of *Aspergillus oryzae* α -amylase (**PDB ID: 7TAA**) are also explored by *in silico* molecular docking studies. Compound **7n** displays high resemblance in binding mode and pose with the standard drug acarbose. Molecular dynamics simulations performed on protein–ligand complex for 100 ns, the protein gets stabilised after 20 ns and remained below 2 Å for the remaining simulation. Moreover, the deviation observed in RMSF during simulation for each amino acid residue with respect to Cα carbon atom is insignificant.



Abbreviations: IIC: index of ideality of correlation; CW: correlation weight; OECD: Organization of Economic Co-operation and Development; QSAR: quantitative structure–activity relationship; CORAL: CORrelation And Logic; AD: applicability domain; MD: molecular dynamics

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Benzothiazole; aroyl hydrazone; α -amylase inhibition; molecular docking; QSAR; IIC

1. Introduction

Diabetes and obesity are a cluster of metabolic disorders related to lifestyle and characterised by high blood glucose over a prolonged time. The increasing occurrence of these two disorders accelerated the discovery of new drugs. α -Amylase (EC 3.2.1.1) is an endoamylase which mainly occurs in plants, microorganism and higher organisms and belongs to 13th family of glycoside hydrolases (GH13). Its

main function is to hydrolyze the α -D-(1,4)-glycosidic linkage in starch (Brayer et al., 1995; Shi et al., 2018; Souza & Magalhães, 2010) and retaining α -anomeric configuration in the products. The over-expression of α -amylase leads to hyperglycaemia which results in the development of diabetes mellitus. This feature established α -amylase as a well-known molecular target for type 2 diabetes mellitus. Marketed drugs prescribed to treat type-II diabetes mellitus are associated with numerous side effects such as diarrhoea,

the QSAR model built by CORAL Software, AD is calculated by arranging SMILES attributes in the training and calibration sets. If a compound falls outside the range of AD, it is labelled as an outlier. In the CORAL QSAR model, the AD is defined in consonance with the distribution of SMILES characteristics in training and calibration sets as two steps:

Step 1: The definition of statistical defects ($d(F_K)$) for each of the SMILES attributes included to construct the model:

$$d(F_K) = \frac{P_T(A_K) - P_C(A_K)}{N_T(A_K) + N_C(A_K)} \quad (11)$$

where $P_T(A_K)$ and $P_C(A_K)$ are probabilities of attributes A_K in training and calibration set, respectively; $N_T(A_K)$ and $N_C(A_K)$ are the frequency of attributes A_K in the training set and calibration sets, respectively.

Step 2: the calculation for all substances the statistical SMILES defect (D_j):

$$D_j = \sum_{K=1}^{NA} d(F_K) \quad (12)$$

where NA is the number of non-blocked SMILES attributes in the SMILES.

In the current statistical calculation, a compound falls in AD if

$$D_j < 2 \times \overline{\text{DefectNS}} \quad (13)$$

Here, $\overline{\text{DefectNS}}$ is the average of the statistical SMILES defect for the training set.

8.2. Docking studies

Marvin sketch was used for preparing the optimized 3D structure of compounds **7n**. The protein data bank was assessed for the PDB structure of α -amylase for *A. oryzae* (**PDB ID: 7TAA**) (<http://www.rcsb.org/pdb>). The protein was prepared by using UCSF Chimera 1.10 (Pettersen et al., 2004) in which co-crystallized ligand and solvent molecules were removed to avoid interference in binding interactions. Missing side-chain gaps were filled using Dun Brack Rotamer Library (Dunbrack, 2002). Gasteiger charges were calculated using AMBERf14SB and antechamber (Wang et al., 2006) and hydrogens were added. The docking studies were performed using Auto Dock Vina 1.1.2 (Trott & Olson, 2010). Grid center with following size center_x=38.1433640994, center_y=39.1685534078, center_z=31.0477751774, size_x=25.0, size_y=25.0 and size_z=25.0 was placed on the active site. The results of docking studies were analysed using Desmond interface.

8.3. MD simulations

The molecular dynamics simulation of the docked complex of 7TAA.pdb with **7n** was performed for 100 ns using Desmond module of Schrödinger 2019-4 to establish the stability of the docked complex (Guo et al., 2010). The docked poses of protein-ligand complexes were used as input structures and each complex was prepared by system setup option in Desmond module. Explicit solvent system with

OPLS2005 force field was used for this simulation study. Orthorhombic periodic boundary condition for 10 Å buffer region was used for solvation of molecular system with crystallographic water (TIP3P) (Jorgensen et al., 1983) and the system was neutralised by adding Na^+ as counter ions. An ensemble (NPT) of Nose–Hoover thermostat (Martyna et al., 1992, 1994) and barostat was applied to maintain the constant temperature (300 K) and pressure (1 bar) of the systems, respectively. A limited memory algorithm (Broyden–Fletcher–Goldfarb–Shanno (LBFGS)) was employed with convergence threshold gradient of 1 kcal/mol/Å for energy minimization. The data were collected for every 100 ps, and the obtained trajectory was analyzed with Desmond interphase.

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Author contributions

Authors have done equivalent contributions to this work.

Disclosure statement

The authors reported no potential conflict of interest.

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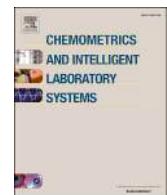
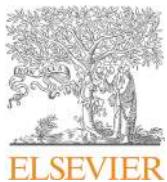
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References

- Achary, P. G. R., Toropova, A. P., & Toropov, A. A. (2019). Combinations of graph invariants and attributes of simplified molecular input-line entry system (SMILES) to build up models for sweetness. *Food Research International* (Ottawa, Ont.), 122, 40–46. <https://doi.org/10.1016/j.foodres.2019.03.067>
- Ahmadi, S. (2020). Mathematical modeling of cytotoxicity of metal oxide nanoparticles using the index of ideality correlation criteria. *Chemosphere*, 242, 125192. <https://doi.org/10.1016/j.chemosphere.2019.125192>
- Ahmadi, S., Ghanbari, H., Lotfi, S., & Azimi, N. (2020). Predictive QSAR modeling for the antioxidant activity of natural compounds derivatives based on Monte Carlo method. *Molecular Diversity*. <https://doi.org/10.1007/s11030-019-10026-9>
- Ahmadi, S., Mehrabi, M., Rezaei, S., & Mardafkan, N. (2019). Structure–activity relationship of the radical scavenging activities of some natural antioxidants based on the graph of atomic orbitals. *Journal of Molecular Structure*, 1191, 165–174. <https://doi.org/10.1016/j.molstruc.2019.04.103>

- Angelova, V., Karabeliov, V., Andreeva-Gateva, P. A., & Tchekalarova, J. (2016). Recent developments of hydrazide/hydrazone derivatives and their analogs as anticonvulsant agents in animal models. *Drug Development Research*, 77(7), 379–392. <https://doi.org/10.1002/ddr.21329>
- Bhatia, R., Kadyan, K., Duhan, M., Devi, M., Singh, R., Kamboj, R. C., & Kumar, P. (2019). A Serendipitous synthesis: $\text{SiO}_2\text{-HNO}_3$ mediated oxidative aromatization and regioselective nitration of 1,3,5-trisubstituted-4,5-dihydro-1*H*-pyrazoles. *ChemistrySelect*, 4(35), 10417–10424. <https://doi.org/10.1002/slct.201902285>
- Bhutani, R., Pathak, D. P., Kapoor, G., Husain, A., Kant, R., & Iqbal, M. A. (2018). Synthesis, molecular modelling studies and ADME prediction of benzothiazole clubbed oxadiazole-Mannich bases, and evaluation of their anti-diabetic activity through *in vivo* model. *Bioorganic Chemistry*, 77, 6–15. <https://doi.org/10.1016/j.bioorg.2017.12.037>
- Brayer, G. D., Luo, Y., & Withers, S. G. (1995). The structure of human pancreatic alpha-amylase at 1.8 Å resolution and comparisons with related enzymes. *Protein Science: A Publication of the Protein Society*, 4(9), 1730–1742. <https://doi.org/10.1002/pro.5560040908>
- Cardoso, L. N. F., Nogueira, T. C. M., Rodrigues, F. A. R., Oliveira, A. C. A., dos Santos Luciano, M. C., Pessoa, C., & de Souza, M. V. N. (2017). N-acylhydrazones containing thiophene nucleus: A new anticancer class. *Medicinal Chemistry Research*, 26(8), 1605–1608. <https://doi.org/10.1007/s00044-017-1832-y>
- Duhan, M., Singh, R., Devi, M., Sindhu, J., Bhatia, R., Kumar, A., & Kumar, P. (2019). Synthesis, molecular docking and QSAR study of thiazole clubbed pyrazole hybrid as α -amylase inhibitor. *Journal of Biomolecular Structure and Dynamics*, 1–31. <https://doi.org/10.1080/07391102.2019.1704885>
- Dunbrack, R. L. Jr. (2002). Rotamer libraries in the 21st century. *Current Opinion in Structural Biology*, 12(4), 431–440. [https://doi.org/10.1016/S0959-440X\(02\)00344-5](https://doi.org/10.1016/S0959-440X(02)00344-5)
- Ferreira, L. G., Dos Santos, R. N., Oliva, G., & Andricopulo, A. D. (2015). Molecular docking and structure-based drug design strategies. *Molecules (Basel, Switzerland)*, 20(7), 13384–13421. <https://doi.org/10.3390/molecules200713384>
- Gollapalli, M., Taha, M., Javid, M. T., Almandil, N. B., Rahim, F., Wadood, A., Mosaddik, A., Ibrahim, M., Alqahtani, M. A., & Bamarouf, Y. A. (2019). Synthesis of benzothiazole derivatives as a potent α -glucosidase inhibitor. *Bioorganic Chemistry*, 85, 33–48. <https://doi.org/10.1016/j.bioorg.2018.12.021>
- Guo, Z., Mohanty, U., Noehre, J., Sawyer, T. K., Sherman, W., & Krilov, G. (2010). Probing the alpha-helical structural stability of stapled p53 peptides: Molecular dynamics simulations and analysis. *Chemical Biology & Drug Design*, 75(4), 348–359. <https://doi.org/10.1111/j.1747-0285.2010.00951.x>
- Hernández-Vázquez, E., Salgado-Barrera, S., Ramírez-Espinosa, J. J., Estrada-Soto, S., & Hernández-Luis, F. (2016). Synthesis and molecular docking of N'-arylidene-5-(4-chlorophenyl)-1-(3,4-dichlorophenyl)-4-methyl-1*H*-pyrazole-3-carbohydrazides as novel hypoglycemic and antioxidant dual agents. *Bioorganic & Medicinal Chemistry*, 24(10), 2298–2306. <https://doi.org/10.1016/j.bmc.2016.04.007>
- Hossain, K. A., & Roy, K. (2018). Chemometric modeling of aquatic toxicity of contaminants of emerging concern (CECs) in *Dugesia japonica* and its interspecies correlation with daphnia and fish: QSTR and QSTTR approaches. *Ecotoxicology and Environmental Safety*, 166, 92–101. <https://doi.org/10.1016/j.ecoenv.2018.09.068>
- Jorgensen, W. L., Chandrasekhar, J., Madura, J. D., Impey, R. W., & Klein, M. L. (1983). Comparison of simple potential functions for simulating liquid water. *The Journal of Chemical Physics*, 79(2), 926–935. <https://doi.org/10.1063/1.445869>
- Karelson, M., Lobanov, V. S., & Katritzky, A. R. (1996). Quantum-chemical descriptors in QSAR/QSPR Studies. *Chemical Reviews*, 96(3), 1027–1044. <https://doi.org/10.1021/cr950202r>
- Kaur, I., Khajuria, A., Ohri, P., Kaur, P., & Singh, K. (2018). Benzothiazole based Schiff-base-A mechanically discrete sensor for HSO_4^- and I^- : Application to bioimaging and vapour phase sensing of ethyl acetate. *Sensors and Actuators B: Chemical*, 268, 29–38. <https://doi.org/10.1016/j.snb.2018.04.072>
- Keharom, S., Mahachai, R., & Chanthai, S. (2016). The optimization study of α -amylase activity based on central composite design-response surface methodology by dinitrosalicylic acid method. *International Food Research Journal*, 23(1), 10–17.
- Khan, K., Benfenati, E., & Roy, K. (2019a). Consensus QSAR modeling of toxicity of pharmaceuticals to different aquatic organisms: Ranking and prioritization of the DrugBank database compounds. *Ecotoxicology and Environmental Safety*, 168, 287–297. <https://doi.org/10.1016/j.ecoenv.2018.10.060>
- Khan, P. M., Roy, K., & Benfenati, E. (2019b). Chemometric modeling of *Daphnia magna* toxicity of agrochemicals. *Chemosphere*, 224, 470–479. <https://doi.org/10.1016/j.chemosphere.2019.02.147>
- Krallinger, M., Rabal, O., Lourenco, A., Oyarzabal, J., & Valencia, A. (2017). Information retrieval and text mining technologies for chemistry. *Chemical Reviews*, 117(12), 7673–7761. <https://doi.org/10.1021/cschemrev.6b00851>
- Kumar, A., & Chauhan, S. (2017). Use of the Monte Carlo method for OECD principles-guided QSAR modeling of SIRT1 inhibitors. *Archiv der Pharmazie (Weinheim)*, 350(1), e1600268. <https://doi.org/10.1002/ardp.201600268>
- Kumar, P., Bhatia, R., Kumar, D., Kamboj, R. C., Kumar, S., Kamal, R., & Kumar, R. (2015). An economic, simple and convenient synthesis of 2-aryl/heteroaryl/styryl/alkylbenzothiazoles using $\text{SiO}_2\text{-HNO}_3$. *Research on Chemical Intermediates*, 41(7), 4283–4292. <https://doi.org/10.1007/s11164-013-1529-x>
- Kumar, P., Duhan, M., Kadyan, K., Bhardwaj, J. K., Saraf, P., & Mittal, M. (2018). Multicomponent synthesis of some molecular hybrid containing thiazole pyrazole as apoptosis inducer. *Drug Research*, 68(2), 72–79. <https://doi.org/10.1055/s-0043-116947>
- Kumar, P., Duhan, M., Kadyan, K., Sindhu, J., Kumar, S., & Sharma, H. (2017a). Synthesis of novel inhibitors of α -amylase based on the thiazolidine-4-one skeleton containing a pyrazole moiety and their configurational studies. *MedChemComm*, 8(7), 1468–1476. <https://doi.org/10.1039/c7md00080d>
- Kumar, P., Kadyan, K., Duhan, M., Sindhu, J., Singh, V., & Saharan, B. S. (2017b). Design, synthesis, conformational and molecular docking study of some novel acyl hydrazone based molecular hybrids as anti-malarial and antimicrobial agents. *Chemistry Central Journal*, 11(1), 115. <https://doi.org/10.1186/s13065-017-0344-7>
- Kumar, P., & Kumar, A. (2019). Nucleobase sequence based building up of reliable QSAR models with the index of ideality correlation using Monte Carlo method. *Journal of Biomolecular Structure and Dynamics*, 38(11), 3296–3306. <https://doi.org/10.1080/07391102.2019.1656109>
- Kumar, P., & Kumar, A. (2020). CORAL: QSAR models of CB1 cannabinoid receptor inhibitors based on local and global SMILES attributes with the index of ideality of correlation and the correlation contradiction index. *Chemometrics and Intelligent Laboratory Systems*, 200, 103982. <https://doi.org/10.1016/j.chemolab.2020.103982>
- Kumar, P., Kumar, A., & Sindhu, J. (2019a). Design and development of novel focal adhesion kinase (FAK) inhibitors using Monte Carlo method with index of ideality of correlation to validate QSAR. *SAR and QSAR in Environmental Research*, 30(2), 63–80. <https://doi.org/10.1080/1062936X.2018.1564067>
- Kumar, P., Kumar, A., & Sindhu, J. (2019b). In silico design of diacylglycerol acyltransferase-1 (DGAT1) inhibitors based on SMILES descriptors using Monte-Carlo method. *SAR and QSAR in Environmental Research*, 30(8), 525–541. <https://doi.org/10.1080/1062936X.2019.1629998>
- Kumar, P., Kumar, A., Sindhu, J., & Lal, S. (2019c). QSAR models for nitrogen containing monophosphonate and bisphosphonate derivatives as human farnesyl pyrophosphate synthase inhibitors based on Monte Carlo method. *Drug Research*, 69(3), 159–167. <https://doi.org/10.1055/a-0652-5290>
- Kumar, S., Rathore, D. S., Garg, G., Khatri, K., Saxena, R., & Sahu, S. K. (2017c). Synthesis and evaluation of some benzothiazole derivatives as antidiabetic agents. *International Journal of Pharmacy and Pharmaceutical Sciences*, 9(2), 60–68. <https://doi.org/10.22159/ijpps.2017v9i2.14359>
- Le, T., Epa, V. C., Burden, F. R., & Winkler, D. A. (2012). Quantitative structure–property relationship modeling of diverse materials properties. *Chemical Reviews*, 112(5), 2889–2919. <https://doi.org/10.1021/cr200066h>

- Macalino, S. J. Y., Gosu, V., Hong, S., & Choi, S. (2015). Role of computer-aided drug design in modern drug discovery. *Archives of Pharmacal Research*, 38(9), 1686–1701. <https://doi.org/10.1007/s12272-015-0640-5>
- Manisha, Chauhan, S., Kumar, P., & Kumar, A. (2019). Development of prediction model for fructose-1,6-bisphosphatase inhibitors using the Monte Carlo method. *SAR and QSAR in Environmental Research*, 30(3), 145–159. <https://doi.org/10.1080/1062936X.2019.1568299>
- Martyna, G. J., Klein, M. L., & Tuckerman, M. (1992). Nose–Hoover chains – The canonical ensemble via continuous dynamics. *The Journal of Chemical Physics*, 97(4), 2635–2643. <https://doi.org/10.1063/1.463940>
- Martyna, G. J., Tobias, D. J., & Klein, M. L. (1994). Constant-pressure molecular dynamics algorithms. *The Journal of Chemical Physics*, 101(5), 4177–4189. <https://doi.org/10.1063/1.467468>
- Mishra, V. R., Ghanavatkar, C. W., Mali, S. N., Qureshi, S. I., Chaudhari, H. K., & Sekar, N. (2019). Design, synthesis, antimicrobial activity and computational studies of novel azo linked substituted benzimidazole, benzoxazole and benzothiazole derivatives. *Computational Biology and Chemistry*, 78, 330–337. <https://doi.org/10.1016/j.compbiochem.2019.01.003>
- Murtuja, S., Shaquiquzzaman, M., & Amir, M. (2018). Design, synthesis, and screening of hybrid benzothiazolyl-oxadiazoles as anticonvulsant agents. *Letters in Drug Design & Discovery*, 15(4), 398–405. <https://doi.org/10.2174/1570180814666170526154914>
- Nickavar, B., & Yousefian, N. (2010). Inhibitory effects of six allium species on α -Amylase enzyme activity. *Iranian Journal of Pharmaceutical Research*, 8, 53–57.
- Nimbhal, M., Bagri, K., Kumar, P., & Kumar, A. (2020). The index of ideality of correlation: A statistical yardstick for better QSAR modeling of glucokinase activators. *Structural Chemistry*, 31(2), 831–839. <https://doi.org/10.1007/s11224-019-01468-w>
- Ojha, P. K., & Roy, K. (2011). Comparative QSARs for antimalarial endochins: Importance of descriptor-thinning and noise reduction prior to feature selection. *Chemometrics and Intelligent Laboratory Systems*, 109(2), 146–161. <https://doi.org/10.1016/j.chemolab.2011.08.007>
- Ojha, P. K., & Roy, K. (2018). Development of a robust and validated 2D-QSPR model for sweetness potency of diverse functional organic molecules. *Food and Chemical Toxicology: An International Journal Published for the British Industrial Biological Research Association*, 112, 551–562. <https://doi.org/10.1016/j.fct.2017.03.043>
- Pettersen, E. F., Goddard, T. D., Huang, C. C., Couch, G. S., Greenblatt, D. M., Meng, E. C., & Ferrin, T. E. (2004). UCSF Chimera – A visualization system for exploratory research and analysis. *Journal of Computational Chemistry*, 25(13), 1605–1612. <https://doi.org/10.1002/jcc.20084>
- Roy, K. (2015). Application of chemometrics and cheminformatics in antimalarial drug research. *Combinatorial Chemistry & High Throughput Screening*, 18(2), 89–90. <https://doi.org/10.2174/138620731802150215154014>
- Sharma, A. K., Sharma, R., & Gangwal, A. (2018). Surface tension studies of ternary system: Cu(II) surfactants-2-amino-6-methyl benzothiazole complex plus methanol plus benzene at 311 K. *Current Physical Chemistry*, 8(2), 151–161. <https://doi.org/10.2174/1877946808666180914164134>
- Shi, J., Deng, Q., Li, Y., Zheng, M., Chai, Z., Wan, C., Zheng, Z., Li, L., Huang, F., & Tang, B. (2018). A rapid and ultrasensitive tetraphenylethylene-based probe with aggregation-induced emission for direct detection of α -amylase in human body fluids. *Analytical Chemistry*, 90(22), 13775–13782. <https://doi.org/10.1021/acs.analchem.8b04244>
- Souza, P. M. D., & Magalhães, P. de O. e. (2010). Application of microbial α -amylase in industry – A review. *Brazilian Journal of Microbiology*, 41(4), 850–861. <https://doi.org/10.1590/S1517-83822010000400004>
- Svensson, B. (1994). Protein engineering in the alpha-amylase family: Catalytic mechanism, substrate specificity, and stability. *Plant Molecular Biology*, 25(2), 141–157. <https://doi.org/10.1007/BF00023233>
- Taha, M., Irshad, M., Imran, S., Rahim, F., Selvaraj, M., Almandil, N. B., Mosaddik, A., Chigurupati, S., Nawaz, F., Ismail, N. H., & Ibrahim, M. (2019). Thiazole based carbohydrazide derivatives as α -amylase inhibitor and their molecular docking study. *Heteroatom Chemistry*, 2019, 1–8. <https://doi.org/10.1155/2019/7502347>
- Taha, M., Ismail, N. H., Lalani, S., Fatmi, M. Q., Atia Tul, W., Siddiqui, S., Khan, K. M., Imran, S., & Choudhary, M. I. (2015). Synthesis of novel inhibitors of α -glucosidase based on the benzothiazole skeleton containing benzohydrazide moiety and their molecular docking studies. *European Journal of Medicinal Chemistry*, 92, 387–400. <https://doi.org/10.1016/j.ejmech.2015.01.009>
- Taha, M., Shah, S. A. A., Imran, S., Afifi, M., Chigurupati, S., Selvaraj, M., Rahim, F., Ullah, H., Zaman, K., & Vijayabalan, S. (2017). Synthesis and in vitro study of benzofuran hydrazone derivatives as novel alpha-amylase inhibitor. *Bioorganic Chemistry*, 75, 78–85. <https://doi.org/10.1016/j.bioorg.2017.09.002>
- Thota, S., Rodrigues, D. A., Pinheiro, P. D. S. M., Lima, L. M., Fraga, C. A., & Barreiro, E. J. (2018). N-acylhydrazones as drugs. *Bioorganic & Medicinal Chemistry Letters*, 28(17), 2797–2806. <https://doi.org/10.1016/j.bmcl.2018.07.015>
- Toropova, A. P., & Toropov, A. A. (2017). The index of ideality of correlation: A criterion of predictability of QSAR models for skin permeability? *The Science of the Total Environment*, 586, 466–472. <https://doi.org/10.1016/j.scitotenv.2017.01.198>
- Toropova, A. P., & Toropov, A. A. (2019a). QSPR and nano-QSPR: What is the difference? *Journal of Molecular Structure*, 1182, 141–149. <https://doi.org/10.1016/j.molstruc.2019.01.040>
- Toropova, A. P., & Toropov, A. A. (2019b). Does the index of ideality of correlation detect the better model correctly? *Molecular Informatics*, 38(8–9), 1800157. <https://doi.org/10.1002/minf.201800157>
- Toropova, A. P., & Toropov, A. A. (2019c). The index of ideality of correlation: Improvement of models for toxicity to algae. *Natural Product Research*, 33(15), 2200–2207. <https://doi.org/10.1080/14786419.2018.1493591>
- Toropova, A. P., Toropov, A. A., Beeg, M., Gobbi, M., & Salmona, M. (2017). Utilization of the Monte Carlo method to build up QSAR models for hemolysis and cytotoxicity of antimicrobial peptides. *Current Drug Discovery Technologies*, 14(4), 229–243. <https://doi.org/10.2174/1570163814666170525114128>
- Toropova, A. P., Toropov, A. A., Benfenati, E., Leszczynska, D., & Leszczynski, J. (2015). QSAR model as a random event: A case of rat toxicity. *Bioorganic & Medicinal Chemistry*, 23(6), 1223–1230. <https://doi.org/10.1016/j.bmc.2015.01.055>
- Toropova, A. P., Toropov, A. A., Veselinovic, A. M., Veselinovic, J. B., Leszczynska, D., & Leszczynski, J. (2019). Semi-correlations combined with the index of ideality of correlation: A tool to build up model of mutagenic potential. *Molecular and Cellular Biochemistry*, 452(1–2), 133–140. <https://doi.org/10.1007/s11010-018-3419-4>
- Toropov, A. A., & Toropova, A. P. (2018). Predicting cytotoxicity of 2-phenoxyindole derivatives against breast cancer cells using index of ideality of correlation. *Anticancer Research*, 38(11), 6189–6194. <https://doi.org/10.21873/anticancres.12972>
- Toropov, A. A., & Toropova, A. P. (2019). Use of the index of ideality of correlation to improve predictive potential for biochemical endpoints. *Toxicology Mechanisms and Methods*, 29(1), 43–52. <https://doi.org/10.1080/15376516.2018.1506851>
- Toropov, A. A., Toropova, A. P., & Benfenati, E. (2019). The index of ideality of correlation: QSAR model of acute toxicity for Zebrafish (*Danio rerio*) embryo. *International Journal of Environmental Research*, 13(2), 387–394. <https://doi.org/10.1007/s41742-019-00183-y>
- Trott, O., & Olson, A. J. (2010). AutoDock Vina: Improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. *Journal of Computational Chemistry*, 31(2), 455–461. <https://doi.org/10.1002/jcc.21334>
- Version, M., & 6.2.2. (2014). *calculation module developed by ChemAxon*. <http://www.chemaxon.com/products/marvin/marvinsketch/>
- Veselinovic, A. M., Milosavljevic, J. B., Toropov, A. A., & Nikolic, G. M. (2013). SMILES-based QSAR model for arylpiperazines as high-affinity 5-HT(1A) receptor ligands using CORAL. *European Journal of Pharmaceutical Sciences: Official Journal of the European Federation for Pharmaceutical Sciences*, 48(3), 532–541. <https://doi.org/10.1016/j.ejps.2012.12.021>
- Veselinovic, J. B., Nikolic, G. M., Trutic, N. V., Zivkovic, J. V., & Veselinovic, A. M. (2015). Monte Carlo QSAR models for predicting organophosphate inhibition of acetylcholinesterase. *SAR and QSAR in Environmental Research*, 26(6), 449–460. <https://doi.org/10.1080/1062936x.2015.1049665>
- Wang, J., Wang, W., Kollman, P. A., & Case, D. A. (2006). Automatic atom type and bond type perception in molecular mechanical calculations. *Journal of Molecular Graphics & Modelling*, 25(2), 247–260. <https://doi.org/10.1016/j.jmgm.2005.12.005>



CORAL: QSAR models of CB1 cannabinoid receptor inhibitors based on local and global SMILES attributes with the index of ideality of correlation and the correlation contradiction index

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ABSTRACT

Obesity has acquired notable attention due to its high occurrence and link with grievous health problems such as hypertension, diabetes and heart disease. It has been reported that the endocannabinoid system executes a pivotal part in the management of food absorption, fat augmentation, and energy balance. In the present manuscript, we report a detailed QSAR analysis for 165 CB1 cannabinoid receptor inhibitors employing the Monte Carlo optimization process incorporated within the CORAL software. Eight splits are made from the whole dataset and sixteen QSAR models are developed from these splits employing two target function TF₁ (without index of ideality of correlation) and TF₂ (with index of ideality of correlation). All the QSAR models developed with TF₂ have better predictive potential than the models developed with TF₁. The model built for split 5 using TF₂ is the leading model due to the higher value of the determination coefficient of the validation set ($R^2_{Valid} = 0.8518$). The index of ideality of correlation (IIC) improves the statistical performance of CORAL-based QSAR-models and gives statistically robust predictive models of the investigated endpoint pIC₅₀. In the present manuscript, a novel criterion "Correlation Contradiction Index (CCI)" is also applied to know its predictive potential. The absolute value of CCI for calibration set is less when QSAR models are developed employing IIC. The promoters of increase and decrease endpoint pIC₅₀ are identified and these are applied to design seven new compounds. All the newly designed molecule were docked into in the active site of human cannabinoid receptor CB1 (PDB ID: 5tgz).

1. Introduction

Obesity is a worldwide health problem. In 2016, about 74 million boys and 50 million girls were overweight and obesity has greatly increased in recent decades [1]. Obesity has acquired notable attention due to its high occurrence and link with grievous health problems such as hypertension, diabetes and heart disease [2–4]. Food and Drug Administration (FDA) has approved the only one medicine, Orlistat, to treat the obesity in children and adolescents. Whereas in adults, the FDA has recommended six drugs i.e. Orlistat, Phentermine, Bupropion, Lorcaserin and Liraglutide for obesity treatment. The therapeutic potential of these drugs has been limited due to their unwanted side effects and variable efficacy [5,6]. Therefore, the novel target development for the anti-obesity drug is still a challenge to the medicinal chemist [7,8]. Literature survey reveals that the endocannabinoid system gave a new platform for anti-obesity drug development [9–13]. It has been reported

that the endocannabinoid system executes a pivotal role in the management of food absorption, fat augmentation, and energy balance [14]. The hyperactivation of this endogenous signalling system appears to be firmly associated with abdominal obesity and the progress of the metabolic syndrome. The two cannabinoid receptors subtypes, CB1 and CB2, are a member of the G-protein coupled seven-transmembrane-spanning receptor family (GPCRs) [15,16]. The CB1 and CB2 receptor are expressed in the central nervous system & peripheral tissues and immune system, respectively. The CB1 receptor is presumed to be associated in the management of perception, motor activity, sensation, idea, understanding through thought, the senses, intuition resulting from the process of cognition and the suppression of transmitter release through its coupling to ions channels [17,18]. The antagonism of CB1 receptor has been discovered by the scientific community as an alluring target for obesity therapy because the specific blockage of this receptor can induce bodyweight reduction [14,15,17].

Quantitative structure-activity relationship (QSAR) is an important

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List of abbreviation

CB1	Cannabinoid Receptor-1
CCC	Concordance Correlation Coefficient
CORAL:	CORrelations And Logic
IIC	Index of Ideality Correlation
MAE	Mean Absolute Error
QSAR	Quantitative Structure Activity Relationship
SMILES	Simplified Input-Line Entry System
CCI	Correlation Contradiction Index
TF	Target fuction

a target function. This method is applied by the different research group for a large number of various physicochemical, biochemical, ecological, and medicinal endpoints [19,23–25]. In the last few years, the index of ideality correlation (IIC) has been calculated to judge the predictive potential of built QSAR model and it helps to predict the better model in place of various statistical parameters such as Q^2 , Q^2F_1 , Q^2F_2 , Q^2F_3 etc [19,28–31].

In continuation to our work on QSAR [32–34] and synthesis of pharmacological active heterocyclic compounds [35–42], we herein publicize the CORAL based QSAR-models for 165 CB1 cannabinoid receptor inhibitors. In this manuscript, two new criteria Index of ideality correlation (IIC) and Correlation Contradiction Index (CCI) is also studied.

2. Materials and method

2.1. Dataset

A total of 165 CB1 cannabinoid receptor inhibitors were taken from the database <https://www.ebi.ac.uk/chembl/old/> (ChEMBL ID are given in Supporting information) and same experimental condition was applied to measure the IC_{50} of CB1 receptor [6,17,43,44]. All IC_{50} (nM) values for CB1 cannabinoid receptor were converted into the corresponding pIC_{50} ($-\log IC_{50}$) and it is taken as a dependent factor for the building of QSAR models. Eight splits were made from the whole dataset and each split was

part of computer-assisted drug design (CADD) and it has been effectively applied for designing of new lead molecules [19–22]. QSAR correlates the endpoints with the structural features of the molecules. Especially, the CORAL software (<http://www.insilico.eu/coral>) based on the Monte Carlo algorithm has been extensively used for designing the QSAR models [19,20,23–27]. In this software, the molecular attributes obtained from simplified input-line entry system (SMILES) and molecular graph have been applied to develop QSAR models. The molecular descriptor in terms of correlation weight (CW) has been calculated by CORAL software and the numerical value of CW gives a maximal value of

Table 1
Percentage of the identity of splits 1–8 CB1 receptor inhibitors.

Split	SET	Split 1 (%)	Split 2 (%)	Split 3 (%)	Split 4 (%)	Split 5 (%)	Split 6 (%)	Split 7 (%)	Split 8 (%)
Split 1	Total	100	0.0	0.0	0.0	25.5	24.8	24.2	25.5
	Training	100	0.0	0.0	0.0	24.4	26.5	24.4	24.4
	Invisible training	100	0.0	0.0	0.0	26.2	24.1	24.1	26.5
	Calibration	100	0.0	0.0	0.0	26.5	24.1	23.8	26.5
	Validation	100	0.0	0.0	0.0	24.7	24.7	24.7	24.4
Split 2	Total	100	0.0	0.0	24.8	25.5	25.5	24.2	
	Training	100	0.0	0.0	24.1	26.2	26.5	24.1	
	Invisible training	100	0.0	0.0	26.5	24.4	24.4	24.4	
	Calibration	100	0.0	0.0	24.7	24.7	24.4	24.7	
	Validation	100	0.0	0.0	24.1	26.5	26.5	23.8	
Split 3	Total		100	0.0	24.2	25.5	25.5	24.8	
	Training		100	0.0	24.7	24.4	24.7	24.7	
	Invisible training		100	0.0	23.8	26.5	26.5	24.1	
	Calibration		100	0.0	24.1	26.5	26.2	24.1	
	Validation		100	0.0	24.4	24.4	24.4	26.5	
Split 4	Total			100	25.5	24.2	24.8	25.5	
	Training			100	26.5	23.8	24.1	26.5	
	Invisible training			100	24.4	24.7	24.7	24.7	
	Calibration			100	24.4	24.4	26.5	24.4	
	Validation			100	26.5	24.1	24.1	26.2	
Split 5	Total				100	0.0	0.0	0.0	
	Training				100	0.0	0.0	0.0	
	Invisible training				100	0.0	0.0	0.0	
	Calibration				100	0.0	0.0	0.0	
	Validation				100	0.0	0.0	0.0	
Split 6	Total					100	0.0	0.0	
	Training					100	0.0	0.0	
	Invisible training					100	0.0	0.0	
	Calibration					100	0.0	0.0	
	Validation					100	0.0	0.0	
Split 7	Total						100	0.0	
	Training						100	0.0	
	Invisible training						100	0.0	
	Calibration						100	0.0	
	Validation						100	0.0	
Split 8	Total							100	
	Training							100	
	Invisible training							100	
	Calibration							100	
	Validation							100	

& editing.

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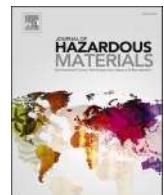
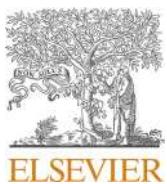
Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.chemolab.2020.103982>.

References

- [1] F. Rossi, F. Punzo, G. Umano, M. Argenziano, E. Miraglia Del Giudice, Role of cannabinoids in obesity, *Int. J. Mol. Sci.* 19 (2018) 2690.
- [2] A. Tremblay, E. Doucet, Obesity: a disease or a biological adaptation? *Obes. Rev.* 1 (2000) 27–35.
- [3] G.A. Bray, Obesity: the disease, *J. Med. Chem.* 49 (2006) 4001–4007.
- [4] P.N. Patel, R. Pathak, Rimonabant: a novel selective cannabinoid-1 receptor antagonist for treatment of obesity, *Am. J. Health Syst. Pharm.* 64 (2007) 481–489.
- [5] D.R. Hou, S. Alam, T.C. Kuan, M. Ramanathan, T.P. Lin, M.S. Hung, 1,2,3-Triazole derivatives as new cannabinoid CB1 receptor antagonists, *Bioorg. Med. Chem. Lett.* 19 (2009) 1022–1025.
- [6] H.J. Seo, M.J. Kim, S.H. Lee, S.-H. Lee, M.E. Jung, M.-S. Kim, K. Ahn, J. Kim, J. Lee, Synthesis and structure-activity relationship of 1,2,4-triazole-containing diarylpyrazolyl carboxamide as CB1 cannabinoid receptor-ligand, *Bioorg. Med. Chem. Lett.* 18 (2010) 1149–1162.
- [7] K.A. Horton, A.V. Goonawardena, J. Sesay, A.C. Howlett, R.E. Hampson, Systemic blockade of the CB1 receptor augments hippocampal gene expression involved in synaptic plasticity but perturbs hippocampus-dependent learning task, *Cannabis Cannabinoid Res.* 4 (2019) 33–41.
- [8] S.D. Banister, K. Krishna Kumar, V. Kumar, B.K. Kobilka, S.V. Malhotra, Selective modulation of the cannabinoid type 1 (CB1) receptor as an emerging platform for the treatment of neuropathic pain, *Medchemcomm* 10 (2019) 647–659.
- [9] K. Yoshida, Y. Kita, S.M. Tokuoka, F. Hamano, M. Yamazaki, K. Sakimura, M. Kano, T. Shimizu, Monoacylglycerol lipase deficiency affects diet-induced obesity, fat absorption, and feeding behavior in CB1 cannabinoid receptor-deficient mice, *Faseb. J.* 33 (2019) 2484–2497.
- [10] X. Xu, S. Jiang, E. Xu, X. Wu, R. Zhao, Inhibition of CB1 receptor ameliorates spatial learning and memory impairment in mice with traumatic brain injury, *Neurosci. Lett.* 696 (2019) 127–131.
- [11] M.C.G. Leite-Avalca, F.T. Staats, D. Verona, P. de Souza, M.C. Almeida, J.E. Silva-Santos, A.R. Zampronio, Cannabinoid CB1 receptor antagonist rimonabant decreases levels of markers of organ dysfunction and alters vascular reactivity in aortic vessels in late sepsis in rats, *Inflammation* 42 (2019) 618–627.
- [12] J.L.C. Lee, F.E. Amorim, L.F. Cassini, O.B. Amaral, Different temporal windows for CB1 receptor involvement in contextual fear memory destabilisation in the amygdala and hippocampus, *PloS One* 14 (2019), e0205781.
- [13] H. Laurikainen, L. Tuominen, M. Tikka, H. Merisaari, R.L. Armio, E. Sormunen, F. Borgan, M. Veronese, O. Howes, M. Haaparanta-Solin, O. Solin, J. Hietala, M. group, Sex difference in brain CB1 receptor availability in man, *Neuroimage* 184 (2019) 834–842.
- [14] B.K. Srivastava, R. Soni, A. Joharapurkar, K.V. Sairam, J.Z. Patel, A. Goswami, S.A. Shedge, S.S. Kar, R.P. Salunke, S.B. Gugale, A. Dhawas, P. Kadam, B. Mishra, N. Sadhwani, V.B. Unadkat, P. Mitra, M.R. Jain, P.R. Patel, Bioisosteric replacement of dihydropyrazole of 4S-(*4*-chlorophenyl)-N-methyl-N’-[*(4*-chlorophenyl)-sulfonyl]-4-phenyl-4,5-di hydro-1H-pyrazole-1-caboxamide (SLV-319) a potent CB1 receptor antagonist by imidazole and oxazole, *Bioorg. Med. Chem. Lett.* 18 (2008) 963–968.
- [15] C. Montero, N.E. Campillo, P. Goya, J.A. Páez, Homology models of the cannabinoid CB1 and CB2 receptors. A docking analysis study, *Eur. J. Med. Chem.* 40 (2005) 75–83.
- [16] C.-J. Qiao, H.I. Ali, K.H. Ahn, S. Kolluru, D.A. Kendall, D. Lu, Synthesis and biological evaluation of indole-2-carboxamides bearing photoactivatable functionalities as novel allosteric modulators for the cannabinoid CB1 receptor, *Eur. J. Med. Chem.* 121 (2016) 517–529.
- [17] J. Lee, H.J. Seo, S.H. Lee, J. Kim, M.E. Jung, S.-H. Lee, K.-S. Song, J. Lee, S.Y. Kang, M.J. Kim, M.-S. Kim, E.-J. Son, M. Lee, H.-K. Han, Discovery of 2-(4-(1*H*-1,2,4-triazol-1-yl)methyl)-5-(4-bromophenyl)-1-(2-chlorophenyl)-1H-pyrazol-3-yl)-5-tert-butyl-1,3,4-thiadiazole (GCC2680) as a potent, selective and orally efficacious cannabinoid-1 receptor antagonist, *Bioorg. Med. Chem.* 18 (2010) 6377–6388.
- [18] M.K. Sharma, P.R. Murunkar, A.M. Kanhed, R. Giridhar, M.R. Yadav, Prospective therapeutic agents for obesity: molecular modification approaches of centrally and peripherally acting selective cannabinoid 1 receptor antagonists, *Eur. J. Med. Chem.* 79 (2014) 298–339.
- [19] P. Kumar, A. Kumar, J. Sindhu, Design and development of novel focal adhesion kinase (FAK) inhibitors using Monte Carlo method with index of ideality of correlation to validate QSAR, *SAR QSAR Environ. Res.* 30 (2019) 63–80.
- [20] P. Kumar, A. Kumar, J. Sindhu, S. Lal, QSAR models for nitrogen containing monophosphonate and bisphosphonate derivatives as human farnesy
- [21] A. Kumar, E. Rathi, S.G. Kini, Identification of potential tumour-associated carbonic anhydrase isozyme IX inhibitors: atom-based 3D-QSAR modelling, pharmacophore-based virtual screening and molecular docking studies, *J. Biomol. Struct. Dyn.* (2019), <https://doi.org/10.1080/07391102.2019.1626285>.
- [22] M. Marzo, G.J. Lavado, F. Como, A.P. Toropova, A.A. Toropov, D. Baderna, C. Cappelli, A. Lombardo, C. Toma, M. Blázquez, E. Benfenati, QSAR models for biocides: the example of the prediction of *Daphnia magna* acute toxicity, *SAR QSAR Environ. Res.* 31 (2020) 227–243.
- [23] A.A. Toropov, A.P. Toropova, The Monte Carlo Method as a tool to build up predictive QSAR/QSAR, *Curr. Comput. Aided Drug Des.* (2019), <https://doi.org/10.2174/1573409915666190328123112>.
- [24] A.P. Toropova, A.A. Toropov, CORAL: QSAR models for carcinogenicity of organic compounds for male and female rats, *Comput. Biol. Chem.* 72 (2018) 26–32.
- [25] P. Kumar, A. Kumar, Monte Carlo method based QSAR studies of mer kinase inhibitors in compliance with OECD principles, *Drug Res.* 68 (2018) 189–195.
- [26] A.P. Toropova, A.A. Toropov, E. Carnesecchi, E. Benfenati, J.L. Dorne, The index of ideality of correlation: models for flammability of binary liquid mixtures, *Chem. Pap.* 74 (2020) 601–609.
- [27] S. Jain, S.A. Amin, N. Adhikari, T. Jha, S. Gayen, Good and bad molecular fingerprints for human rhinovirus 3C protease inhibition: identification, validation, and application in designing of new inhibitors through Monte Carlo-based QSAR study, *J. Biomol. Struct. Dyn.* 38 (2020) 66–77.
- [28] A.P. Toropova, A.A. Toropov, A.M. Veselinovic, J.B. Veselinovic, D. Leszczynska, J. Leszczynski, Semi-correlations combined with the index of ideality of correlation: a tool to build up model of mutagenic potential, *Mol. Cell. Biochem.* 452 (2019) 133–140.
- [29] A.P. Toropova, A.A. Toropov, The index of ideality of correlation: improvement of models for toxicity to algae, *Nat. Prod. Res.* 33 (15) (2019) 2200–2207, <https://doi.org/10.1080/14786419.2018.1493591>.
- [30] A.A. Toropov, A.P. Toropova, Predicting cytotoxicity of 2-phenylindole derivatives against breast cancer cells using index of ideality of correlation, *Anticancer Res.* 38 (2018) 6189–6194.
- [31] S. Ahmadi, Mathematical modeling of cytotoxicity of metal oxide nanoparticles using the index of ideality correlation criteria, *Chemosphere* 242 (2020) 125192.
- [32] P. Kumar, A. Kumar, J. Sindhu, In silico design of diacylglycerol acyltransferase-1 (DGAT1) inhibitors based on SMILES descriptors using Monte-Carlo method, *SAR QSAR Environ. Res.* 30 (2019) 525–541, <https://doi.org/10.1080/1062936X.2019.1629998>.
- [33] Manisha, S. Chauhan, P. Kumar, A. Kumar, Development of prediction model for fructose-1,6- bisphosphatase inhibitors using the Monte Carlo method, *SAR QSAR Environ. Res.* 30 (2019) 145–159.
- [34] A. Kumar, S. Singh, S. Jain, P. Kumar, Synthesis, antimicrobial evaluation, QSAR and in Silico ADMET studies of decanoic acid derivatives, *Acta Pol. Pharm.* 68 (2011) 191–204.
- [35] P. Kumar, M. Duhan, K. Kadyan, J.K. Bhardwaj, P. Saraf, M. Mittal, Multicomponent synthesis of some molecular hybrid containing thiazole pyrazole as apoptosis inducer, *Drug Res.* 68 (2018) 72–79.
- [36] P. Kumar, K. Kadyan, M. Duhan, J. Sindhu, V. Singh, B.S. Saharan, Design, synthesis, conformational and molecular docking study of some novel acyl hydrazone based molecular hybrids as antimalarial and antimicrobial agents, *Chem. Cent. J.* 11 (2017) 115.
- [37] P. Kumar, M. Duhan, K. Kadyan, J. Sindhu, S. Kumar, H. Sharma, Synthesis of novel inhibitors of α -amylase based on thiazolidine-4-one skeleton containing pyrazole moiety and their configurational studies, *MedChemComm* 8 (2017) 1468–1476, <https://doi.org/10.1039/C7MD00080D>.
- [38] P. Kumar, R. Bhatia, R. Khanna, A. Dalal, D. Kumar, P. Surain, R.C. Kamboj, Synthesis of some benzothiazoles by developing a new protocol using urea nitrate as a catalyst and their antimicrobial activities, *J. Sulfur Chem.* 38 (2017) 585–596.
- [39] P. Kumar, R. Bhatia, D. Kumar, R.C. Kamboj, S. Kumar, R. Kamal, R. Kumar, An economic, simple and convenient synthesis of 2-aryl/heteroaryl/styryl/alkylbenzothiazoles using SiO 2-HNO 3, *Res. Chem. Intermed.* 41 (2015) 4283–4292.
- [40] S. Kumar, S. Kumar, P. Kumar, Synthesis and antimicrobial activity of some (3-phenyl-5-(1-phenyl-3-aryl-1H-pyrazol-4-yl)-4-, 5-dihydro-1H-pyrazol-1-yl)(pyridin-4-yl) methanones: new derivatives of 1, 3, 5-trisubstituted pyrazolines, *Med. Chem. Res.* 22 (2013) 433–439.
- [41] P. Kumar, A. Kumar, J. Makrandi, Synthesis and evaluation of bioactivity of thiazolo [3, 2-b]-[1, 2, 4]-triazoles and isomeric thiazolo [2, 3-c]-[1, 2, 4]-triazoles, *J. Heterocycl. Chem.* 50 (2013) 1223–1229.
- [42] P. Kumar, A. Kuamr, L.J. Mohan, J. Makrandi, Heterocyclic systems containing bridgehead nitrogen atom: synthesis and evaluation of biological activity of imidazo [2, 1-b]-1, 3, 4-thiadiazol [2, 3-c]-triazoles, s-triazolo [3, 4-b]-1, 3, 4-thiadiazolo [3, 2-b]-imidazo [4, 5-b] quinoxaline and bis-(s-Triazolo [3, 4-b]-1, 3, 4-thiadiazolo [3, 2-b]-imidazo [4, 5-b]-cyclohexane)-5a, 6a-diene, *Bull. Kor. Chem. Soc.* 31 (2010) 3304–3308.
- [43] K.-S. Song, M.J. Kim, H.J. Seo, S.-H. Lee, M.E. Jung, S.-U. Kim, J. Kim, J. Lee, Synthesis and structure-activity relationship of novel diarylpyrazole imide analogues as CB1 cannabinoid receptor ligands, *Bioorg. Med. Chem.* 17 (2009) 3080–3092.
- [44] N. Khan, S.A. Halim, W. Khan, S.K. Zafar, Z. Ul-Haq, In-silico designing and characterization of binding modes of two novel inhibitors for CB1 receptor against obesity by classical 3D-QSAR approach, *J. Mol. Graph. Model.* 89 (2019) 199–214.
- [45] A.A. Toropov, A.P. Toropova, E. Benfenati, Additive SMILES-based carcinogenicity models: probabilistic principles in the search for robust predictions, *Int. J. Mol. Sci.* 10 (2009) 3106–3127.

- [46] A.P. Toropova, A.A. Toropov, D. Leszczynska, J. Leszczynski, CORAL and Nano-QFAR: quantitative feature - activity relationships (QFAR) for bioavailability of nanoparticles (ZnO, CuO, Co₃O₄, and TiO₂), Ecotoxicol. Environ. Saf. 139 (2017) 404–407.
- [47] P.G.R. Achary, A.P. Toropova, A.A. Toropov, Combinations of graph invariants and attributes of simplified molecular input-line entry system (SMILES) to build up models for sweetness, Food Res. Int. 122 (2019) 40–46.
- [48] M. Duhan, R. Singh, M. Devi, J. Sindhu, R. Bhatia, A. Kumar, P. Kumar, Synthesis, molecular docking and QSAR study of thiazole clubbed pyrazole hybrid as α -amylase inhibitor, J. Biomol. Struct. Dyn. (2019), <https://doi.org/10.1080/07391102.2019.17>.
- [49] A.P. Toropova, A.A. Toropov, QSPR and nano-QSPR: what is the difference? J. Mol. Struct. 1182 (2019) 141–149.
- [50] A.P. Toropova, A.A. Toropov, The index of ideality of correlation: a criterion of predictability of QSAR models for skin permeability? Sci. Total Environ. 586 (2017) 466–472.
- [51] A.A. Toropov, A.P. Toropova, Use of the index of ideality of correlation to improve predictive potential for biochemical endpoints, Toxicol. Mech. Methods 29 (2019) 43–52.
- [52] A.A. Toropov, A.P. Toropova, The Correlation Contradictions Index (CCI): building up reliable models of mutagenic potential of silver nanoparticles under different conditions using quasi-SMILES, Sci. Total Environ. 681 (2019) 102–109.
- [53] R.B. Aher, K. Roy, Exploring the structural requirements in multiple chemical scaffolds for the selective inhibition of Plasmodium falciparum calcium-dependent protein kinase-1 (PfCDPK-1) by 3D-pharmacophore modelling, and docking studies, SAR QSAR Environ. Res. 28 (2017) 390–414.
- [54] S.S. Bhayye, K. Roy, A. Saha, Pharmacophore generation, atom-based 3D-QSAR, HQSAR and activity cliff analyses of benzothiazine and deazaxanthine derivatives as dual A2A antagonists/MAOB inhibitors., SAR QSAR Environ. Res. 27 (3) (2016) 183–202, <https://doi.org/10.1080/1062936X.2015.1136840>.
- [55] A. Golbraikh, A. Tropsha, Beware of q2!, J. Mol. Graph. Model. 20 (2002) 269–276.
- [56] L.M. Shi, H. Fang, W. Tong, J. Wu, R. Perkins, R.M. Blair, W.S. Branham, S.L. Dial, C.L. Moland, D.M. Sheehan, QSAR models using a large diverse set of estrogens, J. Chem. Inf. Comput. Sci. 41 (2001) 186–195.
- [57] G. Schüürmann, R.-U. Ebert, J. Chen, B. Wang, R. Kühne, External validation and prediction employing the predictive squared correlation coefficient — test set activity mean vs training set activity mean, J. Chem. Inf. Model. 48 (2008) 2140–2145.
- [58] P. Pratim Roy, S. Paul, I. Mitra, K. Roy, On two novel parameters for validation of predictive QSAR models, Molecules 14 (2009) 1660–1701.
- [59] K. Roy, P. Chakraborty, I. Mitra, P.K. Ojha, S. Kar, R.N. Das, Some case studies on application of “rm2” metrics for judging quality of quantitative structure-activity relationship predictions: emphasis on scaling of response data, J. Comput. Chem. 34 (2013) 1071–1082.
- [60] P. Gramatica, A. Sangion, A historical excursus on the statistical validation parameters for QSAR models: a clarification concerning metrics and terminology, J. Chem. Inf. Model. 56 (2016) 1127–1131.
- [61] P. Kumar, A. Kumar, Nucleobase sequence based building up of reliable QSAR models with the index of ideality correlation using Monte Carlo method, J. Biomol. Struct. Dyn. (2019), <https://doi.org/10.1080/07391102.2019.1656109>.
- [62] P. Kumar, M. Nimbhal, K. Bagri, P. Kumar, The index of ideality of correlation: A statistical yardstick for better QSAR modeling of glucokinase activators, Struct. Chem. (2019), <https://doi.org/10.1007/s11224-019-01468-w>.
- [63] P.K. Ojha, I. Mitra, R.N. Das, K. Roy, Further exploring rm2 metrics for validation of QSAR models, Chemometr. Intell. Lab. 107 (2011) 194–205.
- [64] A. Ruscifina, G. Floresta, A. Marrazzo, C. Parenti, O. Prezzavento, G. Nastasi, M. Dichiara, E. Amata, Development of a Sigma-2 Receptor affinity filter through a Monte Carlo based QSAR analysis, Eur. J. Pharmaceut. Sci. 106 (2017) 94–101.
- [65] C.A. Lipinski, F. Lombardo, B.W. Dominy, P.J. Feeney, Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings¹ of original article: S0169-409X(96)00423-1. The article was originally published in Advanced Drug Delivery Reviews 23 (1997) 3–25.1, Adv. Drug Deliv. Rev. 46 (2001) 3–26.
- [66] D.A. Griffith, J.R. Hadcock, S.C. Black, P.A. Iredale, P.A. Carpino, P. DaSilva-Jardine, R. Day, J. DiBrino, R.L. Dow, M.S. Landis, R.E. O'Connor, D.O. Scott, Discovery of 1-[9-(4-Chlorophenyl)-8-(2-chlorophenyl)-9H-purin-6-yl]-4-ethylaminopiperidine-4-carboxylic acid amide hydrochloride (CP-945,598), a novel, potent, and selective cannabinoid type 1 receptor antagonist, J. Med. Chem. 52 (2009) 234–237.
- [67] L.E. Klumpers, C. Roy, G. Ferron, S. Turpault, F. Poitiers, J.-L. Pinquier, J.G.C. van Hasselt, L. Zuurman, F.A.S. Erwic, J.M.A. van Gerven, Surinabant, a selective cannabinoid receptor type 1 antagonist, inhibits Δ^9 -tetrahydrocannabinol-induced central nervous system and heart rate effects in humans, Br. J. Clin. Pharmacol. 76 (2013) 65–77.
- [68] R.J. Chorvat, J. Berbaum, K. Seriacki, J.F. McElroy, JD-5006 and JD-5037: peripherally restricted (PR) cannabinoid-1 receptor blockers related to SLV-319 (Ibipinabant) as metabolic disorder therapeutics devoid of CNS liabilities, Bioorg. Med. Chem. Lett 22 (2012) 6173–6180.
- [69] O. Trott, A.J. Olson, AutoDock Vina, Improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading, J. Comput. Chem. 31 (2010) 455–461.
- [70] T. Hua, K. Vemuri, M. Pu, L. Qu, G.W. Han, Y. Wu, S. Zhao, W. Shui, S. Li, A. Korde, R.B. Laprairie, E.L. Stahl, J.H. Ho, N. Zvonok, H. Zhou, I. Kufareva, B. Wu, Q. Zhao, M.A. Hanson, L.M. Bohn, A. Makriyannis, R.C. Stevens, Z.J. Liu, Crystal structure of the human cannabinoid receptor CB1, Cell 167 (2016) 750–762, <https://doi.org/10.1016/j.cell.2016.10.004>, e14.



Cytotoxicity of quantum dots: Use of quasiSMILES in development of reliable models with index of ideality of correlation and the consensus modelling

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ABSTRACT

The assessment of cytotoxicity of quantum dots is very essential for environmental and health risk analysis. In the present work we have modelled HeLa cell cytotoxicity of sixty one CdSe quantum dots with ZnS shell as a function of its experimental conditions and molecular construction using quasiSMILES representations. The index of ideality of correlation helps in the building of ten statistically significant models having good fitting ability with value of R^2 ranging from 0.8414 to 0.9609 for the training set. The split 5 model is rated as the best model with values of R^2 , Q^2_{F1} , Q^2_{F2} and Q^2_{F3} as 0.8964, 0.8267, 0.8264 and 0.8777 respectively for the calibration set. The extraction of features causing increase and decrease of cytotoxicity of quantum dots indicates importance of neutral surface charge, surface modified with protein, 72 h exposure time, combination of MTT assay with surface protein in decreasing the cytotoxicity. Amphiphilic polymer, polyol ligand with neutral charge, 0.5 – 0.6 nm quantum dot diameter with lipid ligand and unmodified positively charged surface are grouped in toxicity enhancer features. Further, consensus modelling using split 5 and 8 patterns enhances the prediction quality by increasing the R^2_{val} to 0.9361 and 0.9656 respectively.

1. Introduction

Quantum Dots (QDs) have wide range of applications in various fields like quantum computing, solar cells, laser diodes, LEDs, transistors, displays and medical imaging (Choi et al., 2018; Imamoglu, 2003; Jin et al., 2011; Kahmann et al., 2020; Reithmaier and Forchel, 2003; Sakho and Oluwafemi, 2019). Extensive work is going on to make these attractive nanomaterials applicable for pharmaceutical and medical purposes because of their potential features of greater quantum yield, broad excitation, narrow emissions and superb photostability (Bajwa et al., 2016; Reshma and Mohanan, 2019). However the increasing opportunities in biological applications for quantum dots have introduced significant concerns with respect to their toxicological effects (Hu et al., 2017).

Many reports show that many QDs are cytotoxic in nature (Shiohara et al., 2004; Rozenzhak et al., 2005; Deka et al., 2009; Lee et al., 2010; Zhang et al., 2010; Bakalova et al., 2011; Chahal et al., 2012; Yeh et al., 2013) and causes inhibition of cell development, mitochondrial dysfunction, DNA destruction, and apoptosis (Nikazar et al., 2020). The

cytotoxicity is influenced by the chemical and physical properties of QDs like size, surface ligand, charge, concentration etc (Oh et al., 2016). Therefore determination of cytotoxicity of QDs is very important. To check the toxicity in vitro and in vivo is expensive and also much harmful for the animals which are being employed for testing (Erhirie et al., 2018) but there exist no other method to easily assess the cytotoxicity of QDs till date.

Computational methods enjoy the top positions in prior determination of toxicity of chemicals (Raies and Bajic, 2016) and among these methods, quantitative structure activity relationship (QSAR) techniques are of paramount importance. In this process the endpoint of any series of structures is correlated with the chemical and physicochemical characteristics of the compounds via mathematical functions (Buglak et al., 2019; Basant et al., 2016). The era of QSAR started with the use of graph based descriptors as independent variables (Bonchev et al., 1980; Randić, 2001). Later on simplified molecular input-line entry system (SMILES) notations of the molecules has been frequently used in the creation of robust and predictive QSAR models for many endpoints including physical, chemical and biological endpoints through the

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illustrates the comparison of MAE (95 %) for individual and three consensus models for validation sets of both splits. All these outcomes suggest that the consensus model can be used for better prediction of the cytotoxicity of the quantum dots of cadmium.

4. Conclusion

This study was designed to develop the quantitative feature toxicity relationship models for the prediction of cytotoxicity of sixty one CdSe quantum dots using quasiSMILES representations. The increase in the weight of index of ideality of correlation from 0.0 to 0.2 resulted in formation of statistically reliable models with high prediction capability. Model generated with split 5 was most successful in calibration set prediction with value of prediction quality parameter Q^2_{F1} as 0.8267. Extraction of cytotoxicity modulating features pointed towards the involvement of 0.6–0.7 nm QDs diameter and its combination with polyol ligand, neutral unmodified surface, WST assay with 72 h exposure time in reducing the cytotoxicity while unmodified surface and positive surface charge with lipid were favourable for cytotoxic effects of quantum dots. Original consensus models made from ten individual models using split 5 and 8 patterns were more predictive for validation sets and the value of determination coefficient for validation sets increased to 0.9361 and 0.9656 respectively. The mean absolute error (95 %) was also reduced to 0.0815 for split 8 validation set. The resultant consensus models also possessed wider applicability domain containing all the calibration and validation set objects and can be used for prediction of cytotoxicity of QDs with reliability. This combination of index of ideality of correlation and consensus modelling can be applied for predictive and accurate modelling of different endpoints related with nano as well as other materials.

CRediT authorship contribution statement

Ashwani Kumar: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Project administration, Resources, Software, Writing - original draft, Writing - review & editing. **Parvin Kumar:** Resources, Supervision, Validation, Visualization, Writing - review & editing.

Declaration of Competing Interest

The authors report no declarations of interest.

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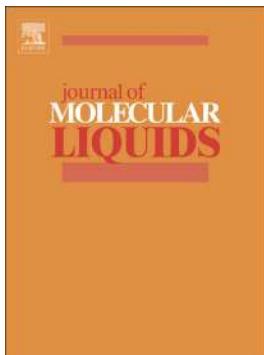
Appendix A. Supplementary data

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References

- Ahmadi, S., 2020. Mathematical modeling of cytotoxicity of metal oxide nanoparticles using the index of ideality correlation criteria. Chemosphere 242, 125192. <https://doi.org/10.1016/j.chemosphere.2019.125192>.
- Bajwa, N., Mehra, N.K., Jain, K., Jain, N.K., 2016. Pharmaceutical and biomedical applications of quantum dots. Artif. Cells Nanomed. Biotechnol. 44 (3), 758–768. <https://doi.org/10.3109/21691401.2015.1052468>.
- Bakalova, R., Zhelev, Z., Aoki, I., Masamoto, K., Mileva, M., Obata, T., Higuchi, M., Gadjeva, V., Kanno, I., 2008. Multimodal silica-shelled quantum dots: direct intracellular delivery, photosensitization, toxic, and microcirculation effects. Bioconjug. Chem. 19 (6), 1135–1142. <https://doi.org/10.1021/bc700431c>.
- Bakalova, R., Zhelev, Z., Kokuryo, D., Spasov, L., Aoki, I., Saga, T., 2011. Chemical nature and structure of organic coating of quantum dots is crucial for their application in imaging diagnostics. Int. J. Nanomed. 6, 1719–1732. <https://doi.org/10.2147/IJN.S17995>.
- Basant, N., Gupta, S., Singh, K.P., 2016. QSAR modeling for predicting reproductive toxicity of chemicals in rats for regulatory purposes. Toxicol. Res. 5 (4), 1029–1038. <https://doi.org/10.1039/c6tx00083e>.
- Boeneman, K., Delehaney, J.B., Blanco-Canosa, J.B., Susumu, K., Stewart, M.H., Oh, E., Huston, A.L., Dawson, G., Ingale, S., Walters, R., Domowicz, M., Deschamps, J.R., Algar, W.R., Dimaggio, S., Manono, J., Spillmann, C.M., Thompson, D., Jennings, T. L., Dawson, P.E., Medintz, I.L., 2013. Selecting improved peptidyl motifs for cytosolic delivery of disparate protein and nanoparticle materials. ACS Nano 7 (5), 3778–3796. <https://doi.org/10.1021/nn400702r>.
- Bonchev, D., Balaban, A.T., Mekenyan, O., 1980. Generalization of the graph center concept, and derived topological centric indexes. J. Chem. Inf. Model. 20 (2), 106–113. <https://doi.org/10.1021/ci60022a011>.
- Buglak, A.A., Zherdev, A.V., Dzantiev, B.B., 2019. Nano-(Q)SAR for cytotoxicity prediction of engineered nanomaterials. Molecules. 24 (24) <https://doi.org/10.3390/molecules24244537>.
- Chahal, D.S., Chahal, H.S., Bayles, A.R., Rudié, E.M., Helms, B.A., 2012. Synthetic development of cell-permeable polymer colloids decorated with nanocrystal imaging probes optimized for cell tracking. Chem. Sci. 3 (7), 2246. <https://doi.org/10.1039/C2SC20206A>.
- Chen, F., Gerion, D., 2004. Fluorescent CdSe/ZnS nanocrystal–Peptide conjugates for long-term, nontoxic imaging and nuclear targeting in living cells. Nano Lett. 4 (10), 1827–1832. <https://doi.org/10.1021/nl049170q>.
- Chirico, N., Gramatica, P., 2011. Real external predictivity of QSAR models: how to evaluate it? Comparison of different validation criteria and proposal of using the concordance correlation coefficient. J. Chem. Inf. Model. 51 (9), 2320–2335. <https://doi.org/10.1021/ci200211n>.
- Choi, M.K., Yang, J., Hyeon, T., Kim, D.H., 2018. Flexible quantum dot light-emitting diodes for next-generation displays. NPJ flex. Electron. 2 (1), 49. <https://doi.org/10.1038/s41528-018-0023-3>.
- Choi, J.S., Trinh, T.X., Yoon, T.H., Kim, J., Byun, H.G., 2019. Quasi-QSAR for predicting the cell viability of human lung and skin cells exposed to different metal oxide nanomaterials. Chemosphere 217, 243–249. <https://doi.org/10.1016/j.chemosphere.2018.11.014>.
- Deka, S., Quarta, A., Lupo, M.G., Falqui, A., Boninelli, S., Giannini, C., Morello, G., Giorgi, M., Lanzani, G., Spinella, C., Cingolani, R., Pellegrino, T., Manna, L., 2009. CdSe/CdS/ZnS double shell nanorods with high photoluminescence efficiency and their exploitation as biolabeling probes. J. Am. Chem. Soc. 131 (8), 2948–2958. <https://doi.org/10.1021/ja808369e>.
- Erhيرhie, E.O., Ihekwereme, C.P., Ilodigwe, E.E., 2018. Advances in acute toxicity testing: strengths, weaknesses and regulatory acceptance. Interdiscip. Toxicol. 11 (1), 5–12. <https://doi.org/10.2478/intox-2018-0001>.
- Fu, Y., Ding, C., Zhu, A., Deng, Z., Tian, Y., Jin, M., 2013. Two-photon ratiometric fluorescent sensor based on specific biomolecular recognition for selective and sensitive detection of copper ions in live cells. Anal. Chem. 85 (24), 11936–11943. <https://doi.org/10.1021/ac403527c>.
- Goto, Y., Matsuno, R., Konno, T., Takai, M., Ishihara, K., 2008. Artificial cell membrane-covered nanoparticles embedding quantum dots as stable and highly sensitive fluorescence bioimaging probes. Biomacromolecules 9 (11), 3252–3257. <https://doi.org/10.1021/bm800819r>.
- Hu, L., Zeng, G., Chen, G., Huang, Z., Wan, J., Chen, A., Yu, Z., Yang, J., He, K., Qin, L., 2017. Bioaccumulation and toxicity of CdSe/ZnS quantum dots in Phanerochaete chrysosporium. Colloids Surf. B Biointerfaces 159, 303–311. <https://doi.org/10.1016/j.colsurfb.2017.08.006>.
- Imamoglu, A., 2003. Are quantum dots useful for quantum computation? Phys. E Low Dimens. Syst. Nanostruct. 16 (1), 47–50. [https://doi.org/10.1016/S1386-9477\(02\)00581-7](https://doi.org/10.1016/S1386-9477(02)00581-7).
- Jin, S., Hu, Y., Gu, Z., Liu, L., Wu, H.C., 2011. Application of quantum dots in biological imaging. J. Nanomat. 2011 (46), 1–13. <https://doi.org/10.1155/2011/834139>.
- Kahmann, S., Shulga, A., Loi, M.A., 2020. Quantum dot light-emitting transistors—powerful research tools and their future applications. Adv. Funct. Mater. 30 (20), 1904174 <https://doi.org/10.1002/adfm.201904174>.
- Kumar, A., Chauhan, S., 2017. QSAR differential model for prediction of SIRT1 modulation using Monte Carlo method. Drug Res. 67 (3), 156–162. <https://doi.org/10.1055/s-0042-119725>.
- Kumar, A., Chauhan, S., 2018. Use of Simplified Molecular Input Line Entry System and molecular graph based descriptors in prediction and design of pancreatic lipase inhibitors. Future Med. Chem. 10 (13), 1603–1622. <https://doi.org/10.4155/fmc-2018-0024>.
- Kumar, P., Kumar, A., 2019. Nucleobase sequence based building up of reliable QSAR models with the index of ideality correlation using Monte Carlo method. J. Biomol. Struct. Dyn. <https://doi.org/10.1080/07391102.2019.1656109>.
- Kumar, A., Kumar, P., 2020a. Construction of pioneering quantitative structure activity relationship screening models for abuse potential of designer drugs using index of ideality of correlation in Monte Carlo optimization. Arch. Toxicol. <https://doi.org/10.1007/s00204-020-02828-w>.

- Kumar, P., Kumar, A., 2020b. CORAL: QSAR models of CB1 cannabinoid receptor inhibitors based on local and global SMILES attributes with the index of ideality of correlation and the correlation contradiction index. *Chemometr. Intell. Lab. Syst.* 200, 103982. <https://doi.org/10.1016/j.chemolab.2020.103982>.
- Lee, J., Im, J.H., Huh, K.M., Lee, Y.K., Shin, H., 2010. Preparation and characterization of CdSe/ZnS quantum dots encapsulated in poly(ethylene glycol)-b-poly(D,L-lactide) micelle nanoparticles. *J. Nanosci. Nanotechnol.* 10 (1), 487–496. <https://doi.org/10.1166/jnn.2010.1736>.
- Towards efficient designing of safe nanomaterials. In: Leszczynski, J., Puzyn, T. (Eds.), 2012. *Nanoscience & Nanotechnology Series*. Royal Society of Chemistry, Cambridge.
- Lin, L.I.K., 1992. Assay validation using the concordance correlation coefficient. *Biometrics* 48 (2), 599. <https://doi.org/10.2307/2532314>.
- Liu, Y.-F., Xie, B., Yin, Z.-G., Fang, S.-M., Zhao, J.-B., 2010. Synthesis of highly stable CdTe/CdS quantum dots with biocompatibility. *Eur. J. Inorg. Chem.* 2010 (10), 1501–1506. <https://doi.org/10.1002/ejic.200900978>.
- Maity, A.R., Saha, A., Roy, A., Jana, N.R., 2013. Folic acid functionalized nanoprobes for fluorescence-, Dark-Field-, and dual-imaging-Based selective detection of cancer cells and tissue. *ChemPlusChem* 78 (3), 259–267. <https://doi.org/10.1002/cplu.201200296>.
- Manisha, Chauhan S., Kumar, P., Kumar, A., 2019. Development of prediction model for fructose-1,6- bisphosphatase inhibitors using the Monte Carlo method. *SAR QSAR Environ. Res.* 30 (3), 145–159. <https://doi.org/10.1080/1062936X.2019.1568299>.
- Mikolajczyk, A., Gajewicz, A., Mulkiewicz, E., Rasulev, B., Marchelek, M., Diak, M., Hirano, S., Zalewska-Medynska, A., Puzyn, T., 2018. Nano-QSAR modeling for ecosafe design of heterogeneous TiO₂ -based nano-photocatalysts. *Environ. Sci. Nano* 5 (5), 1150–1160. <https://doi.org/10.1039/C8EN00085A>.
- Nikazar, S., Sivasankarapillai, V.S., Rahdar, A., Gasmi, S., Anumol, P.S., Shanavas, M.S., 2020. Revisiting the cytotoxicity of quantum dots: an in-depth overview. *Biophys. Rev.* <https://doi.org/10.1007/s12551-020-00653-0>.
- Nimbhal, M., Bagri, K., Kumar, P., Kumar, A., 2020. The index of ideality of correlation: a statistical yardstick for better QSAR modeling of glucokinase activators. *Struct. Chem.* 31 (2), 831–839. <https://doi.org/10.1007/s11224-019-01468-w>.
- Oh, E., Liu, R., Nel, A., Gemmill, K.B., Bilal, M., Cohen, Y., Medintz, I.L., 2016. Meta-analysis of cellular toxicity for cadmium-containing quantum dots. *Nat. Nanotechnol.* 11 (5), 479–486. <https://doi.org/10.1038/NNANO.2015.338>.
- Papa, E., Gramatica, P., 2010. QSPR as a support for the EU REACH regulation and rational design of environmentally safer chemicals: PBT identification from molecular structure. *Green Chem.* 12 (5), 836. <https://doi.org/10.1039/B923843C>.
- Papa, E., van der Wal, L., Arnot, J.A., Gramatica, P., 2014. Metabolic biotransformation half-lives in fish: QSAR modeling and consensus analysis. *Sci. Total Environ.* 470–471, 1040–1046. <https://doi.org/10.1016/j.scitotenv.2013.10.068>.
- Qian, J., Gao, X., 2013. Triblock copolymer-encapsulated nanoparticles with outstanding colloidal stability for siRNA delivery. *ACS Appl. Mater. Interfaces* 5 (8), 2845–2852. <https://doi.org/10.1021/am3021813>.
- Quarta, A., Ragusa, A., Deka, S., Tortiglione, C., Tino, A., Cingolani, R., Pellegrino, T., 2009. Bioconjugation of rod-shaped fluorescent nanocrystals for efficient targeted cell labeling. *Langmuir* 25 (21), 12614–12622. <https://doi.org/10.1021/la901831y>.
- Raies, A.B., Bajic, V.B., 2016. In silico toxicology: computational methods for the prediction of chemical toxicity. *Wiley Interdiscip. Rev. Comput. Mol. Sci.* 6 (2), 147–172. <https://doi.org/10.1002/wcms.1240>.
- Randić, M., 2001. Graph valence shells as molecular descriptors. *J. Chem. Inf. Comput. Sci.* 41 (3), 627–630. <https://doi.org/10.1021/ci000121i>.
- Rasulev, B., Gajewicz, A., Puzyn, T., Leszczynski, J., Leszczynski, J., 2012. Chapter 10. Nano-QSAR: advances and challenges. In: Leszczynski, J., Puzyn, T. (Eds.), *Towards Efficient Designing of Safe Nanomaterials*. Royal Society of Chemistry, Cambridge, pp. 220–256.
- Reithmaier, J.P., Forchel, A., 2003. Recent advances in semiconductor quantum-dot lasers. *C. R. Phys.* 4 (6), 611–619. [https://doi.org/10.1016/S1631-0705\(03\)00075-6](https://doi.org/10.1016/S1631-0705(03)00075-6).
- Reshma, V.G., Mohanan, P.V., 2019. Quantum dots: applications and safety consequences. *J. Lumin.* 205, 287–298. <https://doi.org/10.1016/j.jlumin.2018.09.015>.
- Roy, K., Kar, S., Ambure, P., 2015. On a simple approach for determining applicability domain of QSAR models. *Chemometr. Intell. Lab. Syst.* 145, 22–29. <https://doi.org/10.1016/j.chemolab.2015.04.013>.
- Roy, K., Ambure, P., Kar, S., Ojha, P.K., 2018. Is it possible to improve the quality of predictions from an “intelligent” use of multiple QSAR/QSPR/QSTR models? *J. Chemom.* 32 (4), e2992. <https://doi.org/10.1002/cem.2992>.
- Rozenzhak, S.M., Kadakia, M.P., Caserta, T.M., Westbrook, T.R., Stone, M.O., Naik, R.R., 2005. Cellular internalization and targeting of semiconductor quantum dots. *Chem. Commun.* (17), 2217–2219. <https://doi.org/10.1039/B418454H>.
- Sakho, E.H.M., Oluwafemi, O.S., 2019. Quantum dots for solar cell applications. *Nanomaterials for Solar Cell Applications*. Elsevier, pp. 377–415.
- Shiohara, A., Hoshino, A., Hanaki, K.-I., Suzuki, K., Yamamoto, K., 2004. On the cytotoxicity caused by quantum dots. *Microbiol. Immunol.* 48 (9), 669–675. <https://doi.org/10.1111/j.1348-0421.2004.tb03478.x>.
- Toropov, A.A., Toropova, A.P., 2015. Quasi-SMILES and nano-QFAR: united model for mutagenicity of fullerene and MWCNT under different conditions. *Chemosphere* 139, 18–22. <https://doi.org/10.1016/j.chemosphere.2015.05.042>.
- Toropov, A.A., Toropova, A.P., 2017. The index of ideality of correlation: a criterion of predictive potential of QSPR/QSAR models? *Mutat. Res.* 819, 31–37. <https://doi.org/10.1016/j.mrgentox.2017.05.008>.
- Toropov, A.A., Toropova, A.P., 2019. The Correlation Contradictions Index (CCI): building up reliable models of mutagenic potential of silver nanoparticles under different conditions using quasi-SMILES. *Sci. Total Environ.* 681, 102–109. <https://doi.org/10.1016/j.scitotenv.2019.05.114>.
- Toropov, A.A., Toropova, A.P., Benfenati, E., Gini, G., Leszczynski, J., 2013a. CORAL: QSAR model of water solubility based on local and global SMILES attributes. *Chemosphere* 90 (2), 877–880. <https://doi.org/10.1016/j.chemosphere.2012.07.035>.
- Toropov, A.A., Toropova, A.P., Puzyn, T., Benfenati, E., Gini, G., Leszczynski, J., Leszczynski, J., 2013b. QSAR as a random event: modeling of nanoparticles uptake in PaCa2 cancer cells. *Chemosphere* 92 (1), 31–37. <https://doi.org/10.1016/j.chemosphere.2013.03.012>.
- Toropov, A.A., Rallo, R., Toropova, A.P., 2015. Use of Quasi-SMILES and monte carlo optimization to develop quantitative feature Property/Activity relationships (QFPR/QFAR) for nanomaterials. *Curr. Top. Med. Chem.* 15 (18), 1837–1844. <https://doi.org/10.2174/1568026615666150506152000>.
- Toropov, A.A., Toropova, A.P., Roncaglioni, A., Benfenati, E., 2018. Prediction of biochemical endpoints by the CORAL software: prejudices, paradoxes, and results. *Methods Mol. Biol.* 1800, 573–583. https://doi.org/10.1007/978-1-4939-7899-1_27.
- Toropov, A.A., Toropova, A.P., Raitano, G., Benfenati, E., 2019. CORAL: building up QSAR models for the chromosome aberration test. *Saudi J. Biol. Sci.* 26 (6), 1101–1106. <https://doi.org/10.1016/j.sjbs.2018.05.013>.
- Toropova, A.P., Toropov, A.A., Martyanov, S.E., Benfenati, E., Gini, G., Leszczynska, D., Leszczynski, J., 2012. CORAL: QSAR modeling of toxicity of organic chemicals towards *Daphnia magna*. *Chemometr. Intell. Lab. Syst.* 110 (1), 177–181. <https://doi.org/10.1016/j.chemolab.2011.10.005>.
- Toropova, A.P., Toropov, A.A., Veselinović, A.M., Veselinović, J.B., Leszczynska, D., Leszczynski, J., 2017. Quasi-SMILES as a novel tool for prediction of nanomaterials' endpoints. *Multi-Scale Approaches in Drug Discovery*. Elsevier, pp. 191–221.
- Trinh, T.X., Choi, J.-S., Jeon, H., Byun, H.-G., Yoon, T.-H., Kim, J., 2018. Quasi-SMILES-Based nano-quantitative structure-activity relationship model to predict the cytotoxicity of multiwalled carbon nanotubes to human lung cells. *Chem. Res. Toxicol.* 31 (3), 183–190. <https://doi.org/10.1021/acs.chemrestox.7b00303>.
- Yeh, Y.-C., Saha, K., Yan, B., Miranda, O.R., Yu, X., Rotello, V.M., 2013. The role of ligand coordination on the cytotoxicity of cationic quantum dots in HeLa cells. *Nanoscale* 5 (24), 12140–12143. <https://doi.org/10.1039/c3nr04037b>.
- Yu, M., Yang, Y., Han, R., Zheng, Q., Wang, L., Hong, Y., Li, Z., Sha, Y., 2010. Polyvalent lactose-quantum dot conjugate for fluorescent labeling of live leukocytes. *Langmuir* 26 (11), 8534–8539. <https://doi.org/10.1021/la904488w>.
- Zhang, H.-L., Li, Y.-Q., Wang, J.-H., Li, X.-N., Lin, S., Zhao, Y.-D., Luo, Q.-M., 2010. Special method to prepare quantum dot probes with reduced cytotoxicity and increased optical property. *J. Biomed. Opt.* 15 (1), 15001. <https://doi.org/10.1117/1.3291999>.
- Zhang, M.-Z., Yu, R.-N., Chen, J., Ma, Z.-Y., Zhao, Y.-D., 2012. Targeted quantum dots fluorescence probes functionalized with aptamer and peptide for transferrin receptor on tumor cells. *Nanotechnology* 23 (48), 485104. <https://doi.org/10.1088/0957-4484/23/48/485104>.
- Zhao, Y., Liu, S., Li, Y., Jiang, W., Chang, Y., Pan, S., Fang, X., Wang, Y.A., Wang, J., 2010. Synthesis and grafting of folate-PEG-PAMAM conjugates onto quantum dots for selective targeting of folate-receptor-positive tumor cells. *J. Colloid Interface Sci.* 350 (1), 44–50. <https://doi.org/10.1016/j.jcis.2010.05.035>.
- Zhu, H., Tropsha, A., Fourches, D., Varnek, A., Papa, E., Gramatica, P., Oberg, T., Dao, P., Cherkasov, A., Tetko, I.V., 2008. Combinatorial QSAR modeling of chemical toxicants tested against *Tetrahymena pyriformis*. *J. Chem. Inf. Model.* 48 (4), 766–784. <https://doi.org/10.1021/ci700443v>.



Quantitative structure toxicity analysis of ionic liquids toward acetylcholinesterase enzymes using novel QSTR models with index of ideality of correlation and correlation contradictions index

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Quantitative Structure Toxicity Analysis of Ionic Liquids toward Acetylcholinesterase Enzymes Using Novel QSTR Models with Index of Ideality of Correlation and Correlation Contradictions Index

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Abstract: Ionic liquids (ILs) have enticed the curiosity of chemists due to their vast applications in academic and industrial research. These have many advantages over other conventional solvents such as broad liquid temperature, negligible vapour pressure, non-volatility, etc. But, from an environmental point of view, these advantages can develop heterogeneous toxic results when released into the environment. It is important to predict the toxicity of ionic liquids. A useful tool for predicting ILs toxicity is the quantitative structure-toxicity relationship (QSTR). The toxicity of ionic liquids is evaluated by predicting the acetylcholinesterase (AChE, EC3.1.1.7) enzyme inhibition. In the present manuscript, an exhaustive QSTR analysis for 229 ionic liquids as an acetylcholinesterase enzyme inhibitor is described using the inbuilt Monte Carlo optimization method of CORAL software. Eleven splits are prepared and from these split, 22 QSTR models are developed using two target functions, i.e. TF₁ (without IIC) and TF₂ (with IIC). All models developed by TF₂ are robust and have better predictability. The model developed for split 1 using TF₂ is considered as the best model ($R^2_{Valid} = 0.7782$). In the present work, a novel parameter “Correlation Contradiction Index (CCI)” is studied to recognize its predictability. The docking simulation was also performed to understand the mechanistic interpretation. Further, the mechanistic interpretation of the best QSTR model was in good correlation with the three-dimensional studies of ligand binding. In order to see the true picture of inhibitory potential, ligand transport study of five ILs (IL015, IL040, IL116, IL156 and IL211) was studied in the tunnel leading to the active site of AChE using the services of Caver Web. Result of the transport study showed that these ILs formed a most stable complex in the active site and not in the tunnel and did not obstruct the tunnel for the accessibility of the enzyme active site for the substrate.

Keyword: QSTR, Ionic Liquids, Toxicity, Acetylcholinesterase, IIC, CCI

References

- [1] S. Oguzcan, J. Dvarioniene, A. Tugnoli, J. Kruopiene, Environmental impact assessment model for substitution of hazardous substances by using life cycle approach, *Environmental Pollution* 254 (2019). doi:10.1016/j.envpol.2019.07.113.
- [2] E.B. Wedebye, M. Dybdahl, N.G. Nikolov, S.T. Jónsdóttir, J.R. Niemelä, QSAR screening of 70,983 REACH substances for genotoxic carcinogenicity, mutagenicity and developmental toxicity in the ChemScreen project, *Reproductive Toxicology* 55 (2015) 64. doi:10.1016/j.reprotox.2015.03.002.
- [3] X. Yang, Y. Wang, R. Byrne, G. Schneider, S. Yang, Concepts of Artificial Intelligence for Computer-Assisted Drug Discovery, *Chemical Review*, 119 (2019) 10520. doi:10.1021/acs.chemrev.8b00728.
- [4] P. Heitel, L. Gellrich, L. Kalinowsky, J. Heering, A. Kaiser, J. Ohrndorf, E. Proschak, D. Merk, Computer-Assisted Discovery and Structural Optimization of a Novel Retinoid X Receptor Agonist Chemotype, *ACS Medicinal Chemistry Letters* 10 (2019) 203. doi:10.1021/acsmedchemlett.8b00551.
- [5] R.P. Sharma, V. Kumar, M. Schuhmacher, A. Kol' dkina, H.V. Westerhoff, Development and evaluation of a harmonized whole body physiologically based pharmacokinetic (PBPK) model for flutamide in rats and its extrapolation to humans, *Environmental Research* 182 (2020). doi:10.1016/j.envres.2019.108948.
- [6] K. Roy, G. Ghosh, QSTR with extended topochemical atom indices. Part 5: Modeling of the acute toxicity of phenylsulfonyl carboxylates to *Vibrio fischeri* using genetic function approximation, *Bioorganic and Medicinal Chemistry* 13 (2005) 1185. doi:10.1016/j.bmc.2004.11.014.
- [7] M.Y. Moridani, A. Siraki, T. Chevaldina, H. Scobie, P.J. O'Brien, Quantitative structure toxicity relationships for catechols in isolated rat hepatocytes, *Chemico-Biological Interactions* 147 (2004) 297. doi:10.1016/j.cbi.2004.02.001.
- [8] G. Jana, R. Pal, S. Sural, P.K. Chattaraj, *Methods in Pharmacology and Toxicology*, Humana Press Inc., 2020, p. 661-679.
- [9] G. Jana, R. Pal, S. Sural, P.K. Chattaraj, Quantitative structure-toxicity relationship: An “in silico study” using electrophilicity and hydrophobicity as descriptors, *International Journal of Quantum Chemistry* 120 (2020). doi:10.1002/qua.26097.
- [10] Z. Zhang, J. Song, B. Han, Catalytic Transformation of Lignocellulose into Chemicals and Fuel Products in Ionic Liquids, *Chem. Rev.* (2017).
- [11] B. Wang, L. Qin, T. Mu, Z. Xue, G. Gao, Are Ionic Liquids Chemically Stable?, *Chem. Rev.* (2017).
- [12] Z. Lei, B. Chen, Y.-M. Koo, D.R. MacFarlane, Introduction: Ionic Liquids, *Chemical Reviews* 117 (2017) 6633. doi:10.1021/acs.chemrev.7b00246.

- [13] K.S. Egorova, E.G. Gordeev, V.P. Ananikov, Biological Activity of Ionic Liquids and Their Application in Pharmaceutics and Medicine, *Chemical Reviews* 117 (2017) 7132. doi:10.1021/acs.chemrev.6b00562.
- [14] Z. Zheng, Q. Xu, J. Guo, J. Qin, H. Mao, B. Wang, F. Yan, Structure-Antibacterial Activity Relationships of Imidazolium-Type Ionic Liquid Monomers, Poly(ionic liquids) and Poly(ionic liquid) Membranes: Effect of Alkyl Chain Length and Cations, *ACS Appl. Mater. Interfaces* 8 (2016) 12684.
- [15] H.C. Zhang, C.Y. Shi, H.H. Yang, G.W. Chen, D.Z. Liu, Genotoxicity Evaluation of Ionic Liquid 1-Octyl-3-methylimidazolium Bromide in Freshwater Planarian *Dugesia japonica* Using RAPD Assay, *Ecotoxicol. Environ. Saf.* 134P1 (2016) 17.
- [16] H.C. Zhang, C.Y. Shi, L.Q. Sun, F. Wang, G.W. Chen, Toxic Effects of Ionic Liquid 1-Octyl-3-methylimidazolium Bromide on the Antioxidant Defense System of Freshwater Planarian, *Dugesia japonica*, *Toxicol. Ind. Health* 32 (2016) 1675.
- [17] F. Yan, S. Xia, Q. Wang, P. Ma, Predicting Toxicity of Ionic Liquids in Acetylcholinesterase Enzyme by the Quantitative Structure-Activity Relationship Method Using Topological Indexes, *Journal of Chemical & Engineering Data* 57 (2012) 2252. doi:10.1021/je3002046.
- [18] P. Zhu, X. Kang, Y. Zhao, U. Latif, H. Zhang, Predicting the toxicity of ionic liquids toward acetylcholinesterase enzymes using novel QSAR models, *International Journal of Molecular Sciences* 20 (2019). doi:10.3390/ijms2009184.
- [19] F. Siopa, R.F.M. Fraude, A. Diniz, J.M. Andrade, M. Nicolai, A. Meirinhos, S.D. Lucas, F. Marcelo, C.A.M. Afonso, P. Rijo, Acetylcholinesterase Choline-Based Ionic Liquid Inhibitors: In Vitro and in Silico Molecular Docking Studies, *ACS Omega* 3 (2018) 17145. doi:10.1021/acsomega.8b02347.
- [20] S. Stolte, T. Schulz, C.W. Cho, J. Arning, T. Strassner, Synthesis, toxicity, and biodegradation of tunable aryl alkyl ionic liquids (TAAILs), *ACS Sustainable Chemistry and Engineering* 1 (2013) 410. doi:10.1021/sc300146t.
- [21] M.G. Lionetto, R. Caviglio, A. Calisi, M.E. Giordano, T. Schettino, Acetylcholinesterase as a biomarker in environmental and occupational medicine: new insights and future perspectives, *Biomed Res Int* 2013 (2013) 321213. doi:10.1155/2013/321213.
- [22] M. Sugimoto, A.B. Manggara, K. Yoshida, T. Inoue, T. Ideo, An Electronic-structure Informatics Study on the Toxicity of Alkylphenols to Tetrahymena pyriformis, *Molecular Informatics* 39 (2020). doi:10.1002/minf.201900121.
- [23] M. Marzo, G.J. Lavado, F. Como, A.P. Toropova, A.A. Toropov, D. Baderna, C. Cappelli, A. Lombardo, C. Toma, M. Blázquez, E. Benfenati, QSAR models for biocides: The example of the prediction of *Daphnia magna* acute toxicity, SAR and QSAR in Environmental Research 31 (2020) 227. doi:10.1080/1062936X.2019.1709221.
- [24] Manisha, S. Chauhan, P. Kumar, A. Kumar, Development of prediction model for fructose-1,6- bisphosphatase inhibitors using the Monte Carlo method, SAR and QSAR in Environmental Research 30 (2019) 145. doi:10.1080/1062936X.2019.1568299.

- [25] P. Kumar, A. Kumar, J. Sindhu, In silico design of diacylglycerol acyltransferase-1 (DGAT1) inhibitors based on SMILES descriptors using Monte-Carlo method, SAR and QSAR in Environmental Research 30 (2019) 525. doi:10.1080/1062936X.2019.1629998.
- [26] S. Ahmadi, F. Mardinia, N. Azimi, M. Qomi, E. Balali, Prediction of chalcone derivative cytotoxicity activity against MCF-7 human breast cancer cell by Monte Carlo method, Journal of Molecular Structure 1181 (2019) 305. doi:10.1016/j.molstruc.2018.12.089.
- [27] M.A. Toropova, M. Raškova, I. Raška, Jr., A.P. Toropova, The Index of Ideality of Correlation (IIC): model for sweetness, Monatshefte fur Chemie 150 (2019) 617. doi:10.1007/s00706-019-2368-2.
- [28] A.P. Toropova, A.A. Toropov, Application of the Monte Carlo method for the prediction of behavior of peptides, Current Protein and Peptide Science 20 (2019) 1151. doi:10.2174/1389203720666190123163907.
- [29] A.A. Toropov, A.P. Toropova, G. Raitano, E. Benfenati, CORAL: Building up QSAR models for the chromosome aberration test, Saudi J Biol Sci 26 (2019) 1101. doi:10.1016/j.sjbs.2018.05.013.
- [30] P. Kumar, A. Kumar, J. Sindhu, S. Lal, QSAR Models for Nitrogen Containing Monophosphonate and Bisphosphonate Derivatives as Human Farnesyl Pyrophosphate Synthase Inhibitors Based on Monte Carlo Method, Drug Research 69 (2019) 159. doi:10.1055/a-0652-5290.
- [31] M. Nimbhal, K. Bagri, P. Kumar, A. Kumar, The index of ideality of correlation: A statistical yardstick for better QSAR modeling of glucokinase activators, Structural Chemistry 31 (2020) 831. doi:10.1007/s11224-019-01458-w.
- [32] P. Kumar, A. Kumar, Nucleobase sequence based building up of reliable QSAR models with the index of ideality correlation using Monte Carlo method, J Biomol Struct Dyn 38 (2020) 3296. doi:10.1080/07391102.2019.1556109.
- [33] M. Duhan, R. Singh, M. Devi, J. Sindhu, R. Bhatia, A. Kumar, P. Kumar, Synthesis, molecular docking and QSAR study of thiazole clubbed pyrazole hybrid as alpha-amylase inhibitor, J Biomol Struct Dyn (2019) 1. doi:10.1080/07391102.2019.1704885.
- [34] P. Kumar, K. Kadyan, M. Duhan, J. Sindhu, K. Hussain, S. Lal, Silica-supported ceric ammonium nitrate (CAN): a simple, mild and solid-supported reagent for quickest oxidative aromatization of Hantzsch 1,4-dihydropyridines, Chemical Papers 73 (2019) 1153. doi:10.1007/s11696-018-0666-5.
- [35] P. Kumar, M. Duhan, J. Sindhu, K. Kadyan, S. Saini, N. Panigar, Thiazolidine-4-one clubbed pyrazoles hybrids: Potent α -amylase and α -glucosidase inhibitors with NLO properties, Journal of Heterocyclic Chemistry 57 (2020) 1573. doi:10.1002/jhet.3882.
- [36] R. Bhatia, K. Kadyan, M. Duhan, M. Devi, R. Singh, R.C. Kamboj, P. Kumar, A Serendipitous Synthesis: SiO₂-HNO₃ Mediated Oxidative Aromatization and Regioselective Nitration of 1,3,5-Trisubstituted-4,5-Dihydro-1H-Pyrazoles, Chemistryselect 4 (2019) 10417. doi:10.1002/slct.201902285.

- [37] P. Kumar, K. Kadyan, M. Duhan, J. Sindhu, V. Singh, B.S. Saharan, Design, synthesis, conformational and molecular docking study of some novel acyl hydrazone based molecular hybrids as antimalarial and antimicrobial agents, *Chem Cent J* 11 (2017) 115. doi:10.1186/s13065-017-0344-7.
- [38] P. Kumar, M. Duhan, K. Kadyan, J. Sindhu, S. Kumar, H. Sharma, Synthesis of novel inhibitors of alpha-amylase based on the thiazolidine-4-one skeleton containing a pyrazole moiety and their configurational studies, *Medchemcomm* 8 (2017) 1468. doi:10.1039/c7md00080d.
- [39] P. Kumar, R. Bhatia, R. Khanna, A. Dalal, D. Kumar, P. Surain, R.C. Kamboj, Synthesis of some benzothiazoles by developing a new protocol using urea nitrate as a catalyst and their antimicrobial activities, *Journal of Sulfur Chemistry* 38 (2017) 585. doi:10.1080/17415993.2017.1334781.
- [40] A.P. Toropova, A.A. Toropov, The index of ideality of correlation: improvement of models for toxicity to algae, *Nat Prod Res* 33 (2019) 2200. doi:10.1080/14786419.2018.1493591.
- [41] V.P. Nickovic, Z.N. Vujnovic-Zivkovic, R. Trajkovic, D. Krstic, L. Ristic, M. Radovic, Z. Ceric, D. Sokolovic, A.M. Veselinovic, In silico studies and the design of novel agents for the treatment of systemic tuberculosis, *J Biomol Struct Dyn* 37 (2019) 3198. doi:10.1080/07391102.2018.1511476.
- [42] M. Douziech, R. Oldenkamp, R. van Zelst, H. King, A.J. Hendriks, A.S. Ficheux, M.A.J. Huijbregts, Confronting variability with uncertainty in the ecotoxicological impact assessment of down-the-drain products, *Environ Int* 126 (2019) 37. doi:10.1016/j.envint.2019.01.080.
- [43] A.A. Toropov, A.P. Toropova, The Correlation Contradictions Index (CCI): Building up reliable models of mutagenic potential of silver nanoparticles under different conditions using quasi-SMILES, *Sci Total Environ* 681 (2019) 102. doi:10.1016/j.scitotenv.2019.05.114.
- [44] P. Kumar, A. Kumar, COR AL: QSAR models of CB1 cannabinoid receptor inhibitors based on local and global SMILES attributes with the index of ideality of correlation and the correlation contradiction index, *Chemometrics and Intelligent Laboratory Systems* 200 (2020) 103982. doi:10.1016/j.chemolab.2020.103982.
- [45] A.P. Toropova, A.A. Toropov, E. Carnesecchi, E. Benfenati, J.L. Dorne, The using of the Index of Ideality of Correlation (IIC) to improve predictive potential of models of water solubility for pesticides, *Environ Sci Pollut Res Int* (2020). doi:10.1007/s11356-020-07820-6.
- [46] N. Basant, S. Gupta, K.P. Singh, Predicting acetyl cholinesterase enzyme inhibition potential of ionic liquids using machine learning approaches: An aid to green chemicals designing, *Journal of Molecular Liquids* 209 (2015) 404. doi:<https://doi.org/10.1016/j.molliq.2015.06.001>.
- [47] C.W. Cho, Y.S. Yun, Interpretation of toxicological activity of ionic liquids to acetylcholinesterase inhibition via in silico modelling, *Chemosphere* 159 (2016) 178. doi:10.1016/j.chemosphere.2016.06.005.

- [48] A.P. Toropova, A.A. Toropov, M. Beeg, M. Gobbi, M. Salmona, Utilization of the Monte Carlo Method to Build up QSAR Models for Hemolysis and Cytotoxicity of Antimicrobial Peptides, *Curr Drug Discov Technol* 14 (2017) 229. doi:10.2174/1570163814666170525114128.
- [49] D. Sokolović, J. Ranković, V. Stanković, R. Stefanović, S. Karaleić, B. Mekić, V. Milenković, J. Kocić, A.M. Veselinović, QSAR study of dipeptidyl peptidase-4 inhibitors based on the Monte Carlo method, *Medicinal Chemistry Research* 26 (2017) 796. doi:10.1007/s00044-017-1792-2.
- [50] A.P. Toropova, A.A. Toropov, QSPR and nano-QSPR: What is the difference?, *Journal of Molecular Structure* 1182 (2019) 141. doi:10.1016/j.molstruc.2019.01.040.
- [51] J.B. Veselinović, V. Đorđević, M. Bogdanović, I. Morić, A.M. Veselinović, QSAR modeling of dihydrofolate reductase inhibitors as a therapeutic target for multiresistant bacteria, *Structural Chemistry* 29 (2018) 541. doi:10.1007/s11224-017-1051-7.
- [52] S. Ahmadi, H. Ghanbari, S. Lotfi, N. Azimi, Predictive QSAR modeling for the antioxidant activity of natural compounds derivatives based on Monte Carlo method, *Molecular Diversity* (2020). doi:10.1007/s11030-019-10026-9.
- [53] P. Kumar, A. Kumar, CORAL: QSAR models of CB1 cannabinoid receptor inhibitors based on local and global SMILES attributes with the index of ideality of correlation and the correlation contradiction index, *Chemometrics and Intelligent Laboratory Systems* 200 (2020). doi:10.1016/j.chemolab.2020.103982.
- [54] A.P. Toropova, P.R. Duchowicz, L.M. Saavedra, E.A. Castro, A.A. Toropov, The Use of the Index of Ideality of Correlation to Build Up Models for Bioconcentration Factor, *Molecular Informatics* (2020). doi:10.1002/minf.201900070.
- [55] R.B. Aher, K. Roy, Exploring the structural requirements in multiple chemical scaffolds for the selective inhibition of *Plasmodium falciparum* calcium-dependent protein kinase-1 (PfCDPK-1) by 3D-pharmacophore modelling, and docking studies, *SAR QSAR Environ Res* 28 (2017) 390. doi:10.1080/1062936X.2017.1326401.
- [56] S.S. Bhayye, K. Roy, A. Søha, Pharmacophore generation, atom-based 3D-QSAR, HQSAR and activity cliff analyses of benzothiazine and deazaxanthine derivatives as dual A2A antagonists/MAOB inhibitors, *SAR QSAR Environ Res* 27 (2016) 183. doi:10.1080/1062936X.2015.1136840.
- [57] A. Golbraikh, A. Tropsha, Beware of q2!, *J Mol Graph Model* 20 (2002) 269. doi:10.1016/s1093-3263(01)00123-1.
- [58] L.M. Shi, H. Fang, W. Tong, J. Wu, R. Perkins, R.M. Blair, W.S. Branham, S.L. Dial, C.L. Moland, D.M. Sheehan, QSAR models using a large diverse set of estrogens, *J Chem Inf Comput Sci* 41 (2001) 186. doi:10.1021/ci000066d.
- [59] G. Schuurmann, R.U. Ebert, J. Chen, B. Wang, R. Kuhne, External validation and prediction employing the predictive squared correlation coefficient test set activity mean vs training set activity mean, *J Chem Inf Model* 48 (2008) 2140. doi:10.1021/ci800253u.
- [60] P. Pratim Roy, S. Paul, I. Mitra, K. Roy, On two novel parameters for validation of predictive QSAR models, *Molecules* 14 (2009) 1660. doi:10.3390/molecules14051660.

- [61] K. Roy, P. Chakraborty, I. Mitra, P.K. Ojha, S. Kar, R.N. Das, Some case studies on application of "r(m)2" metrics for judging quality of quantitative structure-activity relationship predictions: emphasis on scaling of response data, *J Comput Chem* 34 (2013) 1071. doi:10.1002/jcc.23231.
- [62] P. Gramatica, A. Sangion, A Historical Excursus on the Statistical Validation Parameters for QSAR Models: A Clarification Concerning Metrics and Terminology, *J Chem Inf Model* 56 (2016) 1127. doi:10.1021/acs.jcim.6b00088.
- [63] A.P. Toropova, A.A. Toropov, The index of ideality of correlation: improvement of models for toxicity to algae, *Nat Prod Res* (2018) 1. doi:10.1080/14786419.2018.1493591.
- [64] A.P. Toropova, A.A. Toropov, The index of ideality of correlation: A criterion of predictability of QSAR models for skin permeability?, *Sci Total Environ* 586 (2017) 466. doi:10.1016/j.scitotenv.2017.01.198.
- [65] A.A. Toropov, A.P. Toropova, Use of the index of ideality of correlation to improve predictive potential for biochemical endpoints, *Toxicol Mech Methods* 29 (2019) 43. doi:10.1080/15376516.2018.1506851.
- [66] G. Gatidou, N. Vazaiou, N.S. Thomaidis, A.S. Sasinakis, Biodegradability assessment of food additives using OECD 301F respirometric test, *Chemosphere* 241 (2020). doi:10.1016/j.chemosphere.2019.125071.
- [67] D. Yordanova, T.W. Schultz, C. Kuseva, K. Tankova, H. Ivanova, I. Dermen, T. Pavlov, S. Temelkov, A. Chapkanov, M. Georgiev, A. Gssi, T. Sobanski, O.G. Mekyan, Automated and standardized workflows in the OECD QSAK Toolbox, *Computational Toxicology* 10 (2019) 89. doi:10.1016/j.comtox.2019.01.006.
- [68] I.V. Tetko, I. Sushko, A.K. Pandey, H. Zhu, A. Tropsha, E. Papa, T. Oberg, R. Todeschini, D. Fourches, A. Varnek, Critical assessment of QSAR models of environmental toxicity against *Tetrahymena pyriformis*: focusing on applicability domain and overfitting by variable selection, *J Chem Inf Model* 48 (2008) 1733. doi:10.1021/ci800151m.
- [69] S. Ahmadi, H. Ghanbari, S. Lotfi, N. Azimi, Predictive QSAR modeling for the antioxidant activity of natural compounds derivatives based on Monte Carlo method, *Mol Divers* (2020). doi:10.1007/s11030-019-10026-9.
- [70] A.A. Toropov, A.P. Toropova, G. Selvestrel, E. Benfenati, Idealization of correlations between optimal simplified molecular input-line entry system-based descriptors and skin sensitization, *SAR and QSAR in Environmental Research* 30 (2019) 447. doi:10.1080/1062936X.2019.1615547.
- [71] P.K. Ojha, I. Mitra, R.N. Das, K. Roy, Further exploring rm2 metrics for validation of QSPR models, *Chemometrics and Intelligent Laboratory Systems* 107 (2011) 194. doi:<https://doi.org/10.1016/j.chemolab.2011.03.011>.
- [72] A.P. Toropova, A.A. Toropov, Does the Index of Ideality of Correlation Detect the Better Model Correctly?, *Molecular Informatics* 38 (2019). doi:10.1002/minf.201800157.
- [73] C. Rücker, G. Rücker, M. Meringer, y-Randomization and Its Variants in QSPR/QSAR, *Journal of Chemical Information and Modeling* 47 (2007) 2345. doi:10.1021/ci700157b.

- [74] P. Kumar, A. Kumar, Monte Carlo Method Based QSAR Studies of Mer Kinase Inhibitors in Compliance with OECD Principles, *Drug Res (Stuttg)* 68 (2018) 189. doi:10.1055/s-0043-119288.
- [75] F. Stock, J. Hoffmann, J. Ranke, R. Störmann, B. Ondruschka, B. Jastorff, Effects of ionic liquids on the acetylcholinesterase - A structure-activity relationship consideration, *Green Chemistry* 6 (2004) 286. doi:10.1039/b402348j.
- [76] O. Trott, A.J. Olson, AutoDock Vina: Improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading, *Journal of Computational Chemistry* 31 (2010) 455. doi:10.1002/jcc.21334.
- [77] M. Harel, I. Schalk, L. Ehret-Sabatier, F. Bouet, M. Goeldner, C. Hirth, P.H. Axelsen, I. Silman, J.L. Sussman, Quaternary ligand binding to aromatic residues in the active-site gorge of acetylcholinesterase, *Proceedings of the National Academy of Sciences of the United States of America* 90 (1993) 9031. doi:10.1073/pnas.90.19.9031.
- [78] E.F. Pettersen, T.D. Goddard, C.C. Huang, G.S. Couch, D.M. Greenblatt, E.C. Meng, T.E. Ferrin, UCSF Chimera—A visualization system for exploratory research and analysis, 25 (2004) 1605. doi:10.1002/jcc.20084.
- [79] J. Stourac, O. Vavra, P. Kokkonen, J. Filipovic, G. Pinto, J. Brezovsky, J. Damborsky, D. Bednar, Caver Web 1.0: identification of tunnels and channels in proteins and analysis of ligand transport, *Nucleic Acids Research* 47 (2019) V414. doi:10.1093/nar/gkz378 %J Nucleic Acids Research.
- [80] E. Chovancova, A. Pavelka, P. Benes, C. Strnad, J. Brezovsky, B. Kozlikova, A. Gora, V. Sustr, M. Klvana, P. Medek, L. Biedermannova, J. Sochor, J. Damborsky, CAVER 3.0: A Tool for the Analysis of Transport Pathways in Dynamic Protein Structures, *PLOS Computational Biology* 8 (2012) e1002708. doi:10.1371/journal.pcbi.1002708.
- [81] J. Filipovic, O. Vavra, J. Mlhal, D. Bednar, S.M. Marques, J. Brezovsky, L. Matyska, J. Damborsky, CaverDock: A Novel Method for the Fast Analysis of Ligand Transport, *IEEE/ACM transactions on computational biology and bioinformatics* (2019). doi:10.1109/tcbb.2019.2907402.
- [82] A. Ghaedi, Predicting the cytotoxicity of ionic liquids using QSAR model based on SMILES optimal descriptors, *Journal of Molecular Liquids* 208 (2015) 269. doi:<https://doi.org/10.1016/j.molliq.2015.04.049>.
- [83] J. Arning, S. Stolte, A. Böschen, F. Stock, W.-R. Pitner, U. Welz-Biermann, B. Jastorff, J. Ranke, Qualitative and quantitative structure activity relationships for the inhibitory effects of cationic head groups, functionalised side chains and anions of ionic liquids on acetylcholinesterase, *Green Chemistry* 10 (2008) 47. doi:10.1039/B712109A.
- [84] J.S. Torrecilla, J. García, E. Rojo, F. Rodríguez, Estimation of toxicity of ionic liquids in Leukemia Rat Cell Line and Acetylcholinesterase enzyme by principal component analysis, neural networks and multiple lineal regressions, *Journal of Hazardous Materials* 164 (2009) 182. doi:<https://doi.org/10.1016/j.jhazmat.2008.08.022>.

- [85] R.N. Das, K. Roy, Predictive in silico Modeling of Ionic Liquids toward Inhibition of the Acetyl Cholinesterase Enzyme of Electrophorus electricus: A Predictive Toxicology Approach, Industrial & Engineering Chemistry Research 53 (2014) 1020. doi:10.1021/ie403636q.
- [86] B. Peric, J. Sierra, E. Martí, R. Cruañas, M.A. Garau, Quantitative structure–activity relationship (QSAR) prediction of (eco)toxicity of short aliphatic protic ionic liquids, Ecotoxicology and Environmental Safety 115 (2015) 257. doi:<https://doi.org/10.1016/j.ecoenv.2015.02.027>.

CRediT author statement:

Both authors have an equal contribution.

Ashwani Kumar: Data curation, Software, Validation, Literature survey, Reviewing and Editing

Parvin Kumar: Conceptualization, Methodology, Software, Writing- original draft, Reviewing and Editing

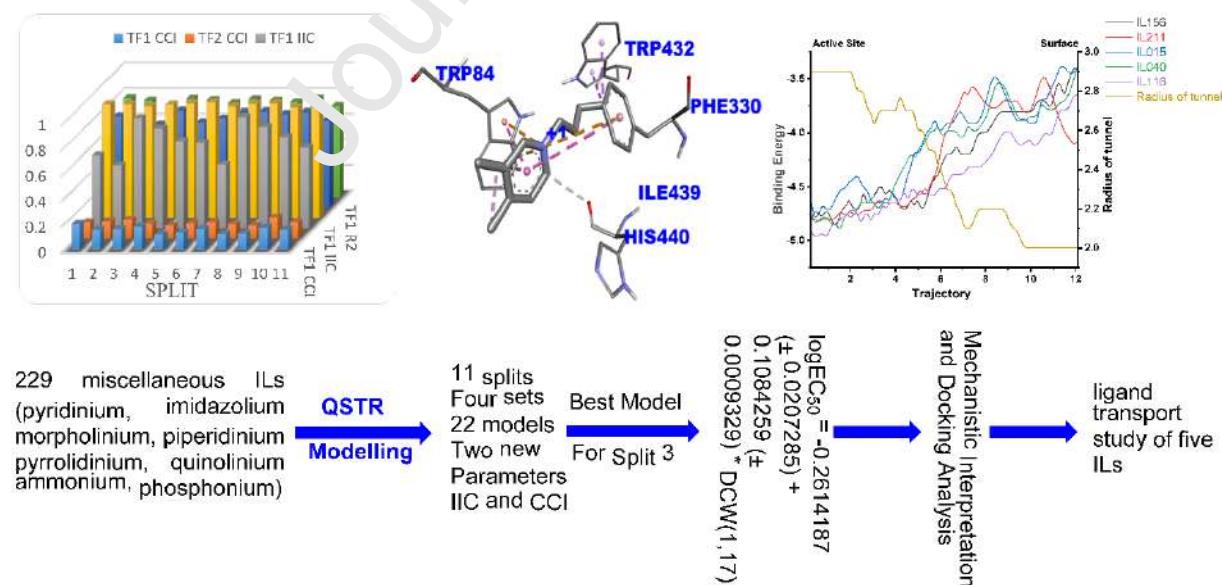
Declaration of interests

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

GRAPHICAL ABSTRACT

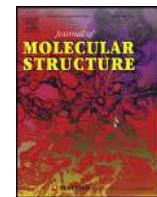
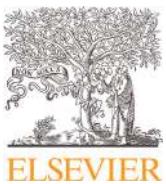
The Correlation Contradictions Index (CCI): Predicting the Toxicity of Ionic Liquids toward Acetylcholinesterase Enzymes Using Novel QSTR Models with Index of Ideality of Correlation

Ashwani Kumar^a, Parvin Kumar^{a,*}



Highlights:

- 22 QSTR models for 229 of ionic liquids are developed from 11 random splits
- Toxicity of ionic liquids are predicted toward acetylcholinesterase Enzymes
- IIC and CCI are examined as a criterion of predictive potential
- The mechanistic interpretation was confirmed by the docking studies
- Ligand transport study of five ILs was studied using the Caver Web Server



Synthesis, Crystal structure and DFT studies of Polyfunctionalized Alkenes: A transition Metal-Free C(sp²)-H Sulfenylation of electron deficient Alkyne

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ABSTRACT

An efficient, novel and transition metal-free protocol has been developed for the synthesis of polyfunctionalized aminothioalkenes *via* direct C–H sulfenylation of *in situ* generated enamines. The reaction was performed using a catalytic amount of inexpensive and nontoxic K₂CO₃ under mild reaction condition. All the reactions resulted in good to excellent yields. The cross-coupling reaction has been achieved by *in situ* aerobic oxidation at room temperature with good functional group tolerance. The molecular architecture and stereochemistry has been established by spectral data, X-ray single crystal diffraction studies and supported by Density Functional theory (DFT). Hirshfeld surface analysis has been used to explore the intramolecular and intermolecular interactions present in the case of **4a**. Moreover, the intramolecular hyperconjugative interactions have been investigated using natural bond orbitals (NBOs) analysis and their intensity was categorized according to their second-order stabilization energy (E(2)). The electrostatic properties such as global reactivity descriptors, local reactivity descriptors, ESP and NLO have been investigated using DFT method and B3LYP/6–311+G(d) level of theory.

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1. Introduction

Development of novel, green and accessible procedures for the synthesis polyfunctionalized functionalities is a worthwhile contribution in organic synthesis. The progression of new C–S cross-dehydrogenative coupling (CDC) strategies attract synthetic organic chemists due to wide application of organosulfur compounds in pharmaceuticals, agrochemicals, organic dyes, and materials chemistry [1–6]. The C–H activation is a class of organic transformations [7–9], wherein the Heck reaction has established itself with high practicality [10,11]. Lately, the cross-coupling (CC) reaction of aryl halides with aryl thiols has become one of the most efficient method for C–S bond formation. A number of CDC reactions have been developed recently to form C_{sp}–C_{sp}, C_{sp}–C_{sp}² and C_{sp}²–C_{sp}² bonds [12–19]. With the focus on emergence of “atom-economy” [20,21] and “green chemistry”[22] in organic synthesis, transition metal-free functionalization and specially, the C–C and C–X bond

formation *via* C–H activation or CDC reactions has become more important.

Several CDC reaction based methodologies have been developed for thiolation/sulfenylation *via* C–H activation which, however, require synthesis of thiolation reagents initially [23–31]. Direct utilization of arylthiols as sulfenylating agent in metal-free protocols have not been explored to a significant extent. Thiolation/sulfenylation of five and six membered heterocycles has been reported using transition metal-free reagents *viz.* I₂/BSA [32], K₂CO₃/DMSO [33], I₂/DMSO [34], N-chlorosuccinimide [35], I₂/DMSO [36], and KClO₃/ethylacetate [37]. Molecular oxygen has been utilized as mild oxidant in organic synthesis by various research groups in C–H oxygenation [38], C–H amination [39] and C–H thiolation of enamines/enaminones [40,41]. To the best of our knowledge, synthesis of polyfunctionalized alkenes *via* CDC mediated sulfenylation of enamines by using environmentally benign method has not been explored yet.

The non-covalent interactions largely affect the molecular architecture by controlling the aggregation process in crystals. [42] The role of strong hydrogen bonds (O–H•••O, N–H•••H, N–H•••O etc.) is very significant in crystal packing [43]. Further, crystal pack-

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Table 1
Optimization of reaction conditions^a.

Entry	Base (equiv.)	Solvent	Time(h)	Yield(4a)
1.	–	CH ₃ CN	24 h	No reaction
2.	K ₂ CO ₃ (2)	CH ₃ CN	24 h	42 ^b
3.	K ₂ CO ₃ (1)	CH ₃ CN	24 h	32 ^b
4.	K ₂ CO ₃ (4)	CH ₃ CN	24 h	42 ^b
5.	K ₂ CO ₃ (4)	EtOH	24 h	42 ^b
6.	K ₂ CO ₃ (4)	THF	24 h	20 ^b
7.	K ₂ CO ₃ (4)	DCM	24 h	36 ^b
8.	K ₂ CO ₃ (4)	DMF	12 h	76
9.	K ₂ CO ₃ (4)	DMSO	09 h	83
10.	NaOH (4)	DMSO	24 h	68
11.	Cs ₂ CO ₃ (4)	DMSO	24 h	78
12.	K ₂ CO ₃ (4)	DMSO	24 h	No reaction ^c

^a Conditions: **1a** (1.0 mmol), **2a** (1.0 mmol) and **3a** (2.0 mmol), base in 5 mL of solvent at room temperature;

^b Incomplete Reaction;

^c reaction was attempted under N₂ atmosphere.

Table 2
Synthesis of polyfunctionalized aminothioalkenes (**4a-4i**).

Compound no	R ¹	R ²	Ar	Time (h)	Yield (%)
4a	OMe	H	C ₆ H ₅	09	83
4b	OMe	H	4-ClC ₆ H ₄	08	82
4c	OMe	H	2-Naphthyl	12	85
4d	Cl	H	4-ClC ₆ H ₄	10	85
4e	Br	H	4-BrC ₆ H ₄	24	78
4f	Me	H	4-MeC ₆ H ₄	12	86
4g	H	H	4-ClC ₆ H ₄	12	81
4h	F	H	4-ClC ₆ H ₄	24	80
4i	Cl	Cl	2-Naphthyl	12	82

ing in molecules devoid of these strong directional forces depends mainly upon weak interactions. The role of -CH group in self-assembly is well established and plays a significant role in crystal packing [44]. DFT based theoretical methods provide an alternative to crystallography for molecular structure prediction. The detailed exploration of structural features from optimized molecular geometry using DFT based methods is one of our interest [45]. [46]

Keeping in view the need of development of novel transition metal-free methodologies and continuing our interest in the same field [47–50], we have carried out metal free C–H sulfenylation of *in situ* prepared enamines and their detailed structural study using X-ray single crystal diffraction studies (XCDS) and DFT methods

2. Results and discussion

2.1. Chemistry

The direct sulfenylation of *in situ* generated enamine was investigated using a one-pot, three-component protocol, where dimethylacetylenedicarboxylate (**1a**), *p*-anisidine (**2a**) and thiophenol (**3a**) were used as model substrates. Firstly, a mixture of dimethylacetylenedicarboxylate (**1a**) (1.0 mmol) and *p*-anisidine (**2a**) (1.0 mmol) was stirred at room temperature in ethanol for 5 min. After the formation of enamine, as monitored by TLC using EtOAc: Pet ether (10:90, v/v) as solvent, ethanol was removed under reduced pressure and 2.0 mmol of thiophenol (**3a**) in 5 mL of CH₃CN was added to the reaction mixture. The contents were stirred at room temperature and the progress of the reaction was monitored using TLC for 24 h. No new product formation was observed in the reaction even after 24 h (Table 1, entry 1) (Scheme 1). The same model reaction was further explored using K₂CO₃ (2 equiv.) as a base. A new spot was observed on TLC, however, the reaction was incomplete even after 24 h. The reaction was quenched by adding ice. The reaction mixture was extracted using DCM, dried over anhyd. Na₂SO₄ and

chromatographed over silica gel (230–400) using EtOAc: Pet ether (2:98, v/v) as eluent. The solid thus separated was characterized as dimethyl 2-((4-methoxyphenyl)amino)-3-(phenylthio)fumarate (**4a**) using ¹H NMR, ¹³C NMR and X-ray single crystal diffraction studies (XCDS) and was obtained in 42% yield (Table 1, entry 2).

In order to develop a highly efficient and convenient methodology for the synthesis of aminothioalkenes (**4**), the scope of the same model reaction was further explored using different concentration of base i.e., 1 eq. and 4 eq. of K₂CO₃ (Table 1, entries 3 & 4). It was observed that reaction went to completion with high yield when higher concentration of base was used (Table 1, entry 4).

A variety of solvents i.e., EtOH, THF, DCM, DMF and DMSO were explored in the same model reaction using K₂CO₃ (4 equiv.) as a base (Table 1, entry 5–9). In first three cases, reactions did not proceed to completion even after 24 h and gave 42%, 20% and 36% of **4a**, respectively (Table 1, entries 5–7) while in case of DMF, the reaction was complete in 12 h but gives **4a** in 76% yield (Table 1, entry 8). To our delight, the reaction carried out in DMSO at room temperature under ideal conditions resulted in 83% yield of **4a** in 9 h (Table 1, entry 9). The effect of different bases like NaOH and Cs₂CO₃ (Table 1, entries 10 & 11) was also explored. However, no significant improvement in yield and reaction time was observed in both the conditions. In order to explore the role of air, the reaction was carried out under N₂ atmosphere and no desired product (**4a**) were detected (Table 1, entry 12). It can be inferred that thiol **3a** undergoes aerobic oxidation *in situ* to disulfide. This shows that the reactions occurring via an aerobic oxidative cross-coupling.

It can be inferred from above results that the reaction of dimethylacetylenedicarboxylate (**1a**) (1 mmol), *p*-anisidine (**2a**) (1 mmol) and thiophenol (**3a**) (2 mmol) in presence of 4 equiv. of K₂CO₃ using DMSO as solvent is the standardized reaction condition for the synthesis of dimethyl 2-(phenylthio)-3-((4-methoxyphenyl)amino)fumarate (**4a**).

The developed methodology was then extended to other substrates by carrying out reactions of dimethylacetylenedicarboxylate (**1a**) with substituted thiophenol and aniline under otherwise identical conditions (Scheme 2). All the reaction gave the corresponding polyfunctionalized alkenes in high yields in 8–24 h under similar condition (**4b–4i**). The developed methodology failed to yield desired product when aliphatic thiol (*n*-hexanethiol) was reacted with *in situ* generated enamine under identical conditions.

We believe that the thioarylation proceeds by a nucleophilic attack of *in situ* generated enamine on diphenyldisulfide linkage followed by isomerization to give the desired product (Scheme 3). The initial formation of enamine from dialkyl acetylenedicarboxylate and aromatic amine in ethanol was followed by removal of ethanol under reduced pressure. This was followed by addition of 2 eq. of thiophenol which undergoes aerial oxidation in K₂CO₃/DMSO, followed by attack of enamine on disulphide to give the desired compound polyfunctionalized aminothioalkenes.

2.2. Molecular structure (X-ray diffraction)

The detailed molecular structure of dimethyl 2-((4-methoxyphenyl)amino)-3-(phenylthio)fumarate (**4a**) was explored using XCDS. The *trans* stereochemistry of double bond in **4a** was confirmed from the structure derived from diffraction studies (Fig. 1). The ORTEP view of the asymmetric unit along with the optimized structure of **4a** are shown in Fig. 1. The refinement details and crystallographic parameters are provided as supplementary material (Table S1). The molecule **4a** crystallises in monoclinic system within *p*21/n space group with lattice parameter *a* = 10.8877(2) Å, *b* = 20.6639(3) Å, *c* = 8.1934(8) Å with β = 90.502(3)° respectively.

Dimethyl 2-((4-chlorophenyl)thio)-3-((4-fluorophenyl)amino)fumarate (4 h)

Off white solid; ^1H NMR (400 MHz, CDCl_3) δ_{H} 10.79 (s, 1H, NH), 7.27–7.12 (m, 6H, ArH), 7.03 (t, J = 8.5 Hz, 2H, ArH), 3.72 (s, 3H, COOCH_3), 3.63 (s, 3H, COOCH_3); ^{13}C NMR (100 MHz, CDCl_3) δ_{C} 170.34, 162.74, 161.03(d, $^1J_{\text{C}-\text{F}} = 246$ Hz), 159.39, 137.01, 134.14, 131.38, 128.77, 127.72, 125.33, 116.26 (d, $^2J_{\text{C}-\text{F}} = 24.7$ Hz), 85.57, 52.54, 52.15.

Dimethyl

2-((3,4-dichlorophenyl)amino)-3-(naphthalen-2-ylthio)fumarate(4i)

Off white solid; ^1H NMR (400 MHz, CDCl_3) δ_{H} 10.95 (s, 1H, NH), 7.81–7.72 (m, 3H, ArH), 7.66 (s, 1H, ArH), 7.49–7.35 (m, 4H, ArH), 7.29 (d, J = 2.7 Hz, 1H, ArH), 7.02 (dd, J = 8.7, 2.7 Hz, 1H, ArH), 3.71 (s, 6H, 2 X COOCH_3); ^{13}C NMR (100 MHz, CDCl_3) δ_{C} 170.65, 163.05, 158.27, 137.49, 135.77, 133.82, 132.68, 131.82, 128.41, 127.83, 127.26, 126.50, 125.46, 124.91, 124.41, 124.12, 119.56, 87.16, 52.87, 52.45.

5.2 X-ray crystal studies

The single crystal of compound **4a** suitable for X-ray analysis was obtained by slow evaporation method using acetonitrile as solvent. Single, clear, whitish block of single crystal of **4a** suitable for X-ray was mounted on Xcalibur, Sapphire3 diffractometer using mylar loop. The data collection was done at a steady temperature of $T = 298$ K. The structure was solved using Olex2 [64] and the model was refined with ShelXL using full matrix least squares minimisation on F^2 [65]. The final completeness is 100% out to 29.556° in Θ . A multi-scan absorption correction was performed using CrysAlisPro 1.171.38.46 (Rigaku Oxford Diffraction, 2015). Empirical absorption correction using spherical harmonics, implemented in SCALE3 ABSPACK scaling algorithm. The absorption coefficient μ of this material is 0.205 mm^{-1} at this wavelength ($\lambda = 0.71073 \text{ \AA}$) and the minimum and maximum transmissions are 0.796 and 1.000. Crystal explorer 17.5 was used for Hirshfeld surface generation and 2d fingerprint analysis [66]. Crystallography data excluding structure factors has been deposited on Cambridge crystallography database with CCDC no. **1,993,445** for **4a**.

5.3 Computational details of dft studies

All DFT calculations presented in the present manuscript were performed with Gaussian 09 program package [67] using hybrid exchange correlation functional B3LYP and 6-311(+G(d) basis set [68]. Initially, the geometry of all the polyfunctionalized aminothiols (**4a-4i**) was optimized using same level of theory. No imaginary frequencies were found for any of the structure, which indicates their stability at global minima. Density of states (DOS) were calculated using Gausssum 3.0. [69] After optimization of the molecular geometries, the global reactivity descriptors were calculated utilizing the information contained in FMO's [70,71]. The NBO calculations were performed using NBO 3.0 program implemented in Gaussian 09 W package using the same level of theory to explore the hyperconjugative interaction present in the molecule [72]. Thereafter, Fukui function were calculated from same NBO analysis at same level of theory to explore the possibility of charge transfer in polyfunctionalized aminothiols (**4a-4i**). Avogardo 2.0 was used for the visualization of the results of DFT calculations. [73]

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:[10.1016/j.molstruc.2020.129089](https://doi.org/10.1016/j.molstruc.2020.129089).

References

- [1] I.P. Beletskaya, V.P. Ananikov, Transition-metal-catalyzed C-S, C-Se, and C-te bond formation via cross-coupling and atom-economic addition reactions, Chem. Rev 111 (2011) 1596–1636, doi:[10.1021/cr100347k](https://doi.org/10.1021/cr100347k).
- [2] G. La Regina, A. Coluccia, A. Brancale, F. Piscitelli, V. Gatti, G. Maga, A. Samuele, C. Pannecouque, D. Schols, J. Balzarini, Indolylarylsulfones as HIV-1 non-nucleoside reverse transcriptase inhibitors: new cyclic substituents at indole-2-carboxamide, J. Med. Chem. 54 (2011) 1587–1598.
- [3] T. Punniyamurthy, S. Velusamy, J. Iqbal, Recent advances in transition metal catalyzed oxidation of organic substrates with molecular oxygen, Chem. Rev 105 (2005) 2329–2364.
- [4] M. Klečka, R. Pohl, J. Čejka, M. Hocek, Direct C–H sulenylation of purines and deazapurines, Org. Biomol. Chem. 11 (2013) 5189–5193.
- [5] I.M. Yonova, C.A. Osborne, N.S. Morrisette, E.R. Jarvo, Diaryl and heteroaryl sulfides: synthesis via sulenyl chlorides and evaluation as selective anti-breast-cancer agents, J. Org. Chem. 79 (2014) 1947–1953.
- [6] T. Kondo, T. Mitsudo, Metal-catalyzed carbon–sulfur bond formation, Chem. Rev 100 (2000) 3205–3220.
- [7] S. Tang, K. Liu, C. Liu, A. Lei, Olefinic C–H functionalization through radical alkenylation, Chem. Soc. Rev 44 (2015) 1070–1082.
- [8] S.I. Kozhushkov, L. Ackermann, Ruthenium-catalyzed direct oxidative alkenylation of arenes through twofold C–H bond functionalization, Chem. Sci. 4 (2013) 886–896.
- [9] D.-H. Wang, K.M. Engle, B.-F. Shi, J.-Q. Yu, Ligand-enabled reactivity and selectivity in a synthetically versatile aryl C–H olefination, Science (80–.) 327 (2010) 315–319.
- [10] A.B. Douay, L.E. Overman, The asymmetric intramolecular Heck reaction in natural product total synthesis, Chem. Rev 103 (2003) 2945–2964.
- [11] D. Mc Cartney, P.J. Guiry, The asymmetric Heck and related reactions, Chem. Soc. Rev 40 (2011) 5122–5150.
- [12] G. Dyker, Handbook of CH transformations: applications in organic synthesis, Wiley-VCH, 2005.
- [13] T.W. Lyons, M.S. Sanford, Palladium-catalyzed ligand-directed C–H functionalization reactions, Chem. Rev 110 (2010) 1147–1169.
- [14] D.A. Colby, R.G. Bergman, J.A. Ellman, Rhodium-catalyzed C–H bond activation via heteroatom-directed C–H bond activation, Chem. Rev 110 (2009) 624–655.
- [15] X. Chen, K.M. Engle, D. Wang, J. Yu, Palladium (II)-catalyzed C–H activation/C–C cross-coupling reactions: versatility and practicality, Angew. Chem. 48 (2009) 5094–5115.
- [16] J. Wencel-Delord, T. Droege, F. Liu, F. Glorius, Towards mild metal-catalyzed C–H bond activation, Chem. Soc. Rev 40 (2011) 4740–4761.
- [17] S.H. Cho, J.Y. Kim, J. Kwak, S. Chang, Recent advances in the transition metal-catalyzed twofold oxidative C–H bond activation strategy for C–C and C–N bond formation, Chem. Soc. Rev 40 (2011) 5068–5083.
- [18] C.-L. Sun, B.-J. Li, Z.-J. Shi, Direct C–H transformation via iron catalysis, Chem. Rev 111 (2010) 1293–1314.
- [19] L. Ackermann, Carboxylate-assisted transition-metal-catalyzed C–H bond functionalizations: mechanism and scope, Chem. Rev 111 (2011) 1315–1345.
- [20] B.M. Trost, Atom economy—A challenge for organic synthesis: homogeneous catalysis leads the way, Angew. Chem 34 (1995) 259–281.
- [21] B.M. Trost, The atom economy—a search for synthetic efficiency, Science (80–.) 254 (1991) 1471–1477.
- [22] N. Winterton, Twelve more principles of green chemistry, Green Chem. 3 (2001) G73–G75.
- [23] F. Yang, S. Tian, Iodine-Catalyzed Regioselective Sulfenylation of Indoles with Sulfonyl Hydrazines, Angew. Chem. 52 (2013) 4929–4932.
- [24] X. Zhao, L. Zhang, T. Li, G. Liu, H. Wang, K. Lu, P-Toluenesulphonic acid-promoted, I2-catalysed sulphenylation of pyrazolones with aryl sulphonyl hydrazides, Chem. Commun. (2014). 10.1039/c4cc05237d.
- [25] X. Kang, R. Yan, G. Yu, X. Pang, X. Liu, X. Li, L. Xiang, G. Huang, Iodine-mediated thiolation of substituted naphthols/naphthylamines and arylsulfonyl hydrazides via C (sp²)-H bond functionalization, J. Org. Chem. 79 (2014) 10605–10610.
- [26] Q. Wu, D. Zhao, X. Qin, J. Lan, J. You, Synthesis of di(hetero)aryl sulfides by directly using arylsulfonyl chlorides as a sulfur source, Chem. Comm.. (2011). 10.1039/c1cc13633j.
- [27] F. Xiao, H. Xie, S. Liu, G. Deng, Iodine-Catalyzed Regioselective Sulfenylation of Indoles with Sodium Sulfinate, Adv. Synth. Catal. 356 (2014) 364–368.
- [28] W. Ge, Y. Wei, Iodine-catalyzed oxidative system for 3-sulfenylation of indoles with disulfides using DMSO as oxidant under ambient conditions in dimethyl carbonate, Green Chem. 14 (2012) 2066–2070.
- [29] W. Ge, X. Zhu, Y. Wei, Iodine-Catalyzed Selective Synthesis of 2-Sulfanylphenols via Oxidative Aromatization of Cyclohexanones and Disulfides, Adv. Synth. Catal. 355 (2013) 3014–3021.

- [30] S. Guo, Y. Yuan, J. Xiang, Metal-free oxidative C (sp³)-H bond thiolation of ethers with disulfides, *Org. Lett.* 15 (2013) 4654–4657.
- [31] B. Du, B. Jin, P. Sun, Syntheses of Sulfides and Selenides through Direct Oxidative Functionalization of C (sp³)-H Bond, *Org. Lett.* 16 (2014) 3032–3035.
- [32] D. Equbal, A.G. Lavekar, A.K. Sinha, Cooperative catalysis by bovine serum albumin-iodine towards cascade oxidative coupling-C(sp²)-H sulfenylation of indoles/hydroxyaryls with thiophenols on water, *Org. Biomol. Chem.* 14 (2016) 6111–6118.
- [33] P. Sang, Z. Chen, J. Zou, Y. Zhang, K₂CO₃ promoted direct sulfenylation of indoles: a facile approach towards 3-sulfenylindoles, *Green Chem.* 15 (2013) 2096–2100.
- [34] S.K.R. Parumala, R.K. Peddinti, Iodine catalyzed cross-dehydrogenative C-S coupling by C(sp²)-H bond activation: direct access to aryl sulfides from aryl thiols, *Green Chem.* 17 (2015) 4068–4072.
- [35] G. Khalili, A new synthesis of \$-\$varvec \$-\$aryl uracils from aryl thiols and 6-amino uracils in the presence of NC, *Mol. Divers.* 20 (2016) 963–968.
- [36] Y. Siddaraju, K.R. Prabhu, Iodine Promoted Regioselective α -Sulfonylation of Carbonyl Compounds using Dimethyl Sulfoxide as an Oxidant, *Org. Lett.* 18 (2016) 6090–6093, doi:10.1021/acs.orglett.6b03084.
- [37] J.-P. Wan, S. Zhong, L. Xie, X. Cao, Y. Liu, L. Wei, KIO₃-catalyzed aerobic cross-coupling reactions of enamines and thiophenols: synthesis of polyfunctionalized alkenes by metal-free C-H sulfenylation, *Org. Lett.* 18 (2016) 584–587.
- [38] X. Liu, R. Cheng, F. Zhao, D. Zhang-Negrerie, Y. Du, K. Zhao, Direct β -acyloxylation of enamines via PhIO-mediated intermolecular oxidative C-O bond formation and its application to the synthesis of oxazoles, *Org. Lett.* 14 (2012) 5480–5483.
- [39] Y. Yuan, W. Hou, D. Zhang-Negrerie, K. Zhao, Y. Du, Direct oxidative coupling of enamines and electron-deficient amines: tBAI/TBHP-mediated synthesis of substituted diaminoalkenes under metal-free conditions, *Org. Lett.* 16 (2014) 5410–5413.
- [40] S. Zhong, Y. Liu, X. Cao, J.-P. Wan, KIO₃-Catalyzed Domino C(sp²)-H Bond Sulfenylation and C–N Bond Oxygenation of Enaminones toward the Synthesis of 3-Sulfenylated Chromones, *ChemCatChem* 9 (2017) 465–468, doi:10.1002/cctc.201601273.
- [41] Y. Siddaraju, K.R. Prabhu, Iodine-Catalyzed Cross Dehydrogenative Coupling Reaction: sulfenylation of Enaminones Using Dimethyl Sulfoxide as an Oxidant, *J. Org. Chem.* 82 (2017) 3084–3093, doi:10.1021/acs.joc.7b00073.
- [42] A.J. Rybarczyk-Pirek, M. Łukomska-Rogala, S. Wojtulewski, M. Palusiak, N-Oxide as a Proton Accepting Group in Multicomponent Crystals: x-ray and Theoretical Studies on New p-Nitropyridine-N-oxide Co-Crystals, *Cryst. Growth Des.* 15 (2015) 5802–5815, doi:10.1021/acs.cgd.5b01177.
- [43] J. Hernández-Paredes, R.C. Carrillo-Torres, A.A. López-Zavala, R.R. Sotelo-Mundo, O. Hernández-Negrete, J.Z. Ramírez, M.E. Alvarez-Ramos, Molecular structure, hydrogen-bonding patterns and topological analysis (QTAIM and NCI) of 5-methoxy-2-nitroaniline and 5-methoxy-2-nitroaniline with 2-amino-5-nitropyridine (1:1) co-crystal, *J. Mol. Struct.* 1119 (2016) 505–516, doi:10.1016/j.molstruc.2016.05.012.
- [44] E. Bosch, N.P. Bowling, J. Darko, The Power of Nonconventional Phenyl C–H•••N Hydrogen Bonds: supportive Crystal-Packaging Force and Dominant Supramolecular Engineering Force, *Cryst. Growth Des.* 15 (2015) 1634–1641, doi:10.1021/cg5014076.
- [45] G. Singh, J. Sindhu, V.Kumar Manisha, V. Sharma, S.K. Sharma, S.K. Mehta, M.H. Mahnashi, A. Umar, R. Kataria, Development of an off-on selective fluorescent sensor for the detection of Fe³⁺ ions based on Schiff base and its Hirshfeld surface and DFT studies, *J. Mol. Liq.* (2019) 296, doi:10.1016/j.molliq.2019.111814.
- [46] R. Kataria, D. Vashisht, J. Sindhu, S. Sharma, S.K. Mehta, R. Kumar, S.C. Sahoo, S. kumar, F. Qu, F.A. Afkhami, A. Gupta, Crystal structure, Hirshfeld surface, DFT and BSA binding studies of dihydropyrazole-1-thiocarboxamides, *J. Mol. Struct.* 1196 (2019) 662–675, doi:10.1016/j.molstruc.2019.06.100.
- [47] A. Chaudhary, J.M. Khurana, G. Khanna, M. Saroha, A Catalyst-Free Domino Protocol for the Chemoselective Synthesis of Multifunctionalised Pyrroles in Aqueous Media via Nitroketene-N, S-Acetal Chemistry, *ChemistrySelect* 3 (2018) 6334–6337.
- [48] M. Saroha, K. Meena, J.M. Khurana, PPh₃ Mediated Stereoselective Synthesis of 4-Fumarate Substituted 3-Acylicoumarins: a Cascade Reaction of 3-Acylic Coumarin with Alkyne Derivatives, *Chem.Select* 3 (2018) 5905–5909.
- [49] M. Saroha, G. Bartwal, J.M. Khurana, Transition metal free K₂CO₃ mediated thioarylation, selenoarylation and arylation of 2-aminomaleimides at ambient temperature, *Tetrahedron* 75 (2019) 130486.
- [50] H. Singh, J. Sindhu, J.M. Khurana, Synthesis of biologically as well as industrially important 1,4,5-trisubstituted-1,2,3-triazoles using a highly efficient, green and recyclable DBU-H<inf>2</inf>O catalytic system, *RSC Adv.* 3 (2013), doi:10.1039/c3ra4440f.
- [51] M. Nardelli, PARST95 - an update to PARST: a system of Fortran routines for calculating molecular structure parameters from the results of crystal structure analyses, *J. App. Cryst.* 28 (1995) 659, doi:10.1107/S0021889895007138.
- [52] R. Taylor, O. Kennard, Comparison of X-ray and neutron diffraction results for the N-H ... O=C hydrogen bond, *Acta Crystallogr. Sect. B* 39 (1983) 133–138, doi:10.1107/S0108768183002116.
- [53] J. Martinez, Local reactivity descriptors from degenerate frontier molecular orbitals, *Chem. Phys. Lett.* 478 (2009) 310–322, doi:10.1016/j.cplett.2009.07.086.
- [54] M.N. Arshad, A.-A.M. Al-Dies, M. Asiri, M. Khalid, A.S. Birinji, K.A. Al-Amry, A.A.C. Braga, Synthesis, crystal structures, spectroscopic and nonlinear optical properties of chalcone derivatives: a combined experimental and theoretical study, *J. Mol. Struct.* 1141 (2017) 142–156, doi:10.1016/j.molstruc.2017.03.090.
- [55] F. Zielinski, V. Tognetti, L. Joubert, Condensed descriptors for reactivity: a methodological study, *Chem. Phys. Lett.* 527 (2012) 67–72, doi:10.1016/j.cplett.2012.01.011.
- [56] T.B. Tai, V.T.T. Huong, M.T. Nguyen, Theoretical Design of π -Conjugated Heterocyclic Compounds Containing a Tricoordinated Boron Center, *J. Phys. Chem. C* 117 (2013) 14999–15008, doi:10.1021/jp4049154.
- [57] P.K. Chattaraj, Chemical Reactivity and Selectivity: Local HSAB Principle versus Frontier Orbital Theory, *J. Phys. Chem. A* 105 (2001) 511–513, doi:10.1021/jp003786w.
- [58] R. Balawender, P. Geerlings, DFT-based chemical reactivity indices in the Hartree-Fock method. II. Fukui function, chemical potential, and hardness, *J. Chem. Phys.* 123 (2005) 124103, doi:10.1063/1.2012330.
- [59] L.R. Domingo, E. Chamorro, P. Pérez, Understanding the reactivity of captodative ethylenes in polar cycloaddition reactions. A theoretical study, *J. Org. Chem.* (2008). 10.1021/jo800572a.
- [60] C. Morell, A. Grand, A. Toro-Labbé, New Dual Descriptor for Chemical Reactivity, *J. Phys. Chem. A* 109 (2005) 205–212, doi:10.1021/jp046577a.
- [61] P. Sjöberg, P. Politzer, Use of the electrostatic potential at the molecular surface to interpret and predict nucleophilic processes, *J. Phys. Chem.* 94 (1990) 3959–3961.
- [62] F. Weinhold, C.R. Landis, E.D. Glendening, What is NBO analysis and how is it useful? *Int. Rev. Phys. Chem.* 35 (2016) 399–440, doi:10.1080/0144235X.2016.1192262.
- [63] M. Nakano, H. Fujita, M. Takahata, K. Yamaguchi, Theoretical study on second hyperpolarizabilities of phenylacetylene dendrimer: toward an understanding of structure-property relation in NLO responses of fractal antenna dendrimers, *J. Am. Chem. Soc.* 124 (2002) 9648–9655, doi:10.1021/ja0115969.
- [64] O.V. Dolomanov, L.J. Bourhis, R.J. Gildea, J.A.K. Howard, H. Puschmann, OLEX2: a complete structure solution, refinement and analysis program, *J. Appl. Crystallogr.* 42 (2009) 339–341, doi:10.1107/S0021889808042726.
- [65] G.M. Sheldrick, Crystal structure refinement with SHELXL, *Acta Crystallogr. Sect. C Struct. Chem.* 71 (2015) 3–8, doi:10.1107/S2053229614024218.
- [66] S.K. Wolff, D.J. Grimwood, J.J. McKinnon, M.J. Turner, D. Jayatilaka, M.A. Spackman, CrystalExplorer (Version 3.1), Univ. West. Aust. (2012).
- [67] M.J. Frisch, G.W. Trucks, H.B. Schlegel, G.E. Scuseria, M.A. Robb, J.R. Cheeseman, G. Scalmani, V. Barone, B. Mennucci, G.A. Petersson, Gaussian 09, rev. (2009).
- [68] P.M.W. Gill, B.G. Johnson, J.A. Pople, M.J. Frisch, The performance of the Becke–Lee–Yang–Parr (B–LVP) density functional theory with various basis sets, *Chem. Phys. Lett.* 197 (1992) 499–505.
- [69] N.M. O’Boyle, A.L. Tenderholt, K.M. Langner, Cclib: a library for package-independent computational chemistry algorithms, *J. Comput. Chem.* (2008). 10.1002/jcc.20823.
- [70] L.R. Domingo, M. Ríos-Gutiérrez, P. Pérez, Applications of the conceptual density functional theory indices to organic chemistry reactivity, *Molecules*. (2016). 10.3390/molecules21060748.
- [71] L.R. Domingo, Molecular electron density theory: a modern view of reactivity in organic chemistry, *Molecules*. (2016). 10.3390/molecules2101319.
- [72] E.D. Glendening, A.E. Reed, J.E. Carpenter, F. Weinhold, NBO 3.0 Program Manual, Theor. Chem. Institute, Univ. Wisconsin, Madison, WI, 1990.
- [73] M.D. Hanwell, D.E. Curtis, D.C. Lonie, T. Vandermeersch, E. Zurek, G.R. Hutchison, Avogadro: an advanced semantic chemical editor, visualization, and analysis platform, *J. Cheminfo* 4 (2012) 17.



A review of antimalarial activity of two or three nitrogen atoms containing heterocyclic compounds

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Abstract

Malaria, a nocuous disease, which has become a major challenge for the health resulting in deaths of millions of people around the globe. Malaria is a parasitic disease propagated by mosquitoes and infects the human beings. Several species of *Plasmodium* are responsible for this life-threatening disease and *Plasmodium falciparum* being the most virulent. In order to eradicate the malarial parasite, the researchers are making consistent efforts in synthesizing new antimalarial drug candidates by paying attention to the various drug targets. In this manuscript, the main focus is on the antimalarial activity of numerous heterocyclic compounds reported by the researchers since 2010 against the different strains of *Plasmodium*. Antimalarial activities of the two and three nitrogen-containing heterocycles along with their structure–activity relationship are described.

Keywords Malaria · *Plasmodium falciparum* · Heterocycles · Antimalarial activity · Structure–activity relationship

Introduction

Malaria, a vector-borne infectious disease, is one of the utmost annihilating diseases of the ever-changing world predominated chiefly in tropical regions (Ridley 2002). The World Health Organization evaluated 219 million malaria cases across the entire globe, an increment of 2 million from the preceding year and 435 thousand quietus at annual frequency as well as 1190 on daily occurrence, predominantly moppets (Tse et al. 2019). Six species of single-celled eukaryotic *Plasmodium* parasites are the causative agents of malaria: *P. knowlesi*, *P. vivax*, *P. ovale curtisi*, *P. ovale wallikeri*, *P. malariae*, and *P. falciparum*. Out of all these, *P. falciparum* is liable for ~90% of the infections (Lee et al. 2019). *Plasmodium falciparum* is the most noxious and has the highest rates of complications, mortality as well as the prevalence of erythrocytic disorders globally (Buffet et al. 2011). *Plasmodium* is an obligate intracellular parasite that requires two hosts for the

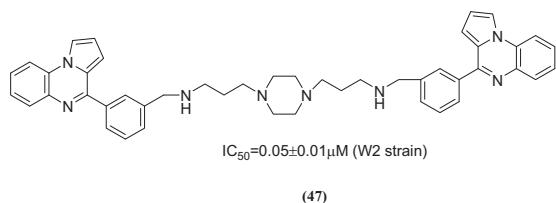
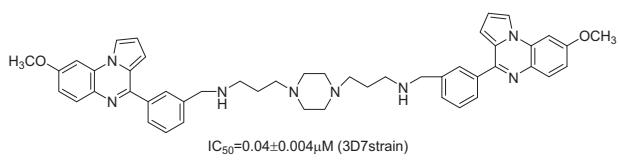
completion of its life process, i.e., for sexual life cycle—an arthropod vector and asexual life cycle-human host (Jensen et al. 2020). Administration of the transmissive form i.e., the sporozoite into the human skin with the assistance of the female *Anopheles* mosquito with an anticoagulant saliva prior to the blood meal. Sporozoites intrude the lymphoid system and get dispersed to defile the liver. In the liver, they turn out to be intracellular and escalate to embody hundreds of merozoites that are actually invasive (the pre- or exo-erythrocytic phase). Merozoites are discharged into the blood stream and assail the erythrocytes. The parasite *Plasmodium* nourishes on its host cell within an erythrocyte, thereafter proliferates to form corresponding merozoites which move out and invade novel erythrocytes, the cycle reoccurs. There is a variation in time within different species from invasion to exit, 48 h for *P. vivax* and *P. falciparum* and 72 h for *P. malariae* and *P. ovale*, the contemporaneous release of merozoites coexisting with fever peaks. Ultimately a sexual phase commences where the parasite develops intrinsically in its host cell into either a macrogametocyte i.e., a female gametocyte or a male gametocyte i.e., a microgametocyte. The reliance on the continuation of the life process is on gametocytes being carried into the feeding female mosquito's gut where twain of gametocytes flee from their host cells. Male gametocytes segregate abruptly into numerous locomotive whip-like microgametes, which can individually fecundate a female macrogamete for the formation of a zygote. The parasite

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susceptibility strain as well as $IC_{50} = 0.05 \pm 0.01 \mu\text{M}$ for chloroquine-resistant strain (Guillon et al. 2017).



Conclusion

Malaria is a disease which posed a great burden on the human beings becoming the cause of mortality of millions of people all over the world. In order to obliterate the parasite responsible for this disease and simultaneous failure of the conventional antimalarial drugs led to the modification of already available drugs as well as development of novel drugs. In this manuscript, a comprehensive review of the two and three nitrogen-containing heterocycles since 2010 has been described. Diverse heterocyclic scaffolds were utilized for the evolution of more drugs with an enhanced bioavailability and improved physicochemical properties. The activity of numerous compounds was evaluated against various *Plasmodium falciparum* strains such as chloroquine-sensitive, chloroquine-resistant as well as multidrug-resistant strains. More and more novel compounds with an improved pharmacokinetic profile would be synthesized which would be more effective, economic as well as safe for human use. These hybrids would also possess least toxicity and a greater antimalarial potency.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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References

- Achan J, Talisuna AO, Erhart A, Yeka A, Tibenderana JK, Baliraine FN, Rosenthal PJ, D'Alessandro U (2011) Quinine, an old anti-malarial drug in a modern world: role in the treatment of malaria. *Malar J* 10:144. <https://doi.org/10.1186/1475-2875-10-144>
- Aggarwal S, Paliwal D, Kaushik D (2018) Pyrazole schiff base hybrids as antimarial agents: synthesis, in vitro screening and computational study. *Comb Chem High Throughput Screen* 21:194–203
- Alano P (2007) *Plasmodium falciparum* gametocytes: still many secrets of a hidden life. *Mol Microbiol* 66:291–302
- Amusengeri A, Tata RB, Bishop OT (2020) Understanding the pyrimethamine drug resistance mechanism via combined molecular dynamics and dynamic residue network analysis. *Molecules* 25:904. <https://doi.org/10.3390/molecules25040904>
- Araujo NCP, Barton V, Jones M, Stocks PA, Ward SA, Davies J, Bray PG, Shone AE, Cristiano MLS, O'Neill PM (2009) Semi-synthetic and synthetic 1,2,4-trioxaquines and 1,2,4-trioxolaquines: synthesis, preliminary structure activity relationship and comparison with acridine endoperoxide conjugates. *Bioorg Med Chem Lett* 19:2038–2043
- Asif M (2017) A mini review: biological significances of nitrogen hetero atom containing heterocyclic compounds. *Int J Bioorg Chem* 2:146–152
- Awan SJ, Hadi F, Fayyaz M, Malik S, Maqbool T (2018) Therapeutic diversification of azoles and their derivatives. *Int J Dev Res* 8:23890–23894
- Bhat HR, Singh UP, Gahtori P, Ghosh SK, Gogoi K, Prakash A, Singh RK (2013a) Antimalarial activity and docking studies of novel bi-functional hybrids derived from 4-aminoquinoline and 1,3,5-triazine against wild and mutant malaria parasites as Pf-DHFR inhibitor. *RSC Adv* 3:2942–2952
- Bhat HR, Singh UP, Gahtori P, Ghosh SK, Gogoi K, Prakash A, Singh RK (2013b) 4-Aminoquinoline-1,3,5-triazine: design, synthesis, in vitro antimalarial activity and docking studies. *N J Chem* 37:2654–2662
- Brack W, Schirmer K (2003) Effect-directed identification of oxygen and sulfur heterocycles as major polycyclic aromatic cytochrome P4501A-inducers in a contaminated sediment. *Environ Sci Technol* 37:3062–3070
- Briolant S, Fusai T, Rogier C, Pradines B (2008) Tetracycline antibiotics in malaria. *Open Trop Med J* 1:31–46
- Broughton HB, Watson IA (2004) Selection of heterocycles for drug design. *J Mol Graph Model* 23:51–58
- Buffet PA, Safeukui I, Deplaine G, Brousse V, Prendki V, Thellier M, Turner GD, Pujalon OM (2011) The Pathogenesis of *Plasmodium falciparum* malaria in humans: insights from splenic physiology. *Blood* 117:381–392
- Cabrera DG, Manach CL, Douelle F, Younis Y, Feng TS, Paquet T, Nchinda AT, Street LJ, Taylor D, Cd Koch, Wiesner L, Duffy S, White KL, Zabiulla KM, Sambandan Y, Bashyam S, Waterson D, Witty MJ, Charman SA, Avery VM, Wittlin S, Chibale K (2014) 2,4-Diaminothienopyrimidines as orally active antimalarial agents. *J Med Chem* 57:1014–1022
- Cairns M, Cisse B, Sokhna C, Cames C, Simondon K, Ba EH, Trape JF, Gaye O, Greenwood BM, Milligan PJM (2010) Amodiaquine dosage and tolerability for intermittent preventive treatment to prevent malaria in children. *Antimicrob Agents Chemother* 54:1265–1274
- Cheng G, Sa W, Cao C, Guo L, Hao H, Liu Z, Wang X, Yuan Z (2016) Quinoxaline 1,4-di-N-oxides: biological activities and mechanisms of actions. *Front Pharm* 7:64. <https://doi.org/10.3389/fphar.2016.00064>

- Chu XM, Wang C, Wang WL, Liang LL, Liu W, Gong KK, Sun KL (2019) Triazole derivatives and their antiplasmodial and antimarial activities. *Eur J Med Chem* 166:206–223
- Dua VK, Sinha SN, Biswas S, Valecha N, Puri SK, Sharma VK (2002) Isolation and antimarial activity of peroxydisulfate oxidation products of primaquine. *Bioorg Med Chem Lett* 12:3587–3589
- Davis TME, Hung TY, Sim IK, Karunajeewa HA, Ilett KF (2005) Piperaquine. *Drugs* 65:75–87
- Ezzet F, Vugt VM, Nosten F, Looareesuwan S, White NJ (2000) Pharmacokinetics and pharmacodynamics of lumefantrine (benflumetol) in acute falciparum malaria. *Antimicrob Agents Chemother* 44:697–704
- Fernando D, Rodrigo C, Rajapakse S (2011) Primaquine in vivax malaria: an update and review on management issues. *Malar J* 10:1–12
- Filho JMDS, Silva DMADQe, Macedo TS, Teixeira HMP, Moreira DRM, Challal S, Wolfender JL, Queiroz EF, Soares MBP (2016) Conjugation of N-acylhydrazone and 1,2,4-oxadiazole leads to the identification of active antimalarial agents. *Bioorg Med Chem* 24:5693–5701
- Faidallah HM, Panda SS, Serrano JC, Grgis AS, Khan KA, Alamry KA, Therathanakorn T, Meyers MJ, Sverdrup FM, Eickhoff CS, Getchell SG, Katritzky (2016) Synthesis, antimarial properties and 2D-QSAR studies of novel triazole-quinine conjugates. *Bioorg Med Chem* 24:3527–3539
- Frohlich T, Reiter C, Ibrahim MM, Beutel J, Hutterer C, Zeitrager I, Bahsi H, Leidenberger M, Friedrich O, Kappes B, Efferth T, Marschall M, Tsogoeva SB (2017) Synthesis of novel hybrids of quinazoline and artemisinin with high activities against *Plasmodium falciparum*, human cytomegalovirus, and leukemia cells. *ACS Omega* 2:2422–2431
- Gellis A, Primas N, Hutter S, Lanzada G, Remusat V, Verhaeghe P, Vanelle P, Azas N (2016) Looking for new antiplasmodial quinazolines: DMAP-catalyzed synthesis of 4-benzyloxy- and 4-aryloxy-2-trichloromethylquinazolines and their in vitro evaluation toward *Plasmodium falciparum*. *Eur J Med Chem* 119:34–44
- Gil A, Pabon A, Galiano S, Burguete A, Silanes SP, Deharo E, Monge A, Aldana I (2014) Synthesis, biological evaluation and structure-activity relationships of new quinoxaline derivatives as anti-*Plasmodium falciparum* agents. *Molecules* 19:2166–2180
- Gomtsyan A (2012) Heterocycles in drugs and drug discovery. *Chem Heterocycl Comp* 48:7–10
- Gopalakrishnan AM, Kumar Nirbhay (2015) Antimalarial action of artesunate involves DNA damage mediated by reactive oxygen species. *Antimicrob Agents Chemother* 59:317–325
- Guillon J, Cohen A, Gueddouda NM, Das RN, Moreau S, Ronga L, Savrimoutou S, Basmaciyan L, Monnier A, Monget M, Rubio S, Garnerin T, Azas N, Mergny JL, Mullie C, Sonnet P (2007) Design, synthesis and antimalarial activity of novel bis {N-[{pyrrolo[1,2-a]quinoxalin-4-yl}benzyl]-3-aminopropyl} amine derivatives. *J Enzyme Inhib Med Chem* 32:547–563
- Gupta V, Kant V (2013) A review on biological activity of imidazole and thiazole moieties and their derivatives. *Sci Int* 1:253–260
- Hong WD, Leung SC, Ampor danai K, Davies J, Priestly SR, Nixon GL, Berry NG, Hasnain SS, Antonyuk SV, Ward SA, Biagini GA, Neil PO (2018) Potent antimalarial 2-pyrazolyl quinolone bc_1 (Q_1) inhibitors with improved drug-like properties. *ACS Med Chem Lett* 9:1205–1210
- Jensen AR, Adams Y, Hviid L (2020) Cerebral *Plasmodium falciparum* malaria: the role of PfEMP1 in its pathogenesis and immunity, and PfEMP1-based vaccines to prevent it. *Immunol Rev* 293:230–252. <https://doi.org/10.1111/imr.12807>
- Joshi MC, Wicht KJ, Taylor D, Hunter R, Smith PJ, Egan TJ (2013) In vitro antimalarial activity, β -haematin inhibition and structure activity relationships in a series of quinoline triazoles. *Eur J Med Chem* 69:338–347
- Kalaria PN, Karad SC, Raval DK (2018) A review on diverse heterocyclic compounds as the privileged scaffolds in antimalarial drug discovery. *Eur J Med Chem* 158:917–936
- Karrouchi K, Radi S, Ramli Y, Taoufik J, Mabkhot YN, Al-aizari FA, Ansar M (2018) Synthesis and pharmacological activities of pyrazole derivatives: a review. *Molecules* 23:134. <https://doi.org/10.3390/molecules23010134>
- Kharb R, Sharma PC, Yar MS (2011) Pharmacological significance of triazole scaffold. *J Enzym Inhib Med Chem* 26:1–21
- Kondaparla S, Manhas A, Dola VR, Srivastava K, Puri SK, Katti SB (2018) Design, synthesis and antiplasmodial activity of novel imidazole derivatives based on 7-chloro-4-aminoquinoline. *Bioorg Chem* 80:204–211
- Kuile FOT, Nosten F, Chongsuphajaisiddhi T, White NJ, Dolan G, Luxemburger C, Phaipun L, Edstein MD, Webster HK (1993) Halofantrine versus mefloquine in treatment of multidrug-resistant falciparum malaria. *Lancet* 341:1044–1049
- Kumar A, Srivastava K, Kumar SR, Siddiqi MI, Puri SK, Sexana JK, Chauhan PMS (2011) 4-Anilinoquinoline triazines: a novel class of hybrid antimalarial agents. *Eur J Med Chem* 46:676–690
- Kumar D, Khan SI, Ponnann P, Rawat DS (2014a) Triazine-pyrimidine based molecular hybrids: synthesis, docking studies and evaluation of antimalarial activity. *N J Chem* 38:5087–5095
- Kumar D, Khan SI, Ponnann P, Rawat DS (2014b) Synthesis, antimalarial activity, heme binding and docking studies of 4-aminoquinoline-pyrimidine based molecular hybrids. *RSC Adv* 4:63655–63669
- Kumar G, Tanwar O, Kumar J, Akhter M, Sharma S, Pillai CR, Alam MM, Zama MS (2018) Pyrazole-pyrazoline as promising novel antimalarial agents: a mechanistic study. *Eur J Med Chem* 149:139–147
- Kumar S, Bhardwaj TR, Prasad DN, Singh RK (2018) Drug targets for resistant malaria: historic to future perspectives. *Biomed Pharmacother* 104:8–27
- Lacerda MVG, Llanos-Cuentas A, Krudsood S, Lon C, Saunders DL, Mohammed R, Yilma D, Pereira DB, Espino FEJ, Mia RZ, Chuquiyauri R, Val F (2019) Single-dose tafenoquine to prevent relapse of Plasmodium vivax malaria. *N Engl J Med* 380:215–228
- Lee WC, Russell B, Renia L (2019) Sticking for a cause: the *falciparum* malaria parasites cytoadherence paradigm. *Front Immunol* 10:1444. <https://doi.org/10.3389/fimmu.2019.01444>
- Manach CL, Cabrera DG, Douelle F, Nchinda AT, Younis Y, Taylor D, Wiesner L, White KL, Ryan E, March C, Duffy S, Avery VM, Waterson D, Witty MJ, Wittlin S, Charman SA, Street LJ, Chibale K (2014) Medicinal chemistry optimization of antiplasmodial imidazopyridazine hits from high throughput screening of a softfocus kinase library: Part 1. *J Med Chem* 57:2789–2798
- Manach CL, Nchinda AT, Paquet T, Cabrera DG, Younis Y, Han Z, Bashyam S, Zabiulla M, Taylor D, Lawrence N, White KL, Charman SA, Waterson D, Witty MJ, Wittlin S, Botha ME, Nondaba SH, Reader J, Birkholtz LM, Diaz MBJ, Martinez MS, Ferrer S, Barturen IA, Meister S, Koch YA, Winzeler EA, Street LJ, Chibale K (2016) Identification of a potential antimalarial drug candidate from a series of 2-aminopyrazines by optimization of aqueous solubility and potency across the parasite life cycle. *J Med Chem* 59:9890–9905
- Makhova NN, Belen'kii LI, Gazieva GA, Dalinger IL, Konstantinova LS, Kuznetsov VV, Kravchenko AN, Krayushkin MM, Rakitin OA, Starosotnikov AM, Fershtat LL, Shevelev SA, Shirinian VZ, Yarovenko VN (2020) Progress in the chemistry of nitrogen-, oxygen- and sulfur-containing heterocyclic systems. *Russ Chem Rev* 89:55–124
- Manlove A, Groziak MP (2009) Chapter 6.2: six-membered ring systems: diazines and benzo derivatives. *Prog Heterocycl Chem* 21:375–414

- Manohar S, Khan SI, Rawat DS (2010a) Synthesis, antimalarial activity and cytotoxicity of 4-aminoquinoline-triazine conjugates. *Bioorg Med Chem Lett* 20:322–325
- Manohar S, Khan SI, Rawat DS (2011b) Synthesis of 4-aminoquinoline-1,2,3-triazole and 4-aminoquinoline-1,2,3-triazole-1,3,5-triazine hybrids as potential antimalarial agents: synthesis of 4-aminoquinoline-1,2,3-triazole and 4-aminoquinoline-1,2,3-triazole-1,3,5-triazine hybrids. *Chem Biol Drug Des* 78:124–136
- Manohar S, Rajesh UC, Khan SI, Tekwani BL, Rawat DS (2012) Novel 4-aminoquinoline-pyrimidine based hybrids with improved and in vivo antimalarial activity. *ACS Med Chem Lett* 3:555–559
- Marella A, Shaquizzaman M, Akhter M, Verma G, Alam MM (2015) Novel pyrazole–pyrazoline hybrids endowed with thioamide as antimalarial agents: their synthesis and 3D-QSAR studies. *J Enzym Inhib Med Chem* 30:597–606
- Meshnick SR (2002) Artemisinin: mechanisms of action, resistance and toxicity. *Int J Parasitol* 32:1655–1660
- Mulla AA (2017) A review: biological importance of heterocyclic compounds. *Der Pharma Chem* 9:141–147
- Narula AK, Azad CS, Nainwal LM (2019) New dimensions in the field of antimalarial research against malaria resurgence. *Eur J Med Chem* 181:111353. <https://doi.org/10.1016/j.ejmech.2019.05.043>
- Noronha M, Pawar V, Prajapati A, Subramanian RB (2020) A literature review on traditional herbal medicines for malaria. *S Afr J Bot* 128:292–303. <https://doi.org/10.1016/j.sajb.2019.11.017>
- Okombo J, Brunschwig C, Singh K, Dzivornu GA, Barnard L, Njoroge M, Wittlin S, Chibale K (2019) Antimalarial pyrid[1,2-a] benzimidazole derivatives with Mannich base side chains: synthesis, pharmacological evaluation and reactive metabolite trapping studies. *ACS Infect Dis* 5:372–384
- Pandya KM, Patel AH, Desai PS (2019) Development of antimicrobial, antimalarial and antitubercular compounds based on a quinoline-pyrazole clubbed scaffold derived via Doebe reaction. *Chem Afr* 3:89–98
- Parhizgar AR, Tahghighi (2017) Introducing new antimalarial analogues of chloroquine and amodiaquine: a narrative review. *Iran J Med Sci* 42:115–128
- Parikh PH, Timaniya JB, Patel MJ, Patel KP (2020) Design, synthesis and characterization of novel substituted 1,2,4-oxadiazole and their biological broadcast. *Med Chem Res* 29:538–548. <https://doi.org/10.1007/s00044-020-02505-8>
- Patel TS, Bhatt JD, Dixit RB, Chudasama CJ, Patel BD, Dixit BC (2019) Green synthesis, biological evaluation, molecular docking studies and 3D-QSAR analysis of novel phenylalanine linked quinazoline-4(3H)-one-sulphonamide hybrid entities distorting the malarial reductase activity in folate pathway. *Bioorg Med Chem* 27:3574–3586
- Patel TS, Vanparia SF, Gandhi SA, Patel UH, Dixit RB, Chudasama CJ, Dixit BC (2015a) Novel stereoselective 2,3-disubstituted quinazoline-4(3H)-one derivatives derived from glycine as a potent antimalarial lead. *N. J Chem* 39:8638–8649
- Patel TS, Vanparia SF, Patel UH, Dixit RB, Chudasama CJ, Patel BD, Dixit BC (2017b) Novel 2,3-disubstituted quinazoline-4(3H)-one molecules derived from amino acid linked sulphonamide as a potent malarial antifolates for DHFR inhibition. *Eur J Med Chem* 129:251–265
- Pathania S, Narang RK, Rawal RK (2019) Role of sulphur heterocycles in medicinal chemistry: an update. *Eur J Med Chem* 180:486–508
- Pinheiro LCS, Feitosa LM, Silveira FFD, Boechat N (2018) Current antimalarial therapies and advances in the development of semi-synthetic artemisinin derivatives. *Acad Bras Cienc* 90:1251–1271
- Pippione AC, Sainas S, Goyal P, Fritzson I, Cassiano GC, Giraudo A, Giorgis M, Tavella TA, Bagnati R, Rolando B, Carlsson CC, Costa FTM, Andrade CH, Karadaghi SA, Boschi D, Friemann R, Lolli ML (2019) Hydroxyazole scaffold-based *Plasmodium falciparum* dihydroorotate dehydrogenase inhibitors: synthesis, biological evaluation and X-ray structural studies. *Eur J Med Chem* 163:266–280
- Ram VJ, Sethi A, Nath M, Pratap R (2019) The chemistry of heterocycles: chemistry of six-to-eight membered N, O, S, P and Se heterocycles. Elsevier Science Publishing Co Inc, United States
- Ramachandran S, Hameed PS, Srivastava A, Shanbhag G, Morayya S, Rautela N, Awasthy D, Kavanagh S, Bharath S, Reddy J, Panduga V, Prabhakar KR, Saralaya R, Nanduri R, Raichurkar A, Menasinakai, Achar V, Diaz MBJ, Martinez MS, Barturen IA, Ferrer S, Sanz LM, Gamo FJ, Duffy S, Avery VM, Waterson D, Lee MC, Flynn OC, Fidock DA, Iyer PS, Narayanan S, Hosagrahara V, Sambandamurthy VK (2014) *N*-aryl-2-aminobenzimidazoles: Novel, efficacious, antimalarial lead compounds. *J Med Chem* 57:6642–6652
- Ridley RG (2002) Medical need, scientific opportunity and the drive for antimalarial drugs. *Nature* 415:686–693
- Rungsirunrat K, Na-Bangchang K, Hawkins VN, Mungthin M, Sibley CH (2007) Sensitivity to antifolates and genetic analysis of *Plasmodium vivax* isolates from Thailand. *Am J Trop Med Hyg* 76:1057–1065
- Ryley JF (1953) The mode of action of proguanil and related antimalarial drugs. *Br J Pharm Chemother* 8:424–430
- Shah DR, Modh RP, Chikhalia KH (2014) Privileged *s*-triazines: structure and pharmacological applications. *Future Med Chem* 6:463–477
- Sharma M, Prasher P (2020) An epigrammatic status of the ‘azole’-based antimalarial drugs. *RSC Med Chem* 11:184–211. <https://doi.org/10.1039/C9MD00479C>
- Shekhar AC, Rao PS, Narsaiah B, Allanki AD, Sijwali PS (2014) Emergence of pyrido quinoxalines as new family of antimalarial agents. *Eur J Med Chem* 77:280–287
- Singh K, Kaur H, Smith P, Kock CD, Chibale K, Balzarini J (2014) Quinoline–pyrimidine hybrids: synthesis, antiplasmoidal activity, structure activity relationship and mode of action studies. *J Med Chem* 57:435–448
- Singh V, Kumar A (2015) The integral *Plasmodium* life cycle phenomenon: gamete genes. *J Bacteriol Parasitol* 06:224. <https://doi.org/10.4172/2155-9597.100024>
- Slater AFG (1993) Chloroquine: mechanism of drug action and resistance in *Plasmodium falciparum*. *Pharm Ther* 57:203–235
- Tariq S, Somakala K, Amir M (2018) Quinoxaline: an insight into the recent pharmacological advances. *Eur J Med Chem* 143:542–557
- Thakkar SS, Thakor P, Doshi H, Ray A (2017) 1,2,4-Triazole and 1,3,4-oxadiazole analogues: synthesis, MO studies, in silico molecular docking studies, antimalarial as DHFR inhibitor and antimicrobial activities. *Bioorg Med Chem* 25:4064–4075
- Tibon NS, Ng CH, Cheong SL (2020) Current progress in antimalarial pharmacotherapy and multi-target drug discovery. *Eur J Med Chem* 188:111983. <https://doi.org/10.1016/j.ejmech.2019.111983>
- Tripathi M, Khan SI, Ponnaiyan P, Kholiya R, Rawat DS (2017) Aminoquinoline-pyrimidine-modified anilines: synthesis, antiplasmoidal activity, cytotoxicity, mechanistic studies and ADME predictions. *Chem Sel* 2:9074–9083
- Tse EG, Korsik M, Todd MH (2019) The past, present and future of antimalarial medicines. *Malar J* 18:93. <https://doi.org/10.1186/s12936-019-2724-z>
- Tukulula M, Sharma RK, Meurillon M, Mahajan A, Naran K, Warner D, Huang J, Mekonnen B, Chibale K (2013) Synthesis and antiplasmoidal and antimycobacterial evaluation of new nitroimidazole and nitroimidazooxazine derivatives. *ACS Med Chem Lett* 4:128–131

- Vaidya AB, Mather MW (2000) Atovaquone resistance in malaria parasites. *Drug Resist Updat* 3:283–287
- de Pilla Varotti F, Botelho ACC, Andrade AA, de Paula RC, Fagundes EMS, Valverde A, Mayer LMU, Mendonca JS, de Souza MVN, Boechat N, Krettli AU (2008) Synthesis, antimarial activity, and intracellular targets of MEFAS, a new hybrid compound derived from mefloquine and artesunate. *Antimicrob Agents Chemother* 52:3868–3874
- Verma G, Chashoo G, Ali A, Khan MF, Akhtar W, Ali I, Akhtar M, Alam MM, Shaquiquzzaman M (2018) Synthesis of pyrazole acrylic acid based oxadiazole and amide derivatives as anti-malarial and anticancer agents. *Bioorg Chem* 77:106–124
- Vinayak S, Alam MT, Mixon-Hayden T, McCollum AM, Sem R, Shah NK, Lim P, Muth S, Rogers WO, Fandeur T, Barnwell JW, Escalante AA, Wongsrichanalai C, Ariey F, Meshnick SR, Udhayakumar V (2010) Origin and evolution of sulfadoxine resistant *Plasmodium falciparum*. *PLoS Pathog* 6:e1000830. <https://doi.org/10.1371/journal.ppat.1000830>
- Younis Y, Douelle F, Cabrera DG, Manach CL, Nchinda AT, Paquet T, Street LJ, White KL, Zabiulla KM, Joseph JT, Bashyam S, Waterson, Witty MJ, Wittlin S, Charman SA, Chibale K (2013) Structure-activity-relationship studies around the 2-amino group and pyridine core of antimarial 3,5-diarylaminopyridines lead to a novel series of pyrazine analogues with oral in vivo activity. *J Med Chem* 56:8860–8871
- Yuvaniyama J, Chitnumsub P, Kamchonwongpaisan S, Vanichtanakul J, Sirawaraporn W, Taylor P, Walkinshaw MD, Yuthavong Y (2003) Insights into antifolate resistance from malarial DHFR-TS structures. *Nat Struct Mol Biol* 10:357–365
- Zhang YK, Plattner JJ, Easom EE, Jacobs RT, Guo D, Sanders V, Freund YR, Campo B, Rosenthal PJ, Bu W, Gamo FJ, Sanz LM, Ge M, Li L, Ding J, Yang Y (2015) Benzoxaborole antimarial agents. Part 4. Discovery of potent 6-(2-(alkoxycarbonyl)pyrazinyl-5-oxy)-1,3-dihydro-1-hydroxy-2,1-benzoxaboroles. *J Med Chem* 58:5344–5354



Tankyrase inhibitors: emerging and promising therapeutics for cancer treatment

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Abstract

Cancer is a disease portrayed by the uncontrolled growth of irregular cells. Tankyrase, a member of poly(ADP-ribose) polymerase family, mediates Wnt signal transduction and has emerged as a new molecular target for the therapy of different kinds of cancer. Wnt/β-catenin signaling functions significantly in a wide scope of biological events, such as the upkeep of genomic stability, transcriptional control, energy metabolism, and apoptosis. ADP-ribosylation is a reversible posttranslational modification process that regulates several cellular signaling pathways in which transferase enzymes such as mono (ADP-ribosyl) and poly(ADP-ribosyl) transferases move a unit of ADP-ribose moiety from the NAD⁺ co-substrate to specific amino acid side chains and/or potentially ADP-ribose units on target proteins. Recently, inhibition of tankyrase has risen as an appealing strategy for the discovery of novel anticancer drugs. The current review offers an understanding of the ongoing improvements on new lead structures as inhibitors of tankyrase and their activities. A special spotlight is set on the structure-activity relationship, molecular docking, polypharmacology profile, and binding mode at the active center.

Keywords Cancer · Tankyrase · ADP-ribosylation · Wnt/β-catenin signaling · Tankyrase inhibitors

Abbreviations

ARTD	ADP-ribose transferase
PARP	Poly(ADP-ribose)polymerase
TNKS	Tankyrase
PKA	Protein kinaseA
APC	Adenomatous polyposis coli gene
CK1	Casein kinase1
GSK3	Glycogen synthase kinase 3
TCF	T-cell factor
LEF	Lymphoid enhancer-binding factor
LRP6	Lipoprotein receptor-related protein
TRF	Telomere repeat-binding factor
MAPK	Mitogen-activated protein kinase
SAM	Sterile alpha motif
NuMA	Nuclear mitotic apparatus protein
IRAP	Insulin-responsive aminopeptidase

Introduction

The human body comprises a huge number of cells; these cells grow and divide to produce new cells as the body needs them. At that point, when cells grow old or damaged, they die and new cells replace them. The mechanism once in a while goes wrong and various cells grow uncontrolled, which prompts cancer [1]. Cancer is a deadly ailment brought about by uncontrolled cell growth and originates from the additional mass tissue known as tumor and spreads all through the circulatory system in the human body [2]. Leukemia affects the blood cells and includes them in their maturity and immaturity. Some of the tumors do not spread all through the body yet they grow wildly like benign tumors [3]. Normal cells regulate their development and kill themselves when they become unhealthy. Various organs can be influenced by cancer cells like the lungs, kidney, eyes, heart, brain, and so forth [4].

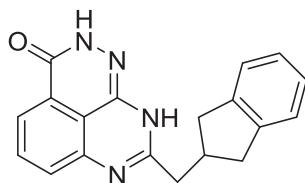
Numerous investigations have indicated that tankyrases assume a fundamental part in the development and progression of distinct types of carcinomas involving fibrosarcoma, pancreatic adenocarcinoma, ovarian cancer, glioblastoma, gastric cancer, breast cancer, and transitional cell carcinoma of the bladder. Tankyrase 1 (TNKS1) and

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a distinct spectrum of clinical opportunity. E7499 currently completed the first period of the clinical trial [88, 89].



(91) E7449

PARP1/2 IC₅₀ = 0.002 μM / 0.001 μM

TNKS1/2 IC₅₀ = 0.05 μM / 0.12 μM

Conclusion

In humans, alteration of TNKS1 and TANK2 and distorted Wnt pathway underlies a wide array of diseases including tumor development, tumor commencement, cell senescence, cell demise, proliferation, and metastasis. In recent decades, tankyrases and the Wnt pathway have proven to be an appealing target for the discovery of anticancer drugs. The tankyrase inhibitors which focus the amino acid of the adenosine binding site along with the residues of the nicotinamide binding site show an exceptionally particular nature for tankyrase over other ARTD isoenzymes. A large number of tankyrase inhibitors that have been developed indicated structural similarity with the significant coenzyme NAD⁺, which may cause toxicities due to off-target binding. High specificity is important for the novel tankyrase inhibitors to avoid the potential adverse effects. The literature review demonstrated the computational studies by using various potent compounds that help the researchers to observe a potential association between the coupling site and inhibitor molecule. It shows that the inhibitory activity of nicotinamide restricting site inhibitors can be expanded by designing new analogs that collaborate with hydrophobic amino acids Phe1188 residue of TNKS1 and Phe1035 of TNKS2. Moreover, it is also indicated that adenosine binding site restricted inhibitors form aryl interaction or π–π stacking interaction with polar amino-acids Histidine. This selective character of TNKS1 and TNKS2 inhibitors provides an excellent platform for dual-site binders. A recently established quinazoline derivative was discovered to be more potent when compared with other tankyrase inhibitors. The quinazoline moiety ties to the nicotinamide binding motif and forms hydrogen bonding with glycine and serine. The central part of the moiety constitutes a hydrogen bond with Tyrosine amino acid, while extended lateral chains form hydrogen bonding with aspartate and glycine amino residues of the protein. All these findings will give meaningful clues in the design and advancement of new specific

inhibitors that can be utilized to treat different kinds of cancer.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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References

- Vanita P, Subrahmanyam V, Jhansi K. A short note on cancer. *J Carcinog Mutagen.* 2011;2:128. <https://doi.org/10.4172/2157-2518.1000128>.
- Majumder P, Saha AK, Majumder M. Selection of significant lifestyle risk factor of cancer by hybrid X-Bar-DEMATEL-TOPSIS method. *J Pharm Sci Res.* 2017;9:878–85.
- Nair LS, Mahesh S, Smitha LS, Sujathan K, Remani P. Expression of *Canavalia gladiata* lectin in leukemic cells. *J Cancer Sci Ther.* 2011;3:88–91.
- Naga Deepthi CH, Pavan Kumar VVL, Rameshbabu A, Indrapriyadarshini U. Role of tumor suppressor protein p53 in apoptosis and cancer therapy. *J Cancer Sci Ther.* 2011;S17:001. <https://doi.org/10.4172/1948-5956.S17-001>.
- Cheng H, Li X, Wang C, Chen Y, Li S, Tan J, et al. Inhibition of tankyrase by a novel small molecule significantly attenuates prostate cancer cell proliferation. *Cancer Lett.* 2019;443:80–90. <https://doi.org/10.1016/j.canlet.2018.11.013>.
- Guo HL, Zhang C, Liu Q, Li Q, Lian G, Wu D, et al. The Axin/TNKS complex interacts with KIF3A and is required for insulin-stimulated GLUT4 translocation. *Cell Res.* 2012;22:1246–57.
- Li Z, Yamauchi Y, Kamakura M, Murayama T, Goshima F, Kimura H, Nishiyama Y. Herpes Simplex Virus Requires Poly (ADP-Ribose) Polymerase Activity for Efficient Replication and Induces Extracellular Signal-Related Kinase-Dependent Phosphorylation and ICP0-Dependent Nuclear Localization of Tankyrase 1. *J Virol.* 2012;86:492–503.
- Che L, Song JY, Lou Y, Li GY. Analysis from the perspective of cilia: the protective effect of PARP inhibitors on visual function during light-induced damage. *Int Ophthalmol.* 2019;40:1017–27.
- Haikarainen T, Krauss S, Lehtio L. Tankyrases: structure, function and therapeutic implications in cancer. *Curr Pharm Des.* 2014;20:6472–88.
- Riffell JL, Lord CJ, Ashworth A. Tankyrase-targeted therapeutics: expanding opportunities in the PARP family. *Nat Rev Drug Discov.* 2012;11:923–36.
- Hottiger MO, Hassa PO, Luscher B, Schuler H, Koch-Nolte F. Toward a unified nomenclature for mammalian ADP-ribosyltransferases. *Trends Biochem Sci.* 2010;35:208–19.
- Guettler S, LaRose J, Petsalaki E, Gish G, Scotter A, Pawson T, et al. Structural basis and sequence rules for substrate recognition by tankyrase explain the basis for cherubism disease. *Cell.* 2011;147:1340–54.

13. Ha GH, Kim HS, Go H, Lee H, Seimiya H, Chung DH, et al. Tankyrase-1 function at telomeres and during mitosis is regulated by Polo-like kinase-1-mediated phosphorylation. *Cell Death Differ.* 2012;19:321–32.
14. Zimmerlin L, Zambidis ET. Pleiotropic roles of tankyrase/PARP proteins in the establishment and maintenance of human naïve pluripotency. *Exp Cell Res.* 2020;390:111935.
15. Curtin NJ, Szabo C. Poly (ADP-ribose) polymerase inhibition: past. *Nat Rev Drug Discov.* 2020;19:711–36.
16. Morrone S, Cheng Z, Moon RT, Cong F, Xu W. Crystal structure of a tankyrase-axin complex and its implications for axin turnover and tankyrase substrate recruitment. *Proc Natl Acad Sci India Sect B Biol Sci.* 2012;109:1500–5.
17. Callow MG, Tran H, Phu L, Lau T, Lee J, Sandoval WN, et al. Ubiquitin ligase RNF146 regulates tankyrase and axin to promote Wnt signaling. *PLoS ONE.* 2011;6:e22595.
18. Jain PG, Patel BD. Medicinal chemistry approaches of poly ADP-ribose polymerase 1 (PARP1) inhibitors as anticancer agents—a recent update. *Eur J Med Chem.* 2019;165:198–215.
19. Banerjee J, Lodhi N, Nguyen BN. The role of Poly (ADP-Ribose) polymerase-1 in cutaneous wound healing. *Adv Wound Care.* 2019;8:634–43.
20. Shirai F, Tsumura T, Yashiroda Y, Yuki H, Niwa H, Sato S, et al. Discovery of novel spiroindoline derivatives as selective tankyrase inhibitors. *J Med Chem.* 2019;62:3407–27.
21. Ferri M, Liscio P, Carotti A, Asciutti S, Sardella R, Macchiarulo A, et al. Targeting Wnt-driven cancers: discovery of novel tankyrase inhibitors. *Eur J Med Chem.* 2017;142:506–22.
22. Guettler S. AXIN shapes tankyrase architecture. *Structure.* 2016;24:1625–7.
23. Pollock K, Liu M, Zaleska M, Meniconi M, Pfuhl M, Collins I, et al. Fragment-based screening identifies molecules targeting the substrate-binding ankyrin repeat domains of tankyrase. *Sci Rep.* 2019;9:1–20.
24. Mariotti L, Templeton CM, Ranes M, Paracuellos P, Cronin N, Beuron F, et al. Tankyrase requires SAM domain-dependent polymerization to support Wnt-β-catenin signaling. *Mol Cell.* 2016;63:498–513.
25. Sidaway JE, Orton TC, Kalaitzi K, Jones HB, Foster A, Lake BG. Analysis of β-catenin gene mutations and gene expression in liver tumours of C57BL/10J mice produced by chronic administration of sodium phenobarbital. *Toxicology.* 2019;430:152343. <https://doi.org/10.1016/j.tox.2019.152343>.
26. Voronkov A, Krauss S. Wnt/β-catenin signaling and small molecule inhibitors. *Curr Pharm Des.* 2013;19:634–64.
27. Stakheev D, Taborska P, Strizova Z, Podrazil M, Bartunkova J, Smrz D. The WNT/β-catenin signaling inhibitor XAV939 enhances the elimination of LNCaP and PC-3 prostate cancer cells by prostate cancer patient lymphocytes in vitro. *Sci Rep.* 2019;9:1–14.
28. Kim MK. Novel insight into the function of tankyrase. *Oncol Lett.* 2018;16:6895–902.
29. Nguyen VHL, Hough R, Bernardo S, Peng C. Wnt/β-catenin signalling in ovarian cancer: insights into its hyperactivation and function in tumorigenesis. *J Ovarian Res.* 2019;12:122. <https://doi.org/10.1186/s13048-019-0596-z>.
30. Huang SMA, Mishina YM, Liu S, Cheung A, Stegmeier F, Michaud GA, et al. Tankyrase inhibition stabilizes axin and antagonizes Wnt signalling. *Nature.* 2009;461:614–20.
31. Taciak B, Pruszynska I, Kiraga L, Bialasek M, Krol M. Wnt signaling pathway in development and cancer. *J Physiol Pharm.* 2018;69:185–96.
32. Jarman EJ, Boulter L. Targeting the Wnt signaling pathway: the challenge of reducing scarring without affecting repair. *Expert Opin Investig Drugs.* 2020;29:179–90. <https://doi.org/10.1080/13543784.2020.1718105>.
33. Velho PI, Fu W, Wang H, Mirkheshti N, Qazi F, Lima FAS, et al. Wnt-pathway activating mutations are associated with resistance to first-line abiraterone and enzalutamide in castration-resistant prostate cancer. *Eur Urol.* 2019;177:14–21.
34. Harvey KF, Zhang X, Thomas DM. The Hippo pathway and human cancer. *Nat Rev Cancer.* 2013;13:246–57.
35. Zhao B, Li L, Lei Q, Guan KL. The Hippo-YAP pathway in organ size control and tumorigenesis: an updated version. *Genes Dev.* 2010;24:862–74.
36. Halder G, Johnson RL. Hippo signaling: growth control and beyond. *Development.* 2011;138:9–22.
37. Dong K, Xue H, Cheng J, Su J, Li D, Zhang J, et al. PRPH2 activates Hippo signalling and suppresses the invasion and anoikis inhibition of laryngeal cancer. *Cancer Manag Res.* 2019;11:10107–15.
38. Mo JS, Park HW, Guan KL. The Hippo signaling pathway in stem cell biology and cancer. *EMBO Rep.* 2014;15:642–56.
39. Flinn MA, Link BA, O'Meara CC. Upstream regulation of the Hippo-Yap pathway in cardiomyocyte regeneration. *Semin Cell Dev Biol.* 2020;100:11–9.
40. Sharma J, Madan P. Characterisation of the Hippo signalling pathway during bovine preimplantation embryo development. *Reprod Fertil Dev.* 2019;32:392–401.
41. Smith S, de Lange T. Tankyrase promotes telomere elongation in human cells. *Curr Biol.* 2000;10:1299–302.
42. Storti CB, De Oliveira RA, De Carvalho M, Hasimoto EN, Cataneo DC, Cataneo AJM, et al. Telomere-associated genes and telomeric lncRNAs are biomarker candidates in lung squamous cell carcinoma (LUSC). *Exp Mol Pathol Suppl.* 2020;112:104354.
43. Smith S, Giriat I, Schmitt A, De Lange T. Tankyrase, a poly (ADP-ribose) polymerase at human telomeres. *Science.* 1998;282:1484–7.
44. Zhao S, Wang F, Liu L. Alternative lengthening of telomeres (ALT) in tumors and pluripotent stem cells. *Genes.* 2019;10:1030. <https://doi.org/10.3390/genes10121030>.
45. Boltz KA, Jasti M, Townley JM, Shippen DE. Analysis of poly (ADP-ribose)polymerases in arabidopsis telomere biology. *PLoS ONE.* 2014;9:e88872.
46. Patel B, Patel A, Bhattacharya H, CoMSIA, molecular docking and MOLCAD studies of pyrimidinone derivatives to design novel and selective tankyrase inhibitors. *J Mol Struct.* 2020;1221:128783.
47. Yang L, Sun L, Teng Y, Chen H, Gao Y, Levine AS, et al. Tankyrase1-mediated poly (ADP-ribosylation) of TRF1 maintains cell survival after telomeric DNA damage. *Nucleic Acids Res.* 2017;45:3906–21.
48. Franchet C, Hoffmann JS. When RAD52 allows mitosis to accept unscheduled DNA synthesis. *Cancers.* 2019;12:26. <https://doi.org/10.3390/cancers12010026>.
49. Daniloski Z, Bisht KK, McStay B, Smith S. Resolution of human ribosomal DNA occurs in anaphase, dependent on tankyrase 1, condensin II, and topoisomerase IIα. *Genes Dev.* 2019;33:276–81.
50. Kim MK, Smith S. Persistent telomere cohesion triggers a prolonged anaphase. *Mol Biol Cell.* 2014;25:30–40.
51. Lehtio L, Chi NW, Krauss S. Tankyrases as drug targets. *FEBS J.* 2013;280:3576–93.
52. Su Z, Deshpande V, James DE, Stockli J. Tankyrase modulates insulin sensitivity in skeletal muscle cells by regulating the stability of GLUT4 vesicle proteins. *J Biol Chem.* 2018;293:8578–87.
53. Di Micco S, Pulvirenti L, Bruno I, Terracciano S, Russo A, Vaccaro MC, et al. Identification by inverse virtual screening of magnolol-based scaffold as new tankyrase-2 inhibitors. *Bioorg Med Chem.* 2018;26:3953–7.
54. Kirubakaran P, Arunkumar P, Premkumar K, Muthusamy K. Sighting of tankyrase inhibitors by structure-and ligand-based screening and in vitro approach. *Mol Biosyst.* 2014;10:2699–712.

55. Wu H, Wu J, Zhang W, Li Z, Fang J, Lian X, et al. Discovery and structure-activity relationship study of phthalimide-phenylpyridine conjugate as inhibitor of Wnt pathway. *Bioorg Med Chem Lett.* 2019;29:870–2.
56. Shultz MD, Cheung AK, Kirby CA, Firestone B, Fan J, Chen CHT, et al. Identification of NVP-TNKS656: the use of structure–efficiency relationships to generate a highly potent, selective, and orally active tankyrase inhibitor. *J Med Chem.* 2013;56:6495–511.
57. Liscio P, Carotti A, Asciutti S, Ferri M, Pires MM, Valloscuro S, et al. Scaffold hopping approach on the route to selective tankyrase inhibitors. *Eur J Med Chem.* 2014;87:611–23.
58. Del Bello F, Farande A, Giannella M, Piergentili A, Quaglia W, Benicchi T, et al. Identification of 2-aminopyrimidine derivatives as inhibitors of the canonical Wnt signaling pathway. *Bioorg Med Chem.* 2015;23:5725–33.
59. Johannes JW, Almeida L, Barlaam B, Boriack-Sjodin PA, Casella R, Croft RA, et al. Pyrimidinone nicotinamide mimetics as selective tankyrase and Wnt pathway inhibitors suitable for in vivo pharmacology. *ACS Med Chem Lett.* 2015;6:254–9.
60. Haikarainen T, Waaler J, Ignatev A, Nkizinkiko Y, Venkannagari H, Obaji E, et al. Development and structural analysis of adenosine site binding tankyrase inhibitors. *Bioorg Med Chem Lett.* 2016;26:328–33.
61. Shultz MD, Majumdar D, Chin DN, Fortin PD, Feng Y, Gould T, et al. Structure–efficiency relationship of [1, 2, 4] Triazol-3-ylamines as novel nicotinamide isosteres that inhibit tankyrases. *J Med Chem.* 2013;56:7049–59.
62. Liscio P, Carotti A, Asciutti S, Karlberg T, Bellocchi D, Llacuna L, et al. Design, synthesis, crystallographic studies, and preliminary biological appraisal of new substituted triazolo [4, 3-b] pyridazin-8-amine derivatives as tankyrase inhibitors. *J Med Chem.* 2014;57:2807–12.
63. Anumala UR, Waaler J, Nkizinkiko Y, Ignatev A, Lazarow K, Lindemann P, et al. Discovery of a novel series of tankyrase inhibitors by a hybridization approach. *J Med Chem.* 2017;60:10013–25.
64. Waaler J, Leenders RGG, Sowa ST, Alam Brinch S, Lycke M, Nieczypor P, et al. Preclinical lead optimization of a 1,2,4-triazole based tankyrase inhibitor. *J Med Chem.* 2020;63:6834–46. <https://doi.org/10.1021/acs.jmedchem.0c00208>.
65. Narwal M, Haikarainen T, Fallarero A, Vuorela PM, Lehtio L. Screening and structural analysis of flavones inhibiting tankyrases. *J Med Chem.* 2013;56:3507–17.
66. Narwal M, Koivunen J, Haikarainen T, Obaji E, Legala OE, Venkannagari H, et al. Discovery of tankyrase inhibiting flavones with increased potency and isoenzyme selectivity. *J Med Chem.* 2013;56:7880–9.
67. James RG, Davidson KC, Bosch KA, Biechele TL, Robin NC, Taylor RJ, et al. WIKI4, a novel inhibitor of tankyrase and Wnt/β-catenin signaling. *PLoS ONE.* 2012;7:e50457. <https://doi.org/10.1371/journal.pone.0050457>.
68. Larsson EA, Jansson A, Ng FM, Then SW, Panicker R, Liu B, et al. Fragment-based ligand design of novel potent inhibitors of tankyrases. *J Med Chem.* 2013;56:4497–508.
69. Paine HA, Nathubhai A, Woon EC, Sunderland PT, Wood PJ, Mahon MF, et al. Exploration of the nicotinamide-binding site of the tankyrases, identifying 3-aryloquinolin-1-ones as potent and selective inhibitors in vitro. *Bioorg Med Chem.* 2015;23: 5891–908.
70. Haikarainen T, Koivunen J, Narwal M, Venkannagari H, Obaji E, Joensuu P, et al. Para-substituted 2-phenyl-3, 4-dihydroquinazolin-4-ones as potent and selective tankyrase inhibitors. *ChemMedChem.* 2013;8:1978–85.
71. Nathubhai A, Wood PJ, Lloyd MD, Thompson AS, Threadgill MD. Design and discovery of 2-arylquinazolin-4-ones as potent and selective inhibitors of tankyrases. *ACS Med Chem Lett.* 2013;4:1173–7.
72. Nathubhai A, Haikarainen T, Hayward PC, Munoz-Descalzo S, Thompson AS, Lloyd MD, et al. Structure-activity relationships of 2-arylquinazolin-4-ones as highly selective and potent inhibitors of the tankyrases. *Eur J Med Chem.* 2016;118:316–27.
73. Bregman H, Chakka N, Guzman-Perez A, Gunaydin H, Gu Y, Huang X, et al. Discovery of novel, induced-pocket binding oxazolidinones as potent, selective, and orally bioavailable tankyrase inhibitors. *J Med Chem.* 2013;56:4320–42.
74. Hua Z, Bregman H, Buchanan JL, Chakka N, Guzman-Perez A, Gunaydin H, et al. Development of novel dual binders as potent, selective, and orally bioavailable tankyrase inhibitors. *J Med Chem.* 2013;56:10003–15.
75. Nkizinkiko Y, Kumar BS, Jeankumar VU, Haikarainen T, Koivunen J, Madhuri C, et al. Discovery of potent and selective nonplanar tankyrase inhibiting nicotinamide mimics. *Bioorg Med Chem.* 2015;23:4139–49.
76. Okada-Iwasaki R, Takahashi Y, Watanabe Y, Ishida H, Saito JI, Nakai R, et al. The discovery and characterization of K-756, a novel Wnt/β-catenin pathway inhibitor targeting tankyrase. *Mol Cancer Ther.* 2016;15:1525–34.
77. Nathubhai A, Haikarainen T, Koivunen J, Murthy S, Koumanov F, Lloyd MD, et al. Highly potent and isoform selective dual site binding tankyrase/Wnt signaling inhibitors that increase cellular glucose uptake and have antiproliferative activity. *J Med Chem.* 2017;60:814–20.
78. Buchstaller HP, Anlauf U, Dorsch D, Kuhn D, Lehmann M, Leuthner B, et al. Discovery and optimization of 2-Arylquinazolin-4-ones into a potent and selective tankyrase inhibitor modulating Wnt pathway activity. *J Med Chem.* 2019;62:7897–909.
79. Voronkov A, Holsworth DD, Waaler J, Wilson SR, Ekblad B, Perdreau-Dahl H, et al. Structural basis and SAR for G007-LK, a lead stage 1, 2, 4-triazole based specific tankyrase 1/2 inhibitor. *J Med Chem.* 2013;56:3012–23.
80. Huang H, Guzman-Perez A, Acquaviva L, Berry V, Bregman H, Dovey J, et al. Structure-based design of 2-aminopyridine oxazolidinones as potent and selective tankyrase inhibitors. *ACS Med Chem Lett.* 2013;4:1218–23.
81. Bregman H, Gunaydin H, Gu Y, Schneider S, Wilson C, DiMauro EF, Huang X. Discovery of a Class of Novel Tankyrase Inhibitors that Bind to Both the Nicotinamide Pocket and the Induced Pocket. *J Med Chem.* 2013;56:1341–5.
82. Thomson DW, Wagner AJ, Bantscheff M, Benson RE, Dittus L, Duempelfeld B, et al. Discovery of a highly selective tankyrase inhibitor displaying growth inhibition effects against a diverse range of tumor derived cell lines. *J Med Chem.* 2017;60:5455–71.
83. De Vicente J, Tivitmahaisoon P, Berry P, Bolin DR, Carvajal D, He W, et al. Fragment-based drug design of novel pyranopyridones as cell active and orally bioavailable tankyrase inhibitors. *ACS Med Chem Lett.* 2015;6:1019–24.
84. Shirai F, Mizutani A, Yashiroda Y, Tsumura T, Kano Y, Muramatsu Y, et al. Design and discovery of an orally efficacious spiroindolinone-based tankyrase inhibitor for the treatment of colon cancer. *J Med Chem.* 2020;63:4183–204.
85. Tomassi S, Pfahler J, Mautone N, Rovere A, Esposito C, Passeri D, et al. From PARP1 to TNKS2 inhibition: a structure-based approach. *ACS Med Chem Lett.* 2020;11:862–8.
86. Li B, Liang J, Lu F, Zeng G, Zhang J, Ma Y, et al. Discovery of novel inhibitor for WNT/β-catenin pathway by tankyrase 1/2 structure-based virtual screening. *Molecules.* 2020;25:1680.
87. Berishvili VP, Kuimov AN, Voronkov AE, Radchenko EV, Kumar P, Choonara YE, et al. Discovery of novel tankyrase inhibitors through molecular docking-based virtual screening and molecular dynamics simulation studies. *Molecules.* 2020;25:1–15.

88. McGonigle S, Chen Z, Wu J, Chang P, Kolber-Simonds D, Ackermann K, et al. E7449: a dual inhibitor of PARP1/2 and tankyrase1/2 inhibits growth of DNA repair deficient tumors and antagonizes Wnt signaling. *Oncotarget*. 2015;6:41307–23.
89. Plummer R, Dua D, Cresti N, Drew Y, Stephens P, Foegh M, et al. First-in-human study of the PARP/tankyrase inhibitor E7449 in patients with advanced solid tumours and evaluation of a novel drug-response predictor. *Br J Cancer*. 2020;111:1–9.



Construction of pioneering quantitative structure activity relationship screening models for abuse potential of designer drugs using index of ideality of correlation in monte carlo optimization

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Abstract

Drug abuse is a worldwide wide problem affecting individual, society and the environment in general and it is nothing less than the attempted ecocide. Designer drugs are the chemical substances used for recreational purposes and have addictive properties. The production of designer drugs at disturbing pace is creating difficulties for the investigators in their testing. Computational evaluation method can be an interesting approach for early checking of abusive drugs. In the present work, quantitative structure activity relationship (QSAR) models are developed for abusive potential of designer drugs using SMILES and graph based parameters. Dopamine transporter/serotonin transporter inhibition (DAT/SERT) ratio was used as endpoint and the whole data set was divided into eight non identical splits for development of the models using balance of correlation technique of Monte Carlo optimization. The internal and external cross validation results confirmed that the models created with index of ideality of correlation were reliable and robust in prediction. The developed models followed all the five principles of the Organisation for Economic Co-operation and Development. The best model split 2 possessed good fitting ability and internal as well as external predictive ability and it was used in explanation of activity trends of different classes of designer drugs.

Keywords Drug abuse · Designer drugs · DAT/SERT ratio · Quantitative structure activity relationship · Index of ideality of correlation

Introduction

Drug addiction is a major issue around the world that imposes immense social and economic pressures on people and on society at large (Han and Gu 2006). According to the World drug report 2019, approximately 35 million people are affected with illicit drug use disorders worldwide. Internationally, around 11 million people took injectable illicit

drugs in 2017, of whom 1.4 million are infected with HIV and 5.6 million with hepatitis C (Hansford 2019). There is burgeoning worldwide apprehension about the synthetic analogs of controlled drugs being produced and marketed to curtail drug laws and escape prohibition. “Designer drugs” or “legal highs” are the psychotropic drugs which are purposely commercialized and delivered for recreational use by perpetuating shortcomings in current controlled substance legislation (Weaver et al. 2015). Many chemicals alluded as designer pharmaceutical products may be licensed medically in various countries, thus not meeting the standard concept of a designer drug (Luethi and Liechti 2020).

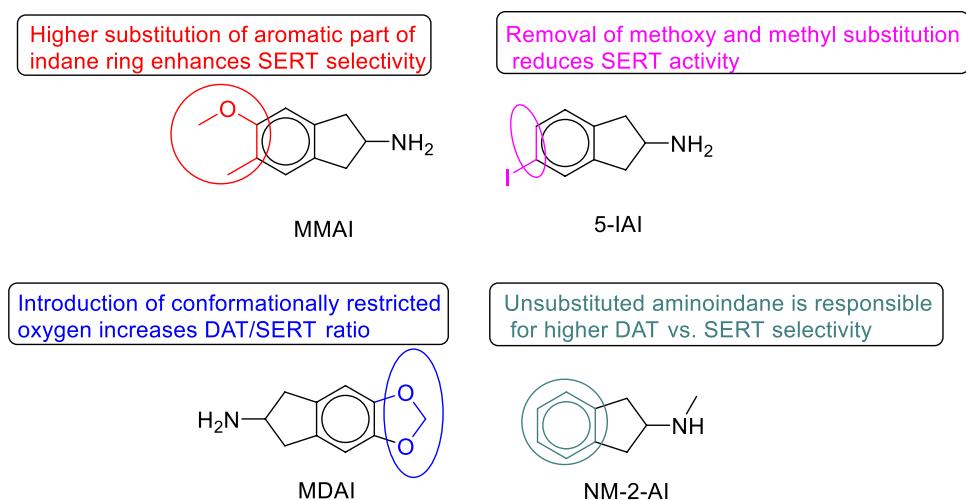
The abuse of designer drugs among youngsters is prevalent, particularly in the “rave and dance club scene” (Richter et al. 2019). Usage of “classic” designer drugs like ecstasy was record high in the 1990s but is still consumed today and that caused the development of most of the “classic” designer drugs in various countries (Langman and Snozek 2019). These drugs generate euphoria and feeling of high energy, and a tendency for socialization. Designer drugs

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Fig. 8 Description of SAR among Aminoindanes

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References

- Amara SG, Arriza JL (1993) Neurotransmitter transporters: three distinct gene families. *Curr Opin Neurobiol* 3(3):337–344. [https://doi.org/10.1016/0959-4388\(93\)90126-j](https://doi.org/10.1016/0959-4388(93)90126-j)
- Carnesecchi E, Toropov AA, Toropova AP, Kramer N, Svendsen C, Dorne JL, Benfenati E (2020) Predicting acute contact toxicity of organic binary mixtures in honey bees (*A. mellifera*) through innovative QSAR models. *Sci Total Environ* 704:135302. <https://doi.org/10.1016/j.scitotenv.2019.135302>
- Cherkasov A, Muratov EN, Fourches D, Varnek A, Baskin II, Cronin M, Dearden J, Gramatica P, Martin YC, Todeschini R, Consonni V, Kuz'min VE, Cramer R, Benigni R, Yang C, Rathman J, Terfloth L, Gasteiger J, Richard A, Tropsha A (2014) QSAR modeling: where have you been? Where are you going to? *J Med Chem* 57(12):4977–5010. <https://doi.org/10.1021/jm4004285>
- Duhan M, Singh R, Devi M, Sindhu J, Bhatia R, Kumar A, Kumar P (2019) Synthesis, molecular docking and QSAR study of thiazole clubbed pyrazole hybrid as α -amylase inhibitor. *J Biomol Struct Dyn*. <https://doi.org/10.1080/07391102.2019.1704885>
- Glennon RA, Dukat M (2017) Structure-activity relationships of synthetic cathinones. *Curr Top Behav Neurosci* 32:19–47. https://doi.org/10.1007/7854_2016_41
- Golbraikh A, Tropsha A (2002) Beware of q2! *J Mol Graph Model* 20(4):269–276. [https://doi.org/10.1016/s1093-3263\(01\)00123-1](https://doi.org/10.1016/s1093-3263(01)00123-1)
- Gramatica P (2007) Principles of QSAR models validation: internal and external. *QSAR Comb Sci* 26(5):694–701. <https://doi.org/10.1002/qsar.200610151>
- Gramatica P (2013) On the development and validation of QSAR models. *Methods Mol Biol* (Clifton N.J.) 930:499–526. https://doi.org/10.1007/978-1-62703-059-5_21
- Halpern P, Moskovich J, Avrahami B, Bentur Y, Soffer D, Peleg K (2011) Morbidity associated with MDMA (ecstasy) abuse: a survey of emergency department admissions. *Hum Exp Toxicol* 30(4):259–266. <https://doi.org/10.1177/0960327110370984>
- Han DD, Gu HH (2006) Comparison of the monoamine transporters from human and mouse in their sensitivities to psychostimulant drugs. *BMC Pharmacol* 6:6. <https://doi.org/10.1186/1471-2210-6-6>
- Hansford B (2019) World Drug Report 2019: 35 million people worldwide suffer from drug use disorders while only 1 in 7 people receive treatment. https://wdr.unodc.org/wdr2019/press/WDR_2019_press_release.pdf. Accessed on 10 May 2020
- Iversen LL (1971) Role of transmitter uptake mechanisms in synaptic neurotransmission. *Br J Pharmacol* 41(4):571–591. <https://doi.org/10.1111/j.1476-5381.1971.tb07066.x>
- Kanner BI, Schuldiner S (1987) Mechanism of transport and storage of neurotransmitters. *CRC Crit Rev Biochem* 22(1):1–38. <https://doi.org/10.3109/10409238709082546>
- Kumar A, Chauhan S (2017a) Monte Carlo method based QSAR modelling of natural lipase inhibitors using hybrid optimal descriptors. *SAR QSAR Environ Res* 28(3):179–197. <https://doi.org/10.1080/1062936X.2017.1293729>
- Kumar A, Chauhan S (2017b) Use of the monte carlo method for OECD principles-guided QSAR modeling of SIRT1 inhibitors. *Arch Pharm*. <https://doi.org/10.1002/ardp.201600268>
- Kumar A, Chauhan S (2018) Use of simplified molecular input line entry system and molecular graph based descriptors in prediction and design of pancreatic lipase inhibitors. *Fut Med Chem* 10(13):1603–1622. <https://doi.org/10.4155/fmc-2018-0024>
- Kumar P, Kumar A (2019) Nucleobase sequence based building up of reliable QSAR models with the index of ideality correlation using Monte Carlo method. *J Biomol Struct Dyn*. <https://doi.org/10.1080/07391102.2019.1656109>
- Kumar P, Kumar A (2020) CORAL: QSAR models of CB1 cannabinoid receptor inhibitors based on local and global SMILES attributes with the index of ideality of correlation and the correlation contradiction index. *Chemomet Intell Lab Syst* 200:103982. <https://doi.org/10.1016/j.chemolab.2020.103982>
- Kumar P, Kumar A, Sindhu J (2019) Design and development of novel focal adhesion kinase (FAK) inhibitors using Monte Carlo method with index of ideality of correlation to validate QSAR. *SAR QSAR Environ Res* 30(2):63–80. <https://doi.org/10.1080/1062936X.2018.1564067>
- Kumar A, Manisha, Bagri K, Kumar P (2020) Use of graph based descriptors for determination of structural features causing

- modulation of fructose-1,6-bisphosphatase. *Drug Res* 70(5):226–232. <https://doi.org/10.1055/a-1138-8725>
- Langman LJ, Snozek CLH (2019) Introduction to drugs of abuse. In: Dasgupta A (ed) Critical issues in alcohol and drugs of abuse testing, 2nd edn. Elsevier Academic Press, London, pp 71–78
- Liechti ME (2003) “Ecstasy” (MDMA): pharmakologie, toxikologie und behandlung der akuten intoxikation (“ecstasy” (MDMA): pharmacology, toxicology, and treatment of acute intoxication (1946). *Dtsch Med Wochenschr* 128(24):1361–1366. <https://doi.org/10.1055/s-2003-39975>
- Luethi D, Liechti ME (2020) Designer drugs: mechanism of action and adverse effects. *Arch Toxicol.* <https://doi.org/10.1007/s00204-020-02693-7>
- Manisha, Chauhan S, Kumar P, Kumar A (2019) Development of prediction model for fructose- 1,6-bisphosphatase inhibitors using the Monte Carlo method. *SAR QSAR Environ Res* 30(3):145–159. <https://doi.org/10.1080/1062936X.2019.1568299>
- Maurer HH, Kraemer T, Springer D, Staack RF (2004) Chemistry, pharmacology, toxicology, and hepatic metabolism of designer drugs of the amphetamine (ecstasy), piperazine, and pyrrolidinophenone types: a synopsis. *Ther Drug Monit* 26(2):127–131. <https://doi.org/10.1097/00007691-200404000-00007>
- Negus SS, Banks ML (2017) Decoding the structure of abuse potential for new psychoactive substances: structure-activity relationships for abuse-related effects of 4-substituted methcathinone analogs. *Curr Top Behav Neurosci* 32:119–131. https://doi.org/10.1007/7854_2016_18
- Nimbal M, Bagri K, Kumar P, Kumar A (2020) The index of ideality of correlation: a statistical yardstick for better QSAR modeling of glucokinase activators. *Struct Chem* 31(2):831–839. <https://doi.org/10.1007/s11224-019-01468-w>
- O’Boyle NM, Banck M, James CA, Morley C, Vandermeersch T, Hutchison GR (2011) Open babel: an open chemical toolbox. *J Cheminform* 3:33. <https://doi.org/10.1186/1758-2946-3-33>
- Ojha PK, Mitra I, Das RN, Roy K (2011) Further exploring rm2 metrics for validation of QSPR models. *Chemomet Intell Lab Syst* 107(1):194–205. <https://doi.org/10.1016/j.chemolab.2011.03.011>
- Papa E, van der Wal L, Arnot JA, Gramatica P (2014) Metabolic biotransformation half-lives in fish: QSAR modeling and consensus analysis. *Sci Total Environ* 470–471:1040–1046. <https://doi.org/10.1016/j.scitotenv.2013.10.068>
- Richter LHJ, Meyer MR, Maurer HH (2019) Overview of common designer drugs. In: Dasgupta A (ed) Critical issues in alcohol and drugs of abuse testing, 2nd edn. Elsevier Academic Press, London, pp 237–246
- Rudnick G, Clark J (1993) From synapse to vesicle: the reuptake and storage of biogenic amine neurotransmitters. *Biochimica et Biophysica Acta (BBA) Bioenerget* 1144(3):249–263. [https://doi.org/10.1016/0005-2728\(93\)90109-S](https://doi.org/10.1016/0005-2728(93)90109-S)
- Ruiz IL, Gómez-Nieto MÁ (2018) Study of the applicability domain of the QSAR classification models by means of the rivalry and modifiability indexes. *Mol (Basel Switzerl)*. <https://doi.org/10.3390/molecules23112756>
- Simmler LD, Hysek CM, Liechti ME (2011) Sex differences in the effects of MDMA (ecstasy) on plasma copeptin in healthy subjects. *J Clin Endocrinol Metab* 96(9):2844–2850. <https://doi.org/10.1210/jc.2011-1143>
- Simmler LD, Buser TA, Donzelli M, Schramm Y, Dieu L-H, Huwyler J, Chaboz S, Hoener MC, Liechti ME (2013a) Pharmacological characterization of designer cathinones in vitro. *Br J Pharmacol* 168(2):458–470. <https://doi.org/10.1111/j.1476-5381.2012.02145.x>
- Simmler LD, Wandeler R, Liechti ME (2013b) Bupropion, methylphenidate, and 3,4-methylenedioxypyrovalerone antagonize methamphetamine-induced efflux of dopamine according to their potencies as dopamine uptake inhibitors: implications for the treatment of methamphetamine dependence. *BMC Res Notes* 6:220. <https://doi.org/10.1186/1756-0500-6-220>
- Toma C, Gadaleta D, Roncaglioni A, Toropov A, Toropova A, Marzo M, Benfenati E (2018) QSAR development for plasma protein binding: influence of the ionization state. *Pharm Res* 36(2):28. <https://doi.org/10.1007/s11095-018-2561-8>
- Toropov AA, Toropova AP (2018) Predicting cytotoxicity of 2-phenylindole derivatives against breast cancer cells using index of ideality of correlation. *Anticancer Res* 38(11):6189–6194. <https://doi.org/10.21873/anticancres.12972>
- Toropov AA, Toropova AP (2019) Use of the index of ideality of correlation to improve predictive potential for biochemical endpoints. *Toxicol Mech Methods* 29(1):43–52. <https://doi.org/10.1080/15376516.2018.1506851>
- Toropov AA, Toropova AP, Martyanov SE, Benfenati E, Gini G, Leszczynska D, Leszczynski J (2011) Comparison of SMILES and molecular graphs as the representation of the molecular structure for QSAR analysis for mutagenic potential of polycyclic aromatic amines. *Chemomet Intell Lab Syst* 109(1):94–100. <https://doi.org/10.1016/j.chemolab.2011.07.008>
- Toropov AA, Raška I, Toropova AP, Raškova M, Veselinović AM, Veselinović JB (2019a) The study of the index of ideality of correlation as a new criterion of predictive potential of QSPR/QSAR-models. *Sci Total Environ* 659:1387–1394. <https://doi.org/10.1016/j.scitotenv.2018.12.439>
- Toropov AA, Toropova AP, Raitano G, Benfenati E (2019b) CORAL: Building up QSAR models for the chromosome aberration test. *Saudi J Biol Sci* 26(6):1101–1106. <https://doi.org/10.1016/j.sjbs.2018.05.013>
- Toropova AP, Toropov AA (2017) The index of ideality of correlation: a criterion of predictability of QSAR models for skin permeability? *Sci Total Environ* 586:466–472. <https://doi.org/10.1016/j.scitotenv.2017.01.198>
- Toropova AP, Toropov AA, Rasulev BF, Benfenati E, Gini G, Leszczynska D, Leszczynski J (2012) QSAR models for ACE-inhibitor activity of tri-peptides based on representation of the molecular structure by graph of atomic orbitals and SMILES. *Struct Chem* 23(6):1873–1878. <https://doi.org/10.1007/s11224-012-9996-z>
- Toropova AP, Toropov AA, Leszczynska D, Leszczynski J (2018) The index of ideality of correlation: hierarchy of Monte Carlo models for glass transition temperatures of polymers. *J Polym Res* 25(10):541. <https://doi.org/10.1007/s10965-018-1618-z>
- Walther D, Shalabi AR, Baumann MH, Glennon RA (2019) Systematic structure-activity studies on selected 2-, 3-, and 4-monosubstituted synthetic methcathinone analogs as monoamine transporter releasing agents. *ACS Chem Neurosci* 10(1):740–745. <https://doi.org/10.1021/acscchemneuro.8b00524>
- Weaver MF, Hopper JA, Gunderson EW (2015) Designer drugs 2015: assessment and management. *Addict Sci Clin Pract* 10:8. <https://doi.org/10.1186/s13722-015-0024-7>
- Wee S, Anderson KG, Baumann MH, Rothman RB, Blough BE, Woolverton WL (2005) Relationship between the serotonergic activity and reinforcing effects of a series of amphetamine analogs. *J Pharmacol Exp Thera* 313(2):848–854. <https://doi.org/10.1124/jpet.104.080101>
- Zvinavashe E, Murk AJ, Rietjens IMCM (2008) Promises and pitfalls of quantitative structure-activity relationship approaches for predicting metabolism and toxicity. *Chem Res Toxicol* 21(12):2229–2236. <https://doi.org/10.1021/tx800252e>



Identification of good and bad fragments of tricyclic triazinone analogues as potential PKC-θ inhibitors through SMILES-based QSAR and molecular docking

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Abstract

Based on the mechanism of action of PKC-θ, the inhibition of this enzyme is considered a potential target for the treatment of autoimmune diseases such as rheumatoid arthritis (RA), inflammatory bowel disease (IBD), and psoriasis. In the present study, 57 structurally diverse tricyclic triazinone analogues as potential PKC-θ inhibitors has been taken into consideration for QSAR analysis through Monte Carlo optimization. QSAR models are developed using the balance of correlation method in the CORAL software with two target functions (TF₁ and TF₂). The models constructed with IIC are found more robust and authentic. The established QSAR model with best $R^2_{\text{calibration}} = 0.9653$ for split 3 is considered the topmost model. The predictabilities of the recommended QSAR model are assessed through various statistical parameters. The outlier of each model is also identified using the applicability domain (AD). The common mechanistic interpretation of the increasing and decreasing attributes has been extracted by evaluating the correlation weights of diverse structural attributes obtained in three Monte Carlo optimization runs from two splits, i.e., split 3 and 4. In the last, the result of mechanistic interpretation is validated by performing the docking studies of compounds PKC03, PKC07, PKC16, PKC25, and PKC56 in the experimental structure of protein kinase C-θ (PDB ID: 4Q9Z). The numerical value of the correlation coefficient between calculated activity and binding affinity is found 0.9597. Hence, the developed QSAR models are descriptive and predictive in nature and the results are in sound agreement with the experimental observations.

Keywords Protein kinase C-θ · QSAR · CORAL software · IIC · Docking

Introduction

The immune system is an intricate network of biochemical aspects that warrants the integrity of the organism by attacking potential pathogens [1]. However, sometimes, our immune system overreacts and attacks at the organism itself. Therefore, the abnormalities in the immune system, mainly the over-activation of T cells, lead to autoimmune and other secondary diseases [2, 3]. It has been cited in the literature that

humans and mice lacking functional regulatory T cells (Tregs) due to mutations in the Foxp3 gene give way to severe lymphoproliferative and inflammatory disease [4–8]. Considering the significant role of T lymphocytes in monitoring and facilitating different types of immune reactions, the T cells are considered major drug targets for treating immunological disorders [9]. The unsatisfactory results with various side effects of the calcineurin inhibitor drugs, such as cyclosporine A (tacrolimus), lead to the development of new immunosuppressive non-calcineurin inhibitors, especially the PKC-θ inhibitors [10].

The protein kinase C (PKC) enzyme family executes an important part in signal transduction pathways that affect cell proliferation and differentiation. The protein kinase C theta (PKC-θ) is a novel member of Ca²⁺-independent novel PKC family and it is predominantly expressed in lymphocytes (T cells) and mast cells [11, 12]. The novel isoform PKC-θ performs a significant function in the activation and survival of T cells by transmitting the T cell receptor (TCR) signaling. Literature survey reveals that the response of PKC-θ deficient

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Comparison of the results with reported models

Literature survey reveals that only one manuscript by Lingwei et al. describing the QSAR study of 54 tricyclic triazinone analogues as potential PKC-θ inhibitors is available [45]. The comparison of the present study and the literature report is given in Table 6. Lingwei et al. have developed the QSAR models based on the CoMFA, CoMSIA, HQSAR, and 2D-QSAR study. The reported QSAR models are quite complex. The reported QSAR models are constructed with six or seven variables, i.e., multiparametric model. The numerical value of R^2 and Q^2 for the training set of reported QSAR models is higher than the present leading QSAR model. On the other hand, the actual predictive power of a QSAR model can be evaluated by the test set/calibration set. The value of the determination coefficient R_{cal}^2 of the QSAR models constructed by TF₂ for split 3 is 0.9653 which is more than all the reported models. The present work describes the QSAR modeling of 57 tricyclic triazinone analogues using four splits because “QSAR is a random event” while the formerly reported models are developed only using one split. All QSAR models or present research work are statistically significant and monoparametric which makes their interpretation very easy compared with the other reported models. In the earlier report, the statistical parameters IIC, CCC, Q^2_{F2} , and Q^2_{F3} are not reported. In the present QSAR modeling, the structural attributes responsible for the increase and decrease of the endpoint are also reported.

Conclusion

The present research demonstrates the development of new robust and reliable QSAR models for tricyclic triazinone analogues as PKC-θ inhibitors based on the Monte Carlo optimization method. The SMILES were used to symbolize the chemical structures of tricyclic triazinone analogues. The QSAR models were developed using the balance of correlation method in the CORAL software with two target functions (TF₁ and TF₂). The IIC was applied to improve the robustness and predictability of the developed QSAR models with second target function (TF2). The models constructed with IIC were found more robust and authentic. The established QSAR model with best $R_{\text{calibration}}^2 = 0.9653$ for split 3 was deliberated as the topmost model. The predictabilities of the recommended QSAR model were assessed through various statistical parameters such as R^2 , CCC, IIC, Q^2 , Q^2_{F1} , Q^2_{F2} , Q^2_{F3} , s , MAE, F , RMSE, R_m^2 , ΔR_m^2 , $C_{R_p^2}$, and Y test. However, all of the established models were appropriate to predict the PKC-θ inhibition of tricyclic triazinone analogues. The outlier of each model was also identified using the applicability domain

(AD). The outliers were absent in all QSAR models developed by TF₁. However, in the case of the QSAR models constructed by TF₂, the number of outliers was 10 and 9 for split 1 and split 3, respectively, while in other splits, outliers were absent. The common mechanistic interpretation of the increasing and decreasing attributes had been extracted by evaluating the correlation weights of diverse structural attributes obtained in three Monte Carlo optimization runs from two splits, i.e., split 3 and 4. In the last, the result of mechanistic interpretation was validated by performing the docking studies of compounds PKC03, PKC07, PKC16, PKC25, and PKC56 in the experimental structure of protein kinase C theta (PDB ID: 4Q9Z). The numerical value of the correlation coefficient between calculated activity and binding affinity was found 0.9597 while it was 0.9837 between experimental activity and binding affinity. Hence, the developed QSAR models were descriptive and predictive in nature and the results were in sound consensus with the experimental observations.

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Authors' contributions Authors have done equivalent contributions to this work.

Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

References

1. Zanin-Zhorov A, Kumari S, Hippen KL, Merkel SC, MacMillan ML, Blazar BR, Dustin ML (2017) Human in vitro-induced regulatory T cells display Dlgh1-dependent and PKC-theta restrained suppressive activity. *Sci Rep* 7(1):4258. <https://doi.org/10.1038/s41598-017-04053-5>
2. Han C, Lei D, Liu L, Xie S, He L, Wen S, Zhou H, Ma T, Li S (2020) Morphine induces the differentiation of T helper cells to Th2 effector cells via the PKC-theta-GATA3 pathway. *Int Immunopharmacol* 80:106133. <https://doi.org/10.1016/j.intimp.2019.106133>
3. von Essen MR, Kongsbak M, Levring TB, Hansen AK, Boding L, Lauritsen JP, Woetmann A, Baier G, Odum N, Bonefeld CM, Geisler C (2013) PKC-theta exists in an oxidized inactive form in naïve human T cells. *Eur J Immunol* 43(6):1659–1666. <https://doi.org/10.1002/eji.201243140>
4. Wachowicz K, Baier G (2014) Protein kinase C theta: the pleiotropic T-cell signalling intermediate. *Biochem Soc Trans* 42(6):1512–1518. <https://doi.org/10.1042/BST20140179>
5. Cole DC, Asselin M, Brennan A, Czerwinski R, Ellingboe JW, Fitz L, Greco R, Huang X, Joseph-McCarthy D, Kelly MF, Kirisits M, Lee J, Li Y, Morgan P, Stock JR, Tsao DH, Wissner A, Yang X, Chaudhary D (2008) Identification, characterization and initial hit-to-lead optimization of a series of 4-arylamino-3-pyridinecarbonitrile as protein kinase C theta (PKCtheta) inhibitors.

- J Med Chem 51(19):5958–5963. <https://doi.org/10.1021/jm800214a>
6. Sakowicz-Burkiewicz M, Nishanth G, Helmuth U, Drogemuller K, Busch DH, Utermohlen O, Naumann M, Deckert M, Schluter D (2008) Protein kinase C-theta critically regulates the proliferation and survival of pathogen-specific T cells in murine listeriosis. J Immunol 180(8):5601–5612. <https://doi.org/10.4049/jimmunol.180.8.5601>
 7. Solomou EE, Juang YT, Tsokos GC (2001) Protein kinase C-theta participates in the activation of cyclic AMP-responsive element-binding protein and its subsequent binding to the -180 site of the IL-2 promoter in normal human T lymphocytes. J Immunol 166(9): 5665–5674. <https://doi.org/10.4049/jimmunol.166.9.5665>
 8. Thuille N, Siegmund K, Klepsch V, Schorgenhuber J, Danklmaier S, Leitges M, Baier G (2019) Loss-of-function phenotype of a PKCtheta(T219A) knockin mouse strain. Cell Commun Signal 17(1):141. <https://doi.org/10.1186/s12964-019-0466-8>
 9. Cywin CL, Dahmann G, Prokopowicz 3rd AS, Young ER, Magolda RL, Cardozo MG, Cogan DA, Disalvo D, Ginn JD, Kashem MA, Wolak JP, Honon CA, Farrell TM, Grbic H, Hu H, Kaplita PV, Liu LH, Spero DM, Jeanfavre DD, O'Shea KM, White DM, Woska Jr JR, Brown ML (2007) Discovery of potent and selective PKC-theta inhibitors. Bioorg Med Chem Lett 17(1): 225–230. <https://doi.org/10.1016/j.bmcl.2006.09.056>
 10. Tanaka Y, Altman A (2002) T cell signaling: protein kinase Cθ the immunological synapse and characterization of SLAT a novel T helper 2-specific adapter protein. Allergol Int 51 (3):167–174. <https://doi.org/10.1046/j.1440-1592.2002.00261.x>
 11. Niu C, Boschelli DH, Tumey LN, Bhagirath N, Subrath J, Shim J, Wang Y, Wu B, Eid C, Lee J, Yang X, Brennan A, Chaudhary D (2009) First generation 5-vinyl-3-pyridinecarbonitrile PKCtheta inhibitors. Bioorg Med Chem Lett 19(20):5829–5832. <https://doi.org/10.1016/j.bmcl.2009.08.086>
 12. Salek-Ardakani S, So T, Halteman BS, Altman A, Croft M (2004) Differential regulation of Th2 and Th1 lung inflammatory responses by protein kinase C theta. J Immunol 173(10):6440–6447. <https://doi.org/10.4049/jimmunol.173.10.6440>
 13. Healy AM, Izmailova E, Fitzgerald M, Walker R, Hattersley M, Silva M, Siebert E, Terkelsen J, Picarella D, Pickard MD, LeClair B, Chandra S, Jaffee B (2006) PKC-theta-deficient mice are protected from Th1-dependent antigen-induced arthritis. J Immunol 177(3):1886–1893. <https://doi.org/10.4049/jimmunol.177.3.1886>
 14. Salek-Ardakani S, So T, Halteman BS, Altman A, Croft M (2005) Protein kinase Ctheta controls Th1 cells in experimental autoimmune encephalomyelitis. J Immunol 175(11):7635–7641. <https://doi.org/10.4049/jimmunol.175.11.7635>
 15. Tan SL, Zhao J, Bi C, Chen XC, Hepburn DL, Wang J, Sedgwick JD, Chintalacharuvu SR, Na S (2006) Resistance to experimental autoimmune encephalomyelitis and impaired IL-17 production in protein kinase C theta-deficient mice. J Immunol 176(5):2872–2879. <https://doi.org/10.4049/jimmunol.176.5.2872>
 16. Nagahama K, Ogawa A, Shirane K, Shimomura Y, Sugimoto K, Mizoguchi A (2008) Protein kinase C theta plays a fundamental role in different types of chronic colitis. Gastroenterology 134(2): 459–469. <https://doi.org/10.1053/j.gastro.2007.11.005>
 17. Dushin RG, Nittoli T, Ingalls C, Boschelli DH, Cole DC, Wissner A, Lee J, Yang X, Morgan P, Brennan A, Chaudhary D (2009) Synthesis and PKCtheta inhibitory activity of a series of 4-indolylamino-5-phenyl-3-pyridinecarbonitriles. Bioorg Med Chem Lett 19(9):2461–2463. <https://doi.org/10.1016/j.bmcl.2009.03.053>
 18. Tumey LN, Bhagirath N, Brennan A, Brooijmans N, Lee J, Yang X, Boschelli DH (2009) 5-Vinyl-3-pyridinecarbonitrile inhibitors of PKCtheta: optimization of enzymatic and functional activity. Bioorg Med Chem 17(23):7933–7948. <https://doi.org/10.1016/j.bmcl.2009.10.020>
 19. Basak SC, Vracko MG (2020) Parsimony principle and its proper use/ application in computer-assisted drug design and QSAR. Curr Comput Aided Drug Des 16(1):1–5. <https://doi.org/10.2174/157340991601200106122854>
 20. Veeravarapu H, Malkhed V, Mustyala KK, Vadija R, Malikanti R, Viruputuri U, Muthyalu MKK (2020) Structure-based drug design, synthesis and screening of MmaA1 inhibitors as novel anti-TB agents. Mol Divers. <https://doi.org/10.1007/s11030-020-10107-0>
 21. Senthil R, Sakthivel M, Usha S (2020) Structure-based drug design of peroxisome proliferator-activated receptor gamma inhibitors: ferulic acid and derivatives. J Biomol Struct Dyn:On Line Published. <https://doi.org/10.1080/07391102.2020.1740790>
 22. Sadhasivam A, Nagarajan H, Umashankar V (2020) Structure-based drug target prioritisation and rational drug design for targeting Chlamydia trachomatis eye infections. J Biomol Struct Dyn 38(11):3131–3143. <https://doi.org/10.1080/07391102.2019.1652691>
 23. Kumar P, Kumar A (2020) CORAL: QSAR models of CB1 cannabinoid receptor inhibitors based on local and global SMILES attributes with the index of ideality of correlation and the correlation contradiction index. Chemometr Intel Lab Syst 200:103982. <https://doi.org/10.1016/j.chemolab.2020.103982>
 24. Kumar A, Sindhu J, Kumar P (2020) In-silico identification of fingerprint of pyrazolyl sulfonamide responsible for inhibition of N-myristoyltransferase using Monte Carlo method with index of ideality of correlation. J Biomol Struct Dyn:On Line Published. <https://doi.org/10.1080/07391102.2020.1784286>
 25. Puratchikody A, Prabu SL, Umamaheswari A (2019) Computer applications in drug discovery and development. Advances in medical technologies and clinical practice (AMTCP) book series. Medical Information Science Reference, Hershey, PA
 26. Worachartcheewan A, Mandi P, Prachayasittikul V, Toropova AP, Toropov AA, Nantasesamat C (2014) Large-scale QSAR study of aromatase inhibitors using SMILES-based descriptors. Chemometr Intel Lab Syst 138:120–126. <https://doi.org/10.1016/j.chemolab.2014.07.017>
 27. Doucet J-P, Panaye A (2010) Three dimensional QSAR : applications in pharmacology and toxicology. QSAR in environmental and health sciences. CRC Press, Boca Raton
 28. Qi R, Pan Y, Cao J, Jia Z, Jiang J (2020) The cytotoxicity of nanomaterials: modeling multiple human cells uptake of functionalized magneto-fluorescent nanoparticles via nano-QSAR. Chemosphere 249:126175. <https://doi.org/10.1016/j.chemosphere.2020.126175>
 29. Mondal D, Ghosh K, Baidya ATK, Gantait AM, Gayen S (2020) Identification of structural fingerprints for in vivo toxicity by using Monte Carlo based QSTR modeling of nitroaromatics. Toxicol Mechan Methods 30 (4):257–265. <https://doi.org/10.1080/15376516.2019.1709238>
 30. Jain S, Bhardwaj B, Amin SA, Adhikari N, Jha T, Gayen S (2020) Exploration of good and bad structural fingerprints for inhibition of indoleamine-2,3-dioxygenase enzyme in cancer immunotherapy using Monte Carlo optimization and Bayesian classification QSAR modeling. J Biomol Struct Dyn 38 (6):1683–1696. <https://doi.org/10.1080/07391102.2019.1615000>
 31. Ahmadi S, Ghanbari H, Lotfi S, Azimi N (2020) Predictive QSAR modeling for the antioxidant activity of natural compounds derivatives based on Monte Carlo method. Mol Divers. <https://doi.org/10.1007/s11030-019-10026-9>
 32. Toropov AA, Toropova AP, Selvestrel G, Benfenati E (2019) Idealization of correlations between optimal simplified molecular input-line entry system-based descriptors and skin sensitization. SAR QSAR Environ Res 30(6):447–455. <https://doi.org/10.1080/1062936X.2019.1615547>
 33. Toropova MA, Veselinovic AM, Veselinovic JB, Stojanovic DB, Toropov AA (2015) QSAR modeling of the antimicrobial activity

- of peptides as a mathematical function of a sequence of amino acids. *Comput Biol Chem* 59 Pt A:126–130. <https://doi.org/10.1016/j.compbiochem.2015.09.009>
34. Toropova AP, Schultz TW, Toropov AA (2016) Building up a QSAR model for toxicity toward Tetrahymena pyriformis by the Monte Carlo method: a case of benzene derivatives. *Environ Toxicol Pharmacol* 42:135–145. <https://doi.org/10.1016/j.etap.2016.01.010>
 35. Toropova AP, Toropov AA, Veselinovic JB, Miljkovic FN, Veselinovic AM (2014) QSAR models for HEPT derivates as NNRTI inhibitors based on Monte Carlo method. *Eur J Med Chem* 77:298–305. <https://doi.org/10.1016/j.ejmech.2014.03.013>
 36. Ibezim E, Duchowicz PR, Ortiz EV, Castro EA (2012) QSAR on aryl-piperazine derivatives with activity on malaria. *Chemometr Intel Lab Syst* 110(1):81–88. <https://doi.org/10.1016/j.chemolab.2011.10.002>
 37. Kumar A, Manisha, Bagri K, Kumar P (2020) Use of graph based descriptors for determination of structural features causing modulation of fructose-1,6-bisphosphatase. *Drug Res (Stuttg)* 70 (5):226–232. <https://doi.org/10.1055/a-1138-8725>
 38. Kumar P, Kumar A, Sindhu J (2019) Design and development of novel focal adhesion kinase (FAK) inhibitors using Monte Carlo method with index of ideality of correlation to validate QSAR. *SAR QSAR Environ Res* 30 (2):63–80. <https://doi.org/10.1080/1062936X.2018.1564067>
 39. Kumar P, Kumar A, Sindhu J (2019) In silico design of diacylglycerol acyltransferase-1 (DGAT1) inhibitors based on SMILES descriptors using Monte-Carlo method. *SAR QSAR Environ Res* 30(8):525–541. <https://doi.org/10.1080/1062936X.2019.1629998>
 40. Toropova AP, Toropov AA, Carnesecchi E, Benfenati E, Dorne JL (2020) The index of ideality of correlation: models for flammability of binary liquid mixtures. *Chem Pap* 74(2):601–609. <https://doi.org/10.1007/s11696-019-00903-w>
 41. Kumar A, Kumar P (2020) Construction of pioneering quantitative structure activity relationship screening models for abuse potential of designer drugs using index of ideality of correlation in Monte Carlo optimization. *Arch Toxicol* 94 (9):3069–3086. <https://doi.org/10.1007/s00204-020-02828-w>
 42. Ahmadi S (2020) Mathematical modeling of cytotoxicity of metal oxide nanoparticles using the index of ideality correlation criteria. *Chemosphere* 242:125192. <https://doi.org/10.1016/j.chemosphere.2019.125192>
 43. Kumar P, Kumar A, Sindhu J, Lal S (2019) QSAR models for nitrogen containing monophosphonate and bisphosphonate derivatives as human farnesyl pyrophosphate synthase inhibitors based on Monte Carlo method. *Drug Res (Stuttg)* 69 (3):159–167. <https://doi.org/10.1055/a-0652-5290>
 44. Achary PGR, Toropova AP, Toropov AA (2019) Combinations of graph invariants and attributes of simplified molecular input-line entry system (SMILES) to build up models for sweetness. *Food Res Int* 122:40–46. <https://doi.org/10.1016/j.foodres.2019.03.067>
 45. Meng L, Feng K, Ren Y (2018) Molecular modelling studies of tricyclic triazinone analogues as potential PKC-θ inhibitors through combined QSAR, molecular docking and molecular dynamics simulations techniques. *J Taiwan Inst Chem Eng* 91:155–175. <https://doi.org/10.1016/j.jtice.2018.06.017>
 46. George DM, Breinlinger EC, Argiriadi MA, Zhang Y, Wang J, Bansal-Pakala P, Duignan DB, Honore P, Lang Q, Mittelstadt S, Rundell L, Schwartz A, Sun J, Edmunds JJ (2015) Optimized protein kinase C theta (PKCtheta) inhibitors reveal only modest anti-inflammatory efficacy in a rodent model of arthritis. *J Med Chem* 58(1):333–346. <https://doi.org/10.1021/jm5013006>
 47. George DM, Breinlinger EC, Friedman M, Zhang Y, Wang J, Argiriadi M, Bansal-Pakala P, Barth M, Duignan DB, Honore P, Lang Q, Mittelstadt S, Potin D, Rundell L, Edmunds JJ (2015) Discovery of selective and orally bioavailable protein kinase C theta (PKCtheta) inhibitors from a fragment hit. *J Med Chem* 58(1):222–236. <https://doi.org/10.1021/jm500669m>
 48. Marvin-Sketch-v.14.11.17.0 (2014). ChemAxon, XchemAxon KFT. Budapest, Hungary
 49. O'Boyle NM, Banck M, James CA, Morley C, Vandermeersch T, Hutchison GR (2011) Open babel: an open chemical toolbox. *J Cheminform* 3 (1):33. <https://doi.org/10.1186/1758-2946-3-33>
 50. Veselinovic JB, Nikolic GM, Trutic NV, Zivkovic JV, Veselinovic AM (2015) Monte Carlo QSAR models for predicting organophosphate inhibition of acetylcholinesterase. *SAR QSAR Environ Res* 26(6):449–460. <https://doi.org/10.1080/1062936X.2015.1049665>
 51. Gissi A, Toropov AA, Toropova AP, Nicolotti O, Carotti A, Benfenati E (2014) Building up QSAR model for toxicity of psychotropic drugs by the Monte Carlo method. *Struct Chem* 25(4): 1067–1073. <https://doi.org/10.1007/s11224-013-0380-4>
 52. Kumar A, Chauhan S (2017) Monte Carlo method based QSAR modelling of natural lipase inhibitors using hybrid optimal descriptors. *SAR QSAR Environ Res* 28(3):179–197. <https://doi.org/10.1080/1062936X.2017.1293729>
 53. Manisha CS, Kumar P, Kumar A (2019) Development of prediction model for fructose-1,6- bisphosphatase inhibitors using the Monte Carlo method. *SAR QSAR Environ Res* 30(3):145–159. <https://doi.org/10.1080/1062936X.2019.1568299>
 54. Toropova AP, Toropov AA (2019) QSPPR and nano-QSPPR: what is the difference? *J Mol Struct* 1182:141–149. <https://doi.org/10.1016/j.molstruc.2019.01.040>
 55. Toropov AA, Toropova AP (2017) The index of ideality of correlation: a criterion of predictive potential of QSPPR/QSAR models? *Mutat Res Genet Toxicol Environ Mutagen* 819:31–37. <https://doi.org/10.1016/j.mrgentox.2017.05.008>
 56. Toropov AA, Toropova AP (2018) Predicting cytotoxicity of 2-Phenylindole derivatives against breast cancer cells using index of ideality of correlation. *Anticancer Res* 38(11):6189–6194. <https://doi.org/10.21873/anticancres.12972>
 57. Environment-directorate guidance document on the validation of (quantitative) structure-activity relationship [(Q)SAR] models. Organisation for Economic Co-operation and Development. [http://www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?doclanguage=en&cote=env/jm/mono\(2007\)2](http://www.oecd.org/officialdocuments/publicdisplaydocumentpdf/?doclanguage=en&cote=env/jm/mono(2007)2). Accessed 07 July 2020
 58. Aher RB, Roy K (2017) Exploring the structural requirements in multiple chemical scaffolds for the selective inhibition of *Plasmodium falciparum* calcium-dependent protein kinase-1 (PfCDPK-1) by 3D-pharmacophore modelling, and docking studies. *SAR QSAR Environ Res* 28 (5):390–414. <https://doi.org/10.1080/1062936X.2017.1326401>
 59. Bhayye SS, Roy K, Saha A (2016) Pharmacophore generation, atom-based 3D-QSAR, HQSAR and activity cliff analyses of benzothiazine and deazaxanthine derivatives as dual A2A antagonists/MAOB inhibitors. *SAR QSAR Environ Res* 27(3): 183–202. <https://doi.org/10.1080/1062936X.2015.1136840>
 60. Golbraikh A, Tropsha A (2002) Beware of q2! *J Mol Graph Model* 20(4):269–276. [https://doi.org/10.1016/s1093-3263\(01\)00123-1](https://doi.org/10.1016/s1093-3263(01)00123-1)
 61. Shi LM, Fang H, Tong W, Wu J, Perkins R, Blair RM, Branham WS, Dial SL, Moland CL, Sheehan DM (2001) QSAR models using a large diverse set of estrogens. *J Chem Inf Comput Sci* 41(1):186–195. <https://doi.org/10.1021/ci000066d>
 62. Schuurmann G, Ebert RU, Chen J, Wang B, Kuhne R (2008) External validation and prediction employing the predictive squared correlation coefficient test set activity mean vs training set activity mean. *J Chem Inf Model* 48(11):2140–2145. <https://doi.org/10.1021/ci800253u>
 63. Pratim Roy P, Paul S, Mitra I, Roy K (2009) On two novel parameters for validation of predictive QSAR models. *Molecules* 14(5): 1660–1701. <https://doi.org/10.3390/molecules14051660>

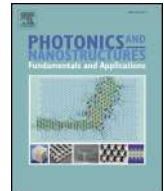
64. Roy K, Chakraborty P, Mitra I, Ojha PK, Kar S, Das RN (2013) Some case studies on application of “ $r(m)2$ ” metrics for judging quality of quantitative structure-activity relationship predictions: emphasis on scaling of response data. *J Comput Chem* 34 (12): 1071–1082. <https://doi.org/10.1002/jec.23231>
65. Gramatica P, Sangion A (2016) A historical excursus on the statistical validation parameters for QSAR models: a clarification concerning metrics and terminology. *J Chem Inf Model* 56 (6): 1127–1131. <https://doi.org/10.1021/acs.jcim.6b00088>
66. Toropova AP, Toropov AA (2019) The index of ideality of correlation: improvement of models for toxicity to algae. *Nat Prod Res* 33(15):2200–2207. <https://doi.org/10.1080/14786419.2018.1493591>
67. Toropov AA, Toropova AP (2019) Use of the index of ideality of correlation to improve predictive potential for biochemical endpoints. *Toxicol Mechan Methods* 29(1):43–52. <https://doi.org/10.1080/15376516.2018.1506851>
68. Duhan M, Singh R, Devi M, Sindhu J, Bhatia R, Kumar A, Kumar P (2019) Synthesis, molecular docking and QSAR study of thiazole clubbed pyrazole hybrid as alpha-amylase inhibitor. *J Biomol Struct Dyn*:1–17. <https://doi.org/10.1080/07391102.2019.1704885>
69. Kumar P, Kumar A (2020) Nucleobase sequence based building up of reliable QSAR models with the index of ideality correlation using Monte Carlo method. *J Biomol Struct Dyn* 38(11):3296–3306. <https://doi.org/10.1080/07391102.2019.1656109>
70. Roy PP, Roy K (2009) QSAR studies of CYP2D6 inhibitor aryloxypropanolamines using 2D and 3D descriptors. *Chem Biol Drug Des* 73(4):442–455. <https://doi.org/10.1111/j.1747-0285.2009.00791.x>
71. Roy K, Mitra I, Kar S, Ojha PK, Das RN, Kabir H (2012) Comparative studies on some metrics for external validation of QSPR models. *J Chem Inf Model* 52(2):396–408. <https://doi.org/10.1021/ci200520g>
72. Golbraikh A, Wang XS, Zhu H, Tropsha A (2012) Predictive QSAR modeling: methods and applications in drug discovery and chemical risk assessment. In: Leszczynski J (ed) *Handbook of computational chemistry*. Springer Netherlands, Dordrecht, pp 1309–1342. https://doi.org/10.1007/978-94-007-0711-5_37
73. Gramatica P (2007) Principles of QSAR models validation: internal and external. *QSAR Comb Sci* 26(5):694–701. <https://doi.org/10.1002/qsar.200610151>
74. Nimbhal M, Bagri K, Kumar P, Kumar A (2020) The index of ideality of correlation: a statistical yardstick for better QSAR modeling of glucokinase activators. *Struct Chem* 31(2):831–839. <https://doi.org/10.1007/s11224-019-01468-w>
75. Toropova AP, Toropov AA (2019) Does the index of ideality of correlation detect the better model correctly? *Mol Inform* 38(8–9): e1800157. <https://doi.org/10.1002/minf.201800157>
76. Rucker C, Rucker G, Meringer M (2007) y -Randomization and its variants in QSPR/QSAR. *J Chem Inf Model* 47 (6):2345–2357. <https://doi.org/10.1021/ci700157b>
77. Ojha PK, Mitra I, Das RN, Roy K (2011) Further exploring $rm2$ metrics for validation of QSPR models. *Chemometr Intel Lab Syst* 107 (1):194–205. <https://doi.org/10.1016/j.chemolab.2011.03.011>
78. Kumar P, Kumar A (2018) Monte Carlo method based QSAR studies of Met kinase inhibitors in compliance with OECD principles. *Drug Res (Stuttg)* 68 (4):189–195. <https://doi.org/10.1055/s-0043-119288>
79. Dassault-Systèmes-BIOVIA (2019). Discovery Studio, San Diego: Dassault Systèmes. Discovery Studio, San Diego: Dassault Systèmes.,

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Control of growth solution on the dimensions of gold nanorods accounted for LSPR sensitivity toward liquid ammonia sensing

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ABSTRACT

In the present study, the effect of dimensions of gold nanorods on its sensing property to detect liquid ammonia was reported. Gold nanorods with two different aspect ratios (GNR1 and GNR2) derived from different lengths and diameters were synthesized using seed-mediated growth method, and the aspect ratio was controlled by changing the silver ion concentration in growth solution. The morphological and size measurement was performed using Transmission Electron Microscopy (TEM), and the average value of aspect ratio (AR) was found to be 3.0 and 3.2 for GNR1 and GNR2, respectively. The characteristics transverse and longitudinal mode of localized surface plasmon resonance (LSPR) have been clearly depicted in UV-vis absorption spectrum of both GNR1 and GNR2. The red shift in longitudinal mode of LSPR from 718 to 732 nm has been observed for GNR with change in aspect ratio from 3.0 to 3.2, respectively. These samples of GNR were tested for liquid ammonia sensing with concentration ranging from 100 to 500 ppm. A clear cut blue shift in longitudinal mode of LSPR of prepared gold nanorod was observed. However, the GNR2 was found to be more sensitive toward liquid ammonia sensing. The origin of such blue shifting and sensitivity of longitudinal mode of LSPR of gold nanorod was explained on the basis of orientation dependence and Dipolar Exciton Coupling Model of coupled plasmon in assemblies of anisotropic plasmonic nanoparticles. With the help of this model, blue shifting in longitudinal plasmon band was correlated with the enhanced formation of H-aggregation induced by dipolar coupling of GNR clusters followed by hydrogen bonding after successive addition of ammonia solution.

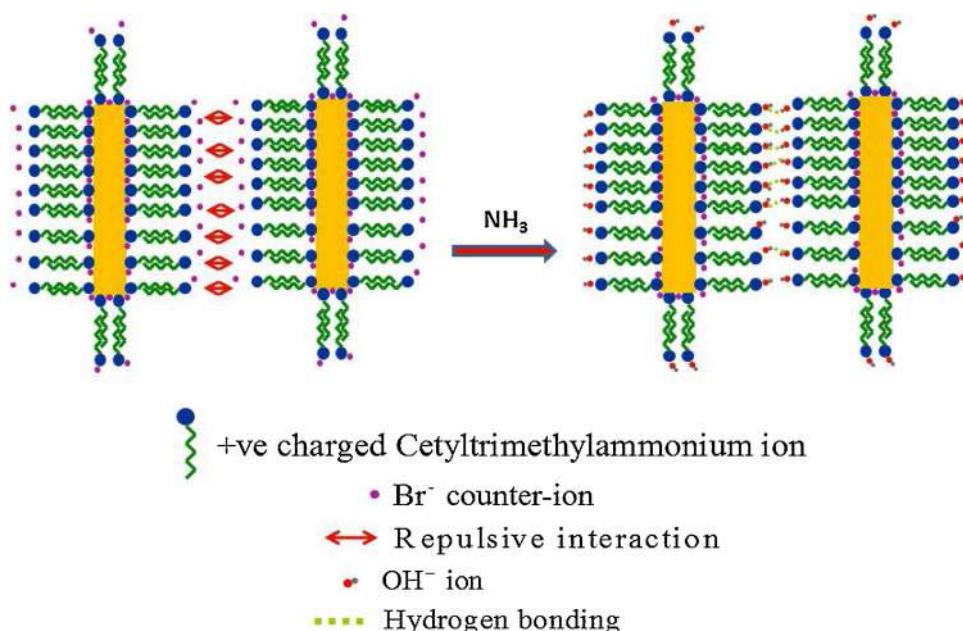
1. Introduction

Over the last decades, nanometer sized structures of noble metal like silver, gold, platinum, etc. have received wide attention due to their special optical response known as surface plasmon resonance (SPR) and localized surface plasmon resonance (LSPR) [1]. Generally, there is a difference in LSPR and SPR; in LSPR, the induced plasmon oscillate locally to the nanostructure upon interaction with incident electromagnetic radiation while in SPR the induced plasmon oscillate along the metal-dielectric interface [2]. Because of this aspect, the decay length of the exponentially decaying electromagnetic field is observed to be shorter in LSPR than that observed in SPR [3,4]. This decay length for plasmon resonance plays vital role and acts as the foundation for biosensing applications. This property reduces the sensitivity response arises due to interference from solution refractive index as well as pH fluctuations whilst providing increased sensitivity to refractive index changes on the surface. Therefore, directing the larger sensitivity of

LSPR to molecular binding and rather than bulk effects. Among various shapes, rod like nanostructures are in a great demand due to anisotropic optical response of LSPR which can be tuned from visible to Infra-red (IR) region of electromagnetic (EM) spectrum, depending upon their aspect ratio [5–7]. The anisotropic optical response relies due to existence of two LSPR absorption band: transverse mode and longitudinal mode of resonance due to the collective oscillation of the quasi-free electrons along the long and short axes of nanorod like structure when excited by EM radiation of comparable wavelength, respectively [8]. The transverse mode is mainly a function of diameter of nanorod observed in a lower side of visible region of EM spectrum and found almost numb to the nanorod morphology. The longitudinal mode observed at higher wavelength side is a function of aspect ratio and refractive index of the surrounding medium and therefore provide tunability of plasmon resonance band covering from green to near IR of EM spectrum wavelength [9,10]. This tunable nature of LSPR benefited from anisotropic geometry of noble metal nanorods provide them a wide

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Scheme 2. Effect of ammonia addition on plasmonic coupling between H-aggregates induced by coupling orientation (c) of scheme 1 .

be more sensitive. The formation of H-aggregates is induced by the formation of microscopic dipole on the surface of GNR, which will change the energy absorption and is thus responsible for plasmon shifting and causing such sensitivity. Thus, on the basis of orientation dependence of coupled plasmon of anisotropic nanoparticles and interaction energy dependence of dipole moment in Dipolar Exciton Coupling Model, the coupling of GNR after the addition of ammonia solution is explained. For nanorod with larger length, such couplings are more, resulting in more shifting in plasmonic wavelength leading to their better sensitivity.

Ethics statement

I certify that this article has not been published previously and is not under consideration for publication elsewhere. There is no conflict of interest regarding this article. I also declare if this article will accepted in this journal, then it will not be published elsewhere in the same form, in English or in any other language, without the written consent of the Publisher.

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References

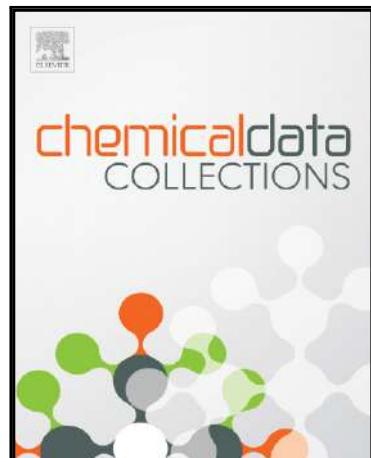
- [1] C.F. Negre, C.G. Sánchez, Metal Clusters and Nanoalloys, Springer, New York, 2013, pp. 105–157.
- [2] J.L. Hammond, N. Bhalla, S.D. Rafiee, P. Estrela, Biosensors 4 (2014) 172–188.
- [3] A.J. Haes, R.P. Van Duyne, Anal. Bioanal. Chem. 379 (2004) 920.
- [4] J.M. Brockman, B.P. Nelson, R.M. Corn, Ann. Rev. Phys. Chem. 51 (2000) 41–63.
- [5] S.S. Verma, J.S. Sekhon, J. Opt. 41 (2012) 89–93.
- [6] C.T. Cooper, M. Rodriguez, S. Blair, J.S. Shumaker-Parry, J. Phys. Chem. C 119 (2015) 11826–11832.
- [7] K.S. Lee, M.A. El-Sayed, J. Phys. Chem. B 109 (2005) 20331–20338.
- [8] J. Cao, T. Sun, K.T. Grattan, Sens. Actuators B Chem. 195 (2014) 332–351.
- [9] S. Link, M.A. El-Sayed, J. Phys. Chem. B 103 (1999) 8410–8426.
- [10] I. Ros, T. Placido, V. Amendola, C. Marinzi, N. Manfredi, R. Comparelli, M. Striccoli, A. Agostiano, A. Abbotto, D. Pedron, R. Pilot, R. Bozio, Plasmonics 9 (2014) 581–593.
- [11] Y. Vlamilidis, V. Voliani, Front. Bioeng. Biotechnol. 6 (143) (2018) 5pp.
- [12] N.G. Khlebtsov, L.A. Dykman, J. Quant. Spectr. Radiat. Transf. 111 (2010) 1–35.
- [13] R.R. Arvizo, S. Bhattacharyya, R.A. Kudgus, K. Giri, R. Bhattacharya, P. Mukherjee, Chem. Soc. Rev. 41 (2012) 2943–2970.
- [14] J. Smolsky, S. Kaur, C. Hayashi, S. Batra, A. Krasnoslobodtsev, Biosensors 7 (2017) 7.
- [15] G. Lu, L. Hou, T. Zhang, W. Li, J. Liu, P. Perriat, Q. Gong, J. Phys. Chem. C 115 (2011) 22877–22885.
- [16] L. Tian, E. Chen, N. Gandra, A. Abbas, S. Singamaneni, Langmuir 28 (2012) 17435–17442.
- [17] H. Chen, L. Shao, K.C. Woo, T. Ming, H.Q. Lin, J. Wang, J. Phys. Chem. C 113 (2009) 17691–17697.
- [18] J. Becker, A. Trügler, A. Jakab, U. Hohenester, C. Sönnichsen, Plasmonics 5 (2010) 161–167.
- [19] M. Rex, F.E. Hernandez, A.D. Campiglia, Anal. Chem. 78 (2006) 445–451.
- [20] C.A. Peng, S. Pachipinde, Nanomater. Nanotechnol. 4 (2014) 9.
- [21] S. Sharma, N. Goswami, N. Gupta, S. Srivastava, Inter. J. Advance. Tech. 3 (2012) 195–202.
- [22] S.T. Dubas, V. Pimpin, Talanta 76 (2008) 29–33.
- [23] B. Timmer, W. Olthuis, A. Van Den Berg, Sens. Actuators B Chem. 107 (2005) 666–677.
- [24] C. Molins-Legua, S. Meseguer-Lloret, Y. Moliner-Martinez, P.A. Campins-Falcó, Trends Analyt. Chem. 25 (2006) 282–290.
- [25] T. Ritthichai, V. Pimpin, J. King Saud Univ. Sci. (2017).
- [26] C. Malins, A. Doyle, B.D. MacCraith, F. Kvasnik, M. Landl, P. Šimon, J. Environ. Monit. 1 (1999) 417–422.
- [27] W. Ament, J.R. Huizenga, E. Kort, T.W. Van Der Mark, R.G. Grevink, G.J. Verkerke, Int. J. Sports Med. 20 (1999) 71–77.
- [28] L.R. Narasimhan, W. Goodman, C.K.N. Patel, Proc. Natl. Acad. Sci. 98 (2001) 4617–4621.
- [29] B. Timmer, W. Olthuis, A. Van Den Berg, Sens. Actuators B Chem. 107 (2005) 666–677.
- [30] Z. Jin, Y. Su, Y. Duan, Sens. Actuators B Chem. 72 (2001) 75–79.
- [31] C. Molins-Legua, S. Meseguer-Lloret, Y. Moliner-Martinez, P. Campins-Falcó, Trends Analyt. Chem. 25 (2006) 282–290.
- [32] J. Li, P.K. Dasgupta, Anal. Chim. Acta 398 (1999) 33–39.
- [33] F. Valentini, V. Biagiotti, C. Lete, G. Palleschi, J. Wang, Sens. Actuators B Chem. 128 (2007) 326–333.
- [34] B. Nikoobakht, M.A. El-Sayed, Chem. Mater. 15 (2003) 1957–1962.
- [35] C.J. Murphy, N.R. Jana, Adv. Mater. 14 (2002) 80–82.
- [36] K.L. Kelly, E. Coronado, L.L. Zhao, G.C. Schatz, J. Phys. Chem. C 107 (2003) 668–677.
- [37] C. Noguez, J. Phys. Chem. C 111 (2007) 3806–3819.

- [38] K.S. Lee, M.A. El-Sayed, *J. Phys. Chem. B* 110 (2006) 19220–19225.
- [39] C. Yu, J. Irudayaraj, *Analytical Chem.* 79 (2007) 572–579.
- [40] P.K. Jain, M.A. El-Sayed, *J. Phys. Chem. C* 111 (2007) 17451–17454.
- [41] P.K. Jain, M.A. El-Sayed, *Nano Lett.* 8 (2008) 4347–4352.
- [42] P.K. Jain, S. Eustis, M.A. El-Sayed, *J. Phys. Chem. B* 110 (2006) 18243–18253.
- [43] S. Link, M.A. El-Sayed, *J. Phys. Chem. B* 109 (2005) 10531–10532.
- [44] J.M. Garnett, *Proc. R. Soc. London* (1904) 385–420.
- [45] J.I. Treu, *Appl. Opt.* 15 (1976) 2746–2750.
- [46] Y.-R. Toh, P. Yu, X. Wen, J. Tang, T.-s. Hsieh, *Nanoscale Res. Lett.* 8 (2013) 103.
- [47] L. Stryer, *Annu Rev. Biochem.* 47 (1978) 819–846.
- [48] S. Saini, G. Srinivas, B. Bagchi, *J. Phys. Chem. B* 113 (7) (2009) 1817–1832.
- [49] S. Bhowmick, S. Saini, V.B. Shenoy, B. Bagchi, *J. Chem. Phys.* 125 (2006) 181102.
- [50] S. Saini, V.B. Shenoy, B. Bagchi, *J. Phys. Chem. C* 112 (16) (2008) 6299–6306.

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Conclusions

The reaction is highly stereospecific and all the products obtained are geometrically pure Z-isomer obtained in higher yield. Fourteen novel aurones were synthesized and screened against MCF-7 cancer cell line for their anticancer activity. All these compounds exhibited moderate to excellent anticancer activity. Among these fourteen compounds, six compounds (**3e**, **3c**, **3i**, **3a**, **3b** and **3n**) have exhibited excellent cytotoxic efficacy against MCF-7 cell line.

Conflict of interest

The authors declare no conflict of interest.

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References

- [1] a) M. Murias, W. Jäger, N. Handler, T. Erker, Z. Horvath, T. Szekeres, H. Nohl, L. Gille, *Biochem. Pharmacol.* 69, (2005), 903-912. <https://doi.org/10.1016/j.bcp.2004.12.001>; b) J. Ferlay, I. Soerjomataram, R. Dikshit, S. Eser, C. Mathers, M. Rebelo, D. M. Parkin, D. Forman, F. Bray, *Int. J. Cancer* 136, (2015), E359-E386. <https://doi.org/10.1002/ijc.29210>.
- [2] R.L. Siegel, K.D. Miller, A. Jemal, *Cancer statistics* 69, (2019), 7-34. <https://doi.org/10.3322/caac.21551>.
- [3] J. Liu, B. Ming, G.-H. Gong, D. Wang, G.-L. Bao, L.-J. Yu, *RSC Adv.* 8, (2018), 4386-4416. <https://doi.org/10.1039/C7RA12912B>.
- [4] A. Saeed, F. A. Larik, P. A. Channar, *Res. Chem. Intermed.* 42, (2016), 6805-6813. <https://doi.org/10.1007/s11164-016-2527-6>.
- [5] M. Leulescu, A. Rotaru, I. Pălărie, A. Moanță, N. Cioateră, M. Popescu, E. Morîntale, M. V. Bubulică, G. Florian, A. Hărăbor, *J. Therm. Anal. Calorim.* 134, (2018), 209-231. <https://doi.org/10.1007/s10973-018-7663-3>.
- [6] S. D. Sawant, G. L. Reddy, M. I. Dar, M. Srinivas, G. Gupta, P. K. Sahu, P. Mahajan, A. Nargotra, S. Singh, S. C. Sharma, *Bioorg. Med. Chem.* 23, (2015), 2121-2128. <https://doi.org/10.1016/j.bmc.2015.03.005>.
- [7] M.A.K.F. Tatsuo, W.M. Carvalho, C.V. Silva, *Inflammation* 18, (1994), 399–405 <https://doi.org/10.1007/BF01534437>
- [8] A. Palomer, F. Cabré, J. Pascual, J. Campos, M. A. Trujillo, A. Entrena, M. A. Gallo, L. García, D. Mauleón, A. Espinosa, *J. Med. Chem.* 45, (2002), 1402-1411. <https://doi.org/10.1021/jm010458r>.
- [9] E. K. Abdelall, G. M. Kamel, *Eur. J. Med. Chem.* 118, (2016), 250-258. <https://doi.org/10.1016/j.ejmech.2016.04.049>.
- [10] a) X. Dai, G. Guo, P. Zou, R. Cui, W. Chen, X. Chen, C. Yin, W. He, R. Vinothkumar, F. Yang, *J. Exp. Clin. Cancer Res.* 36, (2017), 120. <https://doi.org/10.1186/s13046-017-0584-3>; b) M. J. Carr, J. Sun, Z. Eroglu, J. S. Zager, *Expert Opin. Pharmacother.*, 21, (2020), 155-161. <https://doi.org/10.1080/14656566.2019.1694664>; c) M. S. Abdel-Maksoud, M. I. El-Gamal, M. M. G. El-Din, C. H. Oh, *J. Enzyme Inhib. Med. Chem.*, 34,(2019), 97-109. <https://doi.org/10.1080/14756366.2018.1530225>.
- [11] Ş. G. Küçükgüzel, S. Şenkardes, *Eur. J. Med. Chem.* 97, (2015), 786-815. <https://doi.org/10.1016/j.ejmech.2014.11.059>.
- [12] Y. Estevez, M. Quiliano, A. Burguete, B. Cabanillas, M. Zimic, E. Málaga, M. Verástegui, S. Pérez-Silanes, I. Aldana, A. Monge, *Exp. Parasitol.* 127, (2011), 745-751. <https://doi.org/10.1016/j.exppara.2011.01.009>.
- [13] S. Fustero, R. Román, J. F. Sanz-Cervera, A. Simón-Fuentes, J. Bueno, S. Villanova, *J. Org. Chem.* 73, (2008), 8545-8552. <https://doi.org/10.1021/jo801729p>.
- [14] G. M. Nitulescu, L. Matei, I. M. Aldea, C. Draghici, O. T. Olaru, C. Bleotu, *Arabian J. Chem.* 12, (2019), 816-824. <https://doi.org/10.1016/j.arabjc.2015.12.006>.

- [15] F. K. Keter, J. Darkwa, *BioMetals* 25, (2012), 9-21. <https://doi.org/10.1007/s10534-011-9496-4>.
- [16] a) S. M. Gomha, M. A. Abdallah, I. M. Abbas, M. S.H. Kazem, *Med. Chem.* 14, (2018), 344-355, <https://doi.org/10.2174/1573406413666171020114105>; b) S. M. Gomha, M. M. Edrees, R.A.M. Faty *Chem. Cent. J.* 37, (2017), 37, <https://doi.org/10.1186/s13065-017-0266-4>. c) M. A. Abdallah, S. M. Gomha, I. M. Abbas, , M. S. H. Kazem, S. S. Alterary, Y. N. Mabkhot, *Appl. Sci.* 7, (2017), 785, <https://doi.org/10.3390/app7080785>. d) R. Kenchappa, Y.D. Bodke, *Chem. Data Collect.* 28, (2020), 100453. <https://doi.org/10.1016/j.cdc.2020.100453>.
- [17] R. Haudecoeur, A. Boumendjel, *Curr. Med. Chem.* 19, (2012), 2861-2875. <https://doi.org/10.2174/092986712800672085>.
- [18] A.V. Popova, S.P. Bondarenko, M.S. Frasinyuk, *Chem Heterocycl Comp.* 55, (2019), 285–299. <https://doi.org/10.1007/s10593-019-02457-x>.
- [19] C.-Y. Lee, E.-H. Chew, M.-L. Go, *Eur. J. Med. Chem.* 45, (2010), 2957-2971. <https://doi.org/10.1016/j.ejmech.2010.03.023>.
- [20] a) G. S. Hassan, H. H. Georgey, R. F. George, E. R. Mohammed, *Future Med. Chem.* 10, (2018), 27-52. <https://doi.org/10.4155/fmc-2017-0147>; b) A. Detsi, M. Majdalani, C. A. Kontogiorgis, D. Hadjipavlou-Litina, P. Kefalas, *Bioorg. Med. Chem.* 17, (2009), 8073-8085. <https://doi.org/10.1016/j.bmc.2009.10.002>; c) W. Huang, M.-Z. Liu, Y. Li, Y. Tan, G.-F. Yang, *Bioorg. Med. Chem.* 15, (2007), 5191-5197. <https://doi.org/10.1016/j.bmc.2007.05.022>; d) B. P. Bandgar, S. A. Patil, B. L. Korbad, S. C. Biradar, S. N. Nile, C. N. Khobragade, *Eur. J. Med. Chem.* 45, (2010), 3223-3227. <https://doi.org/10.1016/j.ejmech.2010.03.045>; e) M. P. Carrasco, A. S. Newton, L. Gonçalves, A. Góis, M. Machado, J. Gut, F. Nogueira, T. Hänscheid, R. C. Guedes, D. J. dos Santos, *Eur. J. Med. Chem.* 80, (2014), 523-534. <https://doi.org/10.1016/j.ejmech.2014.04.076>; f) R. Haudecoeur, A. Ahmed-Belkacem, W. Yi, A. Fortuné, R. Brillet, C. Belle, E. Nicolle, C. Pallier, J.-M. Pawlotsky, A. Boumendjel, *J. Med. Chem.* 54, (2011), 5395-5402. <https://doi.org/10.1021/jm200242p>; g) S. Y. Shin, M. C. Shin, J.-S. Shin, K.-T. Lee, Y. S. Lee, *Bioorg. Med. Chem. Lett.* 21, (2011), 4520-4523. <https://doi.org/10.1016/j.bmcl.2011.08.020>; h) M. Irshad, Q. Ali, F. Iram, S.A. Ahamad, M. Saleem, M. Saadia, M. Batool, A. Kanwal, S. Tabassum. *Russ. J. Gen. Chem.* 89, (2019), 1519-27. <https://doi.org/10.1134/S1070363219080310>; i) Y. Li, X. Qiang, L. Luo, Y. Li, G. Xiao, Z. Tan, Y. Deng, *Bioorg. Med. Chem.* 24, (2016), 2342-2351. <https://doi.org/10.1016/j.bmc.2016.04.012>; j) R. Haudecoeur, M. Carotti, A. Gouron, M. Maresca, E. Buitrago, R. Hardré, E. Bergantino, H. I. n. Jamet, C. Belle, M. Réglier, *ACS Med. Chem. Lett.* 8, (2017), 55-60. <https://dx.doi.org/10.1021/acsmmedchemlett.6b00369>.
- [21] N. A. Razak, N. Abu, W. Y. Ho, N. R. Zamberi, S. W. Tan, N. B. Alitheen, K. Long, S. K. Yeap, *Sci. Rep.* 9, (2019), 1514. <https://doi.org/10.1038/s41598-018-37796-w>.
- [22] a) S. Kumar, *Green Chem. Lett. Rev.* 7, (2014), 95-99. <https://doi.org/10.1080/17518253.2014.895867>; b) G. Kumar, E. Lathwal, B. Saroha, S. Kumar, S. Kumar, N. S. Chauhan, T. Kumar, *ChemistrySelect* 5, (2020), 3539-3543. <https://doi.org/10.1002/slct.201904912>.
- [23] S. Okombi, D. Rival, S. Bonnet, A.-M. Mariotte, E. Perrier, A. Boumendjel, *J. Med. Chem.* 49, (2006), 329-333. <https://doi.org/10.1021/jm050715i>.

- [24] R. C. Kamboj, G. Sharma, P. Jindal, R. Arora, D. Kumar, S. Kumar, P. Kumar, *Arabian J. Chem.* 10, (2017), S3190-S3196. <https://doi.org/10.1016/j.arabjc.2013.12.013>.
- [25] B. Karale, P. Nirmal, H. Akolkar, *Indian J. Chem.* 54B, (2015), 399-405
- [26] X. Zheng, H. Wang, Y. M. Liu, X. Yao, M. Tong, Y. H. Wang, D. F. Liao, *J. Heterocycl. Chem.* 52, (2015), 296-301. <https://doi.org/10.1002/jhet.1969>.
- [27] E. Rudyakova, V. Savosik, L. Papernaya, A. Albanov, I. Evstaf'eva, G. Levkovskaya, *Russ. J. Org. Chem.* 45, (2009), 1040-1044. <https://doi.org/10.1134/S1070428009070100>.
- [28] S. Mahata, A. C. Bharti, S. Shukla, A. Tyagi, S. A. Husain, B. C. Das, *Mol. Cancer* 10, (2011), 39. <https://doi.org/10.1186/1476-4598-10-39>.
- [29] H.H. Jardosh, M.P. Patel, *Arabian J. Chem*, 10, (2017), S3781-S3791. <https://doi.org/10.1016/j.arabjc.2014.05.014>.

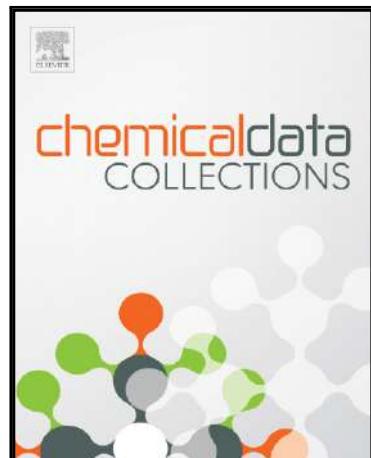
Research Highlights

- Benzofuran-3(2H)-one and pyrazole linked fourteen novel aurones were designed and synthesized under solvent free sustainable conditions.
- Structures of the synthesized aurones were confirmed by their IR, ¹H-NMR, ¹³C-NMR, elemental analysis and Mass spectrometry data.
- Cytotoxic activity of these novel hydroxy aurones derivatives was evaluated toward MCF-7 cancer cell line.
- **3e, 3c, 3i, 3a, and 3n** displayed outstanding potency against MCF-7 (**IC₅₀**: 2.7–15.5 $\mu\text{g/mL}$) in comparison to standard (paclitaxel, $\text{IC}_{50} = \mathbf{18.5 \mu\text{g/mL}}$)

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References

- [1] a) M. Murias, W. Jäger, N. Handler, T. Erker, Z. Horvath, T. Szekeres, H. Nohl, L. Gille, *Biochem. Pharmacol.* 69, (2005), 903-912. <https://doi.org/10.1016/j.bcp.2004.12.001>; b) J. Ferlay, I. Soerjomataram, R. Dikshit, S. Eser, C. Mathers, M. Rebelo, D. M. Parkin, D. Forman, F. Bray, *Int. J. Cancer* 136, (2015), E359-E386. <https://doi.org/10.1002/ijc.29210>.
- [2] R.L. Siegel, K.D. Miller, A. Jemal, *Cancer statistics* 69, (2019), 7-34. <https://doi.org/10.3322/caac.21551>.
- [3] J. Liu, B. Ming, G.-H. Gong, D. Wang, G.-L. Bao, L.-J. Yu, *RSC Adv.* 8, (2018), 4386-4416. <https://doi.org/10.1039/C7RA12912B>.
- [4] A. Saeed, F. A. Larik, P. A. Channar, *Res. Chem. Intermed.* 42, (2016), 6805-6813. <https://doi.org/10.1007/s11164-016-2527-6>.
- [5] M. Leulescu, A. Rotaru, I. Pălărie, A. Moanță, N. Cioateră, M. Popescu, E. Morîntale, M. V. Bubulică, G. Florian, A. Hărăbor, *J. Therm. Anal. Calorim.* 134, (2018), 209-231. <https://doi.org/10.1007/s10973-018-7663-3>.
- [6] S. D. Sawant, G. L. Reddy, M. I. Dar, M. Srinivas, G. Gupta, P. K. Sahu, P. Mahajan, A. Nargotra, S. Singh, S. C. Sharma, *Bioorg. Med. Chem.* 23, (2015), 2121-2128. <https://doi.org/10.1016/j.bmc.2015.03.005>.
- [7] M.A.K.F. Tatsuo, W.M. Carvalho, C.V. Silva, *Inflammation* 18, (1994), 399–405 <https://doi.org/10.1007/BF01534437>
- [8] A. Palomer, F. Cabré, J. Pascual, J. Campos, M. A. Trujillo, A. Entrena, M. A. Gallo, L. García, D. Mauleón, A. Espinosa, *J. Med. Chem.* 45, (2002), 1402-1411. <https://doi.org/10.1021/jm010458r>.
- [9] E. K. Abdelall, G. M. Kamel, *Eur. J. Med. Chem.* 118, (2016), 250-258. <https://doi.org/10.1016/j.ejmech.2016.04.049>.
- [10] a) X. Dai, G. Guo, P. Zou, R. Cui, W. Chen, X. Chen, C. Yin, W. He, R. Vinothkumar, F. Yang, *J. Exp. Clin. Cancer Res.* 36, (2017), 120. <https://doi.org/10.1186/s13046-017-0584-3>; b) M. J. Carr, J. Sun, Z. Eroglu, J. S. Zager, *Expert Opin. Pharmacother.*, 21, (2020), 155-161. <https://doi.org/10.1080/14656566.2019.1694664>; c) M. S. Abdel-Maksoud, M. I. El-Gamal, M. M. G. El-Din, C. H. Oh, *J. Enzyme Inhib. Med. Chem.*, 34,(2019), 97-109. <https://doi.org/10.1080/14756366.2018.1530225>.
- [11] Ş. G. Küçükgüzel, S. Şenkardes, *Eur. J. Med. Chem.* 97, (2015), 786-815. <https://doi.org/10.1016/j.ejmech.2014.11.059>.
- [12] Y. Estevez, M. Quiliano, A. Burguete, B. Cabanillas, M. Zimic, E. Málaga, M. Verástegui, S. Pérez-Silanes, I. Aldana, A. Monge, *Exp. Parasitol.* 127, (2011), 745-751. <https://doi.org/10.1016/j.exppara.2011.01.009>.
- [13] S. Fustero, R. Román, J. F. Sanz-Cervera, A. Simón-Fuentes, J. Bueno, S. Villanova, *J. Org. Chem.* 73, (2008), 8545-8552. <https://doi.org/10.1021/jo801729p>.
- [14] G. M. Nitulescu, L. Matei, I. M. Aldea, C. Draghici, O. T. Olaru, C. Bleotu, *Arabian J. Chem.* 12, (2019), 816-824. <https://doi.org/10.1016/j.arabjc.2015.12.006>.

- [15] F. K. Keter, J. Darkwa, *BioMetals* 25, (2012), 9-21. <https://doi.org/10.1007/s10534-011-9496-4>.
- [16] a) S. M. Gomha, M. A. Abdallah, I. M. Abbas, M. S.H. Kazem, *Med. Chem.* 14, (2018), 344-355, <https://doi.org/10.2174/1573406413666171020114105>; b) S. M. Gomha, M. M. Edrees, R.A.M. Faty *Chem. Cent. J.* 37, (2017), 37, <https://doi.org/10.1186/s13065-017-0266-4>. c) M. A. Abdallah, S. M. Gomha, I. M. Abbas, , M. S. H. Kazem, S. S. Alterary, Y. N. Mabkhot, *Appl. Sci.* 7, (2017), 785, <https://doi.org/10.3390/app7080785>. d) R. Kenchappa, Y.D. Bodke, *Chem. Data Collect.* 28, (2020), 100453. <https://doi.org/10.1016/j.cdc.2020.100453>.
- [17] R. Haudecoeur, A. Boumendjel, *Curr. Med. Chem.* 19, (2012), 2861-2875. <https://doi.org/10.2174/092986712800672085>.
- [18] A.V. Popova, S.P. Bondarenko, M.S. Frasinyuk, *Chem Heterocycl Comp.* 55, (2019), 285–299. <https://doi.org/10.1007/s10593-019-02457-x>.
- [19] C.-Y. Lee, E.-H. Chew, M.-L. Go, *Eur. J. Med. Chem.* 45, (2010), 2957-2971. <https://doi.org/10.1016/j.ejmech.2010.03.023>.
- [20] a) G. S. Hassan, H. H. Georgey, R. F. George, E. R. Mohammed, *Future Med. Chem.* 10, (2018), 27-52. <https://doi.org/10.4155/fmc-2017-0147>; b) A. Detsi, M. Majdalani, C. A. Kontogiorgis, D. Hadjipavlou-Litina, P. Kefalas, *Bioorg. Med. Chem.* 17, (2009), 8073-8085. <https://doi.org/10.1016/j.bmc.2009.10.002>; c) W. Huang, M.-Z. Liu, Y. Li, Y. Tan, G.-F. Yang, *Bioorg. Med. Chem.* 15, (2007), 5191-5197. <https://doi.org/10.1016/j.bmc.2007.05.022>; d) B. P. Bandgar, S. A. Patil, B. L. Korbad, S. C. Biradar, S. N. Nile, C. N. Khobragade, *Eur. J. Med. Chem.* 45, (2010), 3223-3227. <https://doi.org/10.1016/j.ejmech.2010.03.045>; e) M. P. Carrasco, A. S. Newton, L. Gonçalves, A. Góis, M. Machado, J. Gut, F. Nogueira, T. Hänscheid, R. C. Guedes, D. J. dos Santos, *Eur. J. Med. Chem.* 80, (2014), 523-534. <https://doi.org/10.1016/j.ejmech.2014.04.076>; f) R. Haudecoeur, A. Ahmed-Belkacem, W. Yi, A. Fortuné, R. Brillet, C. Belle, E. Nicolle, C. Pallier, J.-M. Pawlotsky, A. Boumendjel, *J. Med. Chem.* 54, (2011), 5395-5402. <https://doi.org/10.1021/jm200242p>; g) S. Y. Shin, M. C. Shin, J.-S. Shin, K.-T. Lee, Y. S. Lee, *Bioorg. Med. Chem. Lett.* 21, (2011), 4520-4523. <https://doi.org/10.1016/j.bmcl.2011.08.020>; h) M. Irshad, Q. Ali, F. Iram, S.A. Ahamad, M. Saleem, M. Saadia, M. Batool, A. Kanwal, S. Tabassum. *Russ. J. Gen. Chem.* 89, (2019), 1519-27. <https://doi.org/10.1134/S1070363219080310>; i) Y. Li, X. Qiang, L. Luo, Y. Li, G. Xiao, Z. Tan, Y. Deng, *Bioorg. Med. Chem.* 24, (2016), 2342-2351. <https://doi.org/10.1016/j.bmc.2016.04.012>; j) R. Haudecoeur, M. Carotti, A. Gouron, M. Maresca, E. Buitrago, R. Hardré, E. Bergantino, H. I. n. Jamet, C. Belle, M. Réglier, *ACS Med. Chem. Lett.* 8, (2017), 55-60. <https://dx.doi.org/10.1021/acsmmedchemlett.6b00369>.
- [21] N. A. Razak, N. Abu, W. Y. Ho, N. R. Zamberi, S. W. Tan, N. B. Alitheen, K. Long, S. K. Yeap, *Sci. Rep.* 9, (2019), 1514. <https://doi.org/10.1038/s41598-018-37796-w>.
- [22] a) S. Kumar, *Green Chem. Lett. Rev.* 7, (2014), 95-99. <https://doi.org/10.1080/17518253.2014.895867>; b) G. Kumar, E. Lathwal, B. Saroha, S. Kumar, S. Kumar, N. S. Chauhan, T. Kumar, *ChemistrySelect* 5, (2020), 3539-3543. <https://doi.org/10.1002/slct.201904912>.
- [23] S. Okombi, D. Rival, S. Bonnet, A.-M. Mariotte, E. Perrier, A. Boumendjel, *J. Med. Chem.* 49, (2006), 329-333. <https://doi.org/10.1021/jm050715i>.

- [24] R. C. Kamboj, G. Sharma, P. Jindal, R. Arora, D. Kumar, S. Kumar, P. Kumar, *Arabian J. Chem.* 10, (2017), S3190-S3196. <https://doi.org/10.1016/j.arabjc.2013.12.013>.
- [25] B. Karale, P. Nirmal, H. Akolkar, *Indian J. Chem.* 54B, (2015), 399-405
- [26] X. Zheng, H. Wang, Y. M. Liu, X. Yao, M. Tong, Y. H. Wang, D. F. Liao, *J. Heterocycl. Chem.* 52, (2015), 296-301. <https://doi.org/10.1002/jhet.1969>.
- [27] E. Rudyakova, V. Savosik, L. Papernaya, A. Albanov, I. Evstaf'eva, G. Levkovskaya, *Russ. J. Org. Chem.* 45, (2009), 1040-1044. <https://doi.org/10.1134/S1070428009070100>.
- [28] S. Mahata, A. C. Bharti, S. Shukla, A. Tyagi, S. A. Husain, B. C. Das, *Mol. Cancer* 10, (2011), 39. <https://doi.org/10.1186/1476-4598-10-39>.
- [29] H.H. Jardosh, M.P. Patel, *Arabian J. Chem*, 10, (2017), S3781-S3791. <https://doi.org/10.1016/j.arabjc.2014.05.014>.

Research Highlights

- Benzofuran-3(2H)-one and pyrazole linked fourteen novel aurones were designed and synthesized under solvent free sustainable conditions.
- Structures of the synthesized aurones were confirmed by their IR, ¹H-NMR, ¹³C-NMR, elemental analysis and Mass spectrometry data.
- Cytotoxic activity of these novel hydroxy aurones derivatives was evaluated toward MCF-7 cancer cell line.
- **3e, 3c, 3i, 3a, and 3n** displayed outstanding potency against MCF-7 (**IC₅₀**: 2.7–15.5 $\mu\text{g/mL}$) in comparison to standard (paclitaxel, $\text{IC}_{50} = \mathbf{18.5 \mu\text{g/mL}}$)